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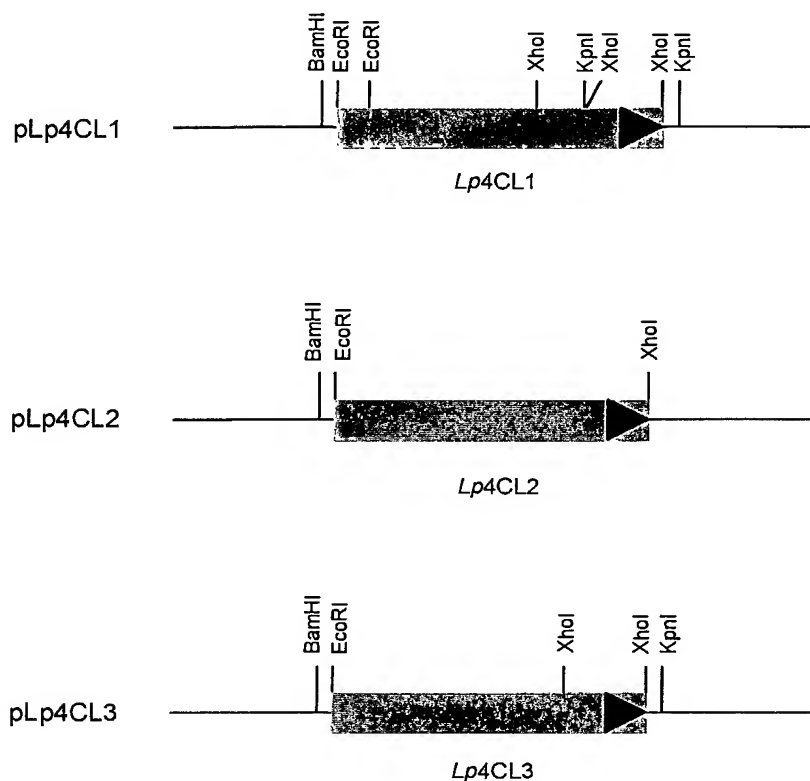
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(54) Title: MODIFICATION OF LIGNIN BIOSYNTHESIS



(57) Abstract: The present invention relates to the modification of lignin biosynthesis in plants, using the nucleotide sequences encoding the enzymes 4-coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR), cinnamyl alcohol dehydrogenase (CAD) of the lignin biosynthetic pathway, from ryegrass (*Lolium*) and fescue (*Festuca*). The present invention also relates to regulatory elements, promoters capable of causing expression of exogenous genes in plants, wherein the regulatory elements are from the genes for caffeic acid O-methyl transferase (OMT), 4CL, CCR or CAD. The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, plant seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors and methods using the nucleic acids, regulatory elements and vectors.



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MODIFICATION OF LIGNIN BIOSYNTHESIS

The present invention relates to the modification of lignin biosynthesis in plants and, more particularly, to enzymes involved in the lignin biosynthetic pathway and nucleic acids encoding such enzymes.

- 5 The present invention also relates to a regulatory element and, more particularly, to a promoter capable of causing expression of an exogenous gene in plant cells, such as a gene encoding an enzyme involved in the lignin biosynthetic pathway in plants.

- 10 The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors, and methods of using the nucleic acids, regulatory elements and vectors.

- 15 Lignins are complex phenolic polymers that strengthen plant cell walls against mechanical and chemical degradation. The process of lignification typically occurs during secondary thickening of the walls of cells with structural, conductive or defensive roles. Three monolignol precursors, sinapyl, coniferyl and *p*-coumaryl alcohol combine by dehydrogenative polymerisation to produce respectively the syringyl(S), guaiacyl(G) and hydroxyl(H) subunits of the lignin polymer, which can also become linked to cell-wall polysaccharides through the action of peroxidases and other oxidative isozymes. In grasses, biosynthesis of the monolignol precursors is a multistep process beginning with the aromatic amino-acids phenylalanine and tyrosine. It is the final two reduction/ dehydrogenation steps of the pathway, catalysed by Cinnamoyl CoA Reductase (CCR) and Cinnamyl Alcohol Dehydrogenase (CAD) that are
20
25 considered to be specific to lignin biosynthesis. The proportions of monolignols incorporated into the lignin polymer vary depending on plant species, tissue, developmental stage and sub-cellular location.

Caffeic acid *O*-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase

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(CAD) are key enzymes involved in lignin biosynthesis.

Worldwide permanent pasture is estimated to cover 70% of agriculturally cultivated area. Ryegrasses (*Lolium* spp.) together with the closely related fescues (*Festuca* spp.) are of significant value in temperate grasslands. The commercially most important ryegrasses are Italian or annual ryegrass (*L. multiflorum* Lam.) and perennial ryegrass (*L. perenne* L.). They are the key forage species in countries where livestock production is an intensive enterprise, such as the Netherlands, United Kingdom and New Zealand. The commercially most important fescues are tall fescue (*F. anundinacea* Schreb.), meadow fescue (*F. pratensis*) and red fescue (*F. rubra*).

Perennial ryegrass (*Lolium perenne* L.) is the major grass species sown in temperate dairy pastures in Australia, and the key pasture grass in temperate climates throughout the world. A marked decline of the feeding value of grasses is observed in temperate pastures of Australia during late spring and early summer, where the nutritive value of perennial ryegrass based pasture is often insufficient to meet the metabolic demands of lactating dairy cattle. Perennial ryegrass is also an important turf grass.

Grass and legume *in vitro* dry matter digestibility has been negatively correlated with lignin content. In addition, natural mutants of lignin biosynthetic enzymes in maize, sorghum and pearl millet that have higher rumen digestibility have been characterised as having lower lignin content and altered S/G subunit ratio. Thus, lignification of plant cell walls is the major factor identified as responsible for lowering digestibility of forage tissues as they mature.

It would be desirable to have methods of altering lignin biosynthesis in plants, including grass species such as ryegrasses and fescues, by reducing the activity of key biosynthetic enzymes in order to reduce lignin content and/or alter lignin composition for enhancing dry matter digestibility and improving herbage quality. However, for some applications it may be desirable to

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enhance lignin biosynthesis to increase lignin content and/or alter lignin composition, for example to increase mechanical strength of wood, to increase mechanical strength of turf grasses, to reduce plant height and reduce lodging or improve disease resistance.

5 While nucleic acid sequences encoding some of the enzymes involved in the lignin biosynthetic pathway have been isolated for certain species of plants, there remains a need for materials useful in the modification of lignin biosynthesis in plants, particularly grass species such as ryegrasses and fescues.

10 Other phenotypic traits which may be improved by transgenic manipulation of plants include disease resistance, mineral content, nutrient quality and drought tolerance.

15 However, transgenic manipulation of phenotypic traits in plants requires the availability of regulatory elements capable of causing the expression of exogenous genes in plant cells.

It is an object of the present invention to overcome, or at least alleviate, one or more of the difficulties or deficiencies associated with the prior art.

20 In one aspect, the present invention provides substantially purified or isolated nucleic acids or nucleic acid fragments encoding the following enzymes from a ryegrass (*Lolium*) or fescue (*Festuca*) species: 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

25 The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall fescue, meadow fescue and red fescue. Preferably the ryegrass or fescue species is a ryegrass, more preferably perennial ryegrass (*Lolium perenne*).

The nucleic acid or nucleic acid fragment may be of any suitable type

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and includes DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

5 The term "isolated" means that the material is removed from its original environment (eg. the natural environment if it is naturally occurring). For example, a naturally occurring nucleic acid present in a living plant is not isolated, but the same nucleic acid separated from some or all of the coexisting materials in the natural system, is isolated. Such nucleic acids could be part of a vector and/or such nucleic acids could be part of a
10 composition, and still be isolated in that such a vector or composition is not part of its natural environment.

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding 4CL includes a nucleotide sequence selected from the group consisting of (a)
15 sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

20 In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding CCR includes a nucleotide sequence selected from the group consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7);
25 (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding
30 CAD includes a nucleotide sequence selected from the group consisting of (a)

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the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b);
5 and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

By "functionally active" is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of modifying lignin biosynthesis in a plant. Such variants include naturally occurring allelic variants and non-
10 naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above mentioned
15 sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Such functionally active variants and fragments include, for example, those having nucleic acid changes which result in conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment
20 has a size of at least 10 nucleotides, more preferably at least 15 nucleotides, most preferably at least 20 nucleotides.

In a second aspect of the present invention there is provided a vector including a nucleic acid or nucleic acid fragment according to the present invention.

25 In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element such as a promoter, a nucleic acid or nucleic acid fragment according to the present invention and a terminator; said regulatory element, nucleic acid or nucleic acid fragment and terminator being operatively linked.

30 By "operatively linked" is meant that said regulatory element is capable

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of causing expression of said nucleic acid or nucleic acid fragment in a plant cell and said terminator is capable of terminating expression of said nucleic acid or nucleic acid fragment in a plant cell. Preferably, said regulatory element is upstream of said nucleic acid or nucleic acid fragment and said terminator is downstream of said nucleic acid or nucleic acid fragment.

The vector may be of any suitable type and may be viral or non-viral. The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable or integrative or viable in the plant cell.

The regulatory element and terminator may be of any suitable type and may be endogenous to the target plant cell or may be exogenous, provided that they are functional in the target plant cell.

Preferably the regulatory element is a promoter. A variety of promoters which may be employed in the vectors of the present invention are well known to those skilled in the art. Factors influencing the choice of promoter include the desired tissue specificity of the vector, and whether constitutive or inducible expression is desired and the nature of the plant cell to be transformed (eg. monocotyledon or dicotyledon). Particularly suitable promoters include the Cauliflower Mosaic Virus 35S (CaMV 35S) promoter, the maize Ubiquitin promoter, the rice Actin promoter, and ryegrass endogenous OMT, 4CL, CCR or CAD promoters.

A variety of terminators which may be employed in the vectors of the present invention are also well known to those skilled in the art. The terminator may be from the same gene as the promoter sequence or a different gene. Particularly suitable terminators are polyadenylation signals, such as the

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CaMV 35S polyA and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the nucleic acid or nucleic acid fragment of the present invention and the terminator, may include further elements necessary for expression of the nucleic acid or nucleic acid fragment, in different combinations, for example vector backbone, origin of replication (*ori*), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other selectable marker genes [such as the neomycin phosphotransferase (*npt2*) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *pat*) gene], and reporter genes (such as beta-glucuronidase (GUS) gene (*gusA*)]. The vector may also contain a ribosome binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said nucleic acid or nucleic acid fragment. Techniques for operatively linking the components of the vector of the present invention are well known to those skilled in the art. Such techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction enzyme sites.

The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons (such as grasses from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, corn, oat, sugarcane, wheat and barley), dicotyledons (such as arabidopsis,

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tobacco, legumes, alfalfa, oak, eucalyptus, maple, canola, soybean and chickpea) and gymnosperms. In a preferred embodiment, the vectors are used to transform monocotyledons, preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably
5 perennial ryegrass (*Lolium perenne*) including forage and turf type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, protoplast fusion, injection into reproductive organs, injection into immature
10 embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce
15 successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell, plant, plant seed or other plant part, including, eg transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part may be from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably a ryegrass, most preferably perennial ryegrass, including forage- and turf-type
25 cultivars.

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The present invention also provides a plant, plant seed or other plant part derived from a plant cell of the present invention.

The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

- 5 In a further aspect of the present invention there is provided a method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment and/or a vector according to the present invention.

- 10 By "an effective amount" is meant an amount sufficient to result in an identifiable phenotypic trait in said plant, or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant, the route of administration and other relevant factors. Such a person will readily be able to determine a suitable amount and method of administration. See, for example,
15 Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

- Using the methods and materials of the present invention, plant lignin biosynthesis may be increased, decreased or otherwise modified relative to an
20 untransformed control plant. It may be increased or otherwise modified, for example, by incorporating additional copies of a sense nucleic acid or nucleic acid fragment of the present invention. It may be decreased, for example, by incorporating an antisense nucleic acid or nucleic acid fragment of the present invention. In addition, the number of copies of genes encoding for different
25 enzymes in the lignin biosynthetic pathway may be manipulated to modify the relative amount of each monolignol synthesized, thereby leading to the formation of lignin having altered composition.

In a still further aspect of the present invention there is provided use of a nucleic acid or nucleic acid fragment according to the present invention, and/or

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nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof, as a molecular genetic marker.

More particularly, nucleic acids or nucleic acid fragments according to the present invention, and/or nucleotide sequence information thereof, and/or
5 single nucleotide polymorphisms thereof, may be used as a molecular genetic marker for qualitative trait loci (QTL) tagging, mapping, DNA fingerprinting and in marker assisted selection, and may be used as candidate genes or perfect markers, particularly in ryegrasses and fescues. Even more particularly, nucleic acids or nucleic acid fragments according to the present invention,
10 and/or nucleotide sequence information thereof, may be used as molecular genetic markers in forage and turf grass improvement, eg. tagging QTLs for dry matter digestibility, herbage quality, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

In a still further aspect of the present invention there is provided a
15 substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the enzymes 4CL, CCR and CAD.

The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall
20 fescue, meadow fescue and red fescue. Preferably the species is a ryegrass, more preferably perennial ryegrass *L. perenne*).

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme 4CL includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4
25 hereto (Sequence ID Nos: 2, 4 and 6, respectively); and functionally active fragments and variants thereof.

In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CCR includes an amino acid sequence selected from the group consisting of the sequence shown in Figure

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10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

5 In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CAD includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

10 By "functionally active" in this context is meant that the fragment or variant has one or more of the biological properties of the enzymes 4CL, CCR and CAD, respectively. Additions, deletions, substitutions and derivatizations of one or more of the amino acids are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the fragment or variant has at least approximately 60% identity to the relevant part of the above mentioned sequence, more preferably
15 at least approximately 80% identity, most preferably at least approximately 90% identity. Such functionally active variants and fragments include, for example, those having conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment has a size of at least 10 amino acids, more preferably at least 15 amino acids,
20 most preferably at least 20 amino acids.

In a further embodiment of this aspect of the invention, there is provided a polypeptide recombinantly produced from a nucleic acid or nucleic acid fragment according to the present invention. Techniques for recombinantly producing polypeptides are well known to those skilled in the art.

25 In a still further aspect of the present invention there is provided a lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part of the present invention.

Such lignins may be modified from naturally occurring lignins in terms of the length, the degree of polymerisation (number of units), degree of branching

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and/or nature of linkages between units.

In a still further aspect, the present invention provides an isolated regulatory element capable of causing expression of an exogenous gene in plant cells. Preferably the regulatory element is isolated from a nucleic acid or
5 nucleic acid fragment encoding OMT, 4CL, CCR or CAD.

The regulatory element may be a nucleic acid molecule, including DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

10 Preferably the regulatory element includes a promoter, more preferably an *O*-methyltransferase promoter, even more preferably an *O*-methyltransferase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

15 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the caffeic acid *O*-methyltransferase gene corresponding to the cDNA homologue *LpOMT1* from perennial ryegrass.

20 Preferably the regulatory element includes a nucleotide sequence including the first approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing
25 expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional

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activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a
5 size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

- 10 Nucleotides – 4581 to –1
- Nucleotides –4285 to –1
- Nucleotides –4020 to –1
- Nucleotides –2754 to –1
- Nucleotides – 1810 to –1
- 15 Nucleotides –831 to –1
- Nucleotides –560 to –1
- Nucleotides –525 to –1
- Nucleotides –274 to –1
- Nucleotides –21 to –1

20 of Figure 18 hereto (Sequence ID No: 13);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a 4 coumarate-CoA ligase promoter, even more preferably a 4 coumarate-CoA ligase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more
25 preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

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In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the 4 coumarate-CoA ligase gene corresponding to the cDNA homologue *Lp4CL2* from perennial ryegrass.

5 Preferably the regulatory element includes a nucleotide sequence including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto (Sequence ID No: 17); or a functionally active fragment or variant thereof.

10 By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or
15 variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

20 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

Nucleotides – 2206 to –1

Nucleotides -1546 to –1

25 Nucleotides –1186 to –1

Nucleotides –406 to –1

Nucleotides – 166 to –1

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of Figure 38 hereto (Sequence ID No: 17);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a cinnamoyl-CoA reductase promoter, even more preferably a cinnamoyl-CoA reductase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the cinnamoyl-CoA reductase gene corresponding to the *LpCCR1* cDNA from perennial ryegrass.

Preferably the regulatory element includes a nucleotide sequence including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

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In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

- Nucleotides – 6735 to –1
 - 5 Nucleotides –5955 to –1
 - Nucleotides –5415 to –1
 - Nucleotides –4455 to –1
 - Nucleotides – 4035 to –1
 - Nucleotides –3195 to –1
 - 10 Nucleotides –2595 to –1
 - Nucleotides –1755 to –1
 - Nucleotides –1275 to –1
 - Nucleotides –495 to –1
 - Nucleotides –255 to –1
 - 15 Nucleotides –75 to –1
- of Figure 39 hereto (Sequence ID No: 18);

or a functionally active fragment or variant thereof.

By an "exogenous gene" is meant a gene not natively linked to said regulatory element. In certain embodiments of the present invention the
20 exogenous gene is also not natively found in the relevant plant or plant cell.

The exogenous gene may be of any suitable type. The exogenous gene may be a nucleic acid such as DNA (e.g. cDNA or genomic DNA) or RNA (e.g. mRNA), and combinations thereof. The exogenous gene may correspond to a target gene, for example a gene capable of influencing disease resistance,
25 herbage digestibility, nutrient quality, mineral content or drought tolerance or be a fragment or variant (such as an analogue, derivative or mutant) thereof

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which is capable of modifying expression of said target gene. Such variants include nucleic acid sequences which are antisense to said target gene or an analogue, derivative, mutant or fragment thereof. The transgene may code for a protein or RNA sequence depending the target condition and whether down
5 or up-regulation of gene expression is required. Preferably, the target gene is selected from exogenous coding sequences coding for mRNA for a protein, this protein may be of bacterial origin (such as enzymes involved in cell wall modification and cell wall metabolism, cytokinin biosynthesis), or eukaryotic origin (such as pharmaceutically active polypeptides) or of plant origin (such as
10 enzymes involved in the synthesis of phenolic compounds, cell wall metabolism, sugar metabolism, lignin biosynthesis). Preferably, the target gene is selected from the group comprising *O*-methyltransferase, 4 coumarate CoA-ligase, cinnamoyl CoA reductase, cinnamyl alcohol dehydrogenase, cinnamate 4 hydroxylase, phenolase, laccase, peroxidase, coniferol glucosyl
15 transferase, coniferin beta-glucosidase, phenylalanine ammonia lyase, ferulate 5-hydroxylase, chitinase, glucanase, isopentenyltransferase, xylanase.

The plant cells, in which the regulatory element of the present invention is capable of causing expression of an exogenous gene, may be of any suitable type. The plant cells may be from monocotyledons (such as grasses
20 from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, corn, grains, oat, sugarcane, wheat and barley), dicotyledons (such as arabidopsis, tobacco, legumes, alfalfa, oak, eucalyptus and maple) and gymnosperms. Preferably the plant cells are from a monocotyledon, more preferably a grass species such as a ryegrass (*Lolium*)
25 or fescue (*Festuca*) species, even more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

The regulatory element according to the present invention may be used to express exogenous genes to which it is operatively linked in the production of transgenic plants.

30 Accordingly, in a further aspect of the present invention there is provided a vector including a regulatory element according to the present

invention.

In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element according to the present invention, an exogenous gene as hereinbefore described, and a terminator; said regulatory
5 element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells. Preferably, said regulatory element is upstream of said exogenous gene and said terminator is downstream of said exogenous gene.

The vector may be of any suitable type and may be viral or non-viral.
10 The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial
15 chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable on integrative or viable in the plant cell.

The terminator may be of any suitable type and includes for example polyadenylation signals, such as the Cauliflower Mosaic Virus 35S polyA
20 (CaMV 35S polyA) and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the exogenous nucleic acid and the terminator, may include further elements necessary for expression of the nucleic acid, in different combinations, for example vector
25 backbone, origin of replication (*ori*), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other selectable marker genes [such as the neomycin phosphotransferase (*npt2*) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *pat*) gene], and reporter
30 genes (such as beta-glucuronidase (GUS) gene (*gusA*)). The vector may also

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contain a ribosome binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

The regulatory element of the present invention may also be used with other full promoters or partial promoter elements.

5 As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot
10 hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said transgene. Techniques for operatively linking the components of the vector of the present invention are well known to those skilled in the art. Such
15 techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction sites.

The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the vectors are used to transform monocotyledons,
20 preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably perennial ryegrass (*Lolium perenne*) including forage- and turf- type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are
25 well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, protoplast fusion, injection into reproductive organs, injection into immature embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely

- 20 -

on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture
5 conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell,
10 plant, plant seed or other plant part, including, eg. transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part is
15 from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably perennial ryegrass (*Lolium perenne*), including forage- and turf-type cultivars.

The present invention also provides a plant, plant seed, or other plant part derived from a plant cell of the present invention.

20 The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

In a still further aspect of the present invention there is provided a recombinant plant genome including a regulatory element according to the present invention.

25 In a preferred embodiment of this aspect of the invention the recombinant plant genome further includes an exogenous gene operatively linked to said regulatory element.

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In a further aspect of the present invention there is provided a method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element and/or a vector according to the present invention.

5 By "an effective amount" is meant an amount sufficient to result in an identifiable phenotypic change in said plant cells or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant cell, the route of administration and other relevant factors. Such a person will readily be able
10 to determine a suitable amount and method of administration. See, for example, Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

The present invention will now be more fully described with reference to
15 the accompanying Examples and drawings. It should be understood, however, that the description following is illustrative only and should not be taken in any way as a restriction on the generality of the invention described above.

In the Figures

Figure 1 shows plasmid maps of the three cDNAs encoding perennial
20 ryegrass 4CL homologues.

Figure 2 shows the nucleotide (Sequence ID No: 1) and amino acid (Sequence ID No: 2) sequences of *Lp4CL1*.

Figure 3 shows the nucleotide (Sequence ID No: 3) and amino acid (Sequence ID No: 4) sequences of *Lp4CL2*.

25 Figure 4 shows the nucleotide (Sequence ID No: 5) and amino acid (Sequence ID No: 6) sequences of *Lp4CL3*.

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Figure 5 shows amino acid sequence alignment of deduced proteins encoded by *Lp4CL1* (Sequence ID No: 2), *Lp4CL2* (Sequence ID No: 4) and *Lp4CL3* (Sequence ID No: 6).

Figure 6 shows northern hybridisation analysis of developing perennial ryegrass using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. SR: roots from seedlings (3-5 d post-germination), SS: shoots from seedlings (3-5 d post-germination), ML: leaves from 12-week-old plants, MS: stems from 12-week-old plants. Blots were washed in 0.2 X SSPE, 0.1 % SDS at 65 °C. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* do not cross hybridise at this stringency. Sizes are given in kb.

Figure 7 shows northern hybridisation analysis showing the time course of expression of 4CL mRNA in wounded perennial ryegrass leaves. Sizes are given in kb.

Figure 8 shows genomic Southern hybridisation analysis using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. 10 µg of digested perennial ryegrass genomic DNA or 20 µg of digested tall fescue genomic DNA were separated on a 1.0 % agarose gel, transferred to Hybond N⁺ membranes and then hybridised with ³²P labelled *Lp4CL1*, *Lp4CL2* or *Lp4CL3* probes. The ryegrass *Lp4CL1*, *Lp4CL2* and *Lp4CL3* genes reveal homologous sequences in tall fescue and indicate that the ryegrass 4CL genes can be used to isolate and to manipulate the expression of the tall fescue (*Festuca arundinacea*) 4CL genes.

Figure 9 shows restriction map of *LpCCR1*. An *L. perenne* seedling cDNA library constructed in Uni-ZAPTM (Stratagene) was screened in a solution containing 10xPIPES, 50% deionised formamide and 10% SDS at 42°C. Filters were washed at room temperature, three times in 0.1% SDS, 2x SSPE and then twice in 0.1% SDS, 0.2x SSPE. The location of the probe used for northern and Southern hybridisation analyses is indicated by the black line labelled *LpCCR531*.

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Figure 10 shows the nucleotide (Sequence ID No: 7) and amino acid (Sequence ID No: 8) sequences of *LpCCR1*.

Figure 11 shows Southern hybridisation analysis of DNA from double haploid (DH) perennial ryegrass using *LpCCR1* as hybridisation probe. 10µg of DH genomic DNA was digested with *DraI*, *BamHI*, *EcoRI*, *EcoRV*, *HindIII* or *XbaI*, separated on a 1% agarose gel and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was probed with the digoxigenin (DIG) labelled *LpCCR531* fragment at 25ng/ml in the hybridisation solution. Hybridisation was in 4x SSC, 50% formamide, 0.1% N-Lauroyl-sarcosine, 0.02% SDS, 2% Blocking solution at 42°C. The membrane was washed twice for five minutes in 2x SSC, 0.1% SDS at room temperature, then twice for fifteen minutes in 0.5x SSC, 0.1%SDS at 68°C. Molecular weight was determined by comparison to a DIG-labelled marker (Roche Molecular Biochemicals).

Figure 12 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using *LpCCR1* probe. Roots from seedlings (3-5 d post-germination), shoots from seedlings (3-5 d post-germination), roots from seedlings (7-10 d post-germination), leaves from seedlings (7-10 d post-germination), roots from 6 and 10 week old plants, leaves from 6 and 10 week old plants, stems from 6 and 10 week old plants, whole seedling from 11 day old *Phalaris* and 7 day old *Festuca*.

Total RNA was isolated using Trizol (GibcoBRL) and 15 µg was separated on a 1.2% Agarose gel containing 6% formamide and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was stained with 0.2% methylene blue/0.3M sodium acetate to visualise the marker and ensure that RNA was evenly loaded. 50 ng *LpCCR531* was random-labelled with ³²P-dCTP (Amersham Megaprime) and hybridisation conditions were 4x SSC, 50% formamide, 0.5% SDS, 5x denhardt solution, 5% dextrane sulphate, 0.1% Herring sperm DNA at 42°C over-night. The ryegrass *LpCCR1*

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gene reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CCR gene can be used to manipulate the expression of the tall fescue (*Festuca arundinacea*) and *Phalaris* CCR endogenous genes.

Figure 13 shows the nucleotide (Sequence ID No: 9) and amino acid
5 (Sequence ID No: 10) sequences of *LpCAD1*.

Figure 14 shows the nucleotide (Sequence ID No: 11) and amino acid
(Sequence ID No: 12) sequences of *LpCAD2*.

Figure 15 shows a plasmid map of a cDNA clone encoding perennial ryegrass CAD homologue *LpCAD1*.

10 Figure 16 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using A) *LpCAD1* and B) *LpCAD2* as hybridisation probes. Roots from seedlings 3-5 d post-germination, 7-10 d post-germination, 6 weeks and 10 weeks, Shoots from seedlings 3-5 d post-germination and 7-10 d post-germination, Leaves
15 from 6 week old and 10 week old plants, stem tissue from 6 and 10 week old plants. RNA isolated from *Phalaris* and *Festuca* 11 and 7 day old seedlings. The ryegrass CAD genes reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CAD gene can be used to manipulate the expression of the tall fescue and *Phalaris* CAD endogenous
20 genes.

Figure 17 shows genomic Southern hybridisation analysis. 10 µg of perennial ryegrass genomic DNA digested with a range of restriction enzymes was separated on a 0.8% agarose gel, transferred to Hybond N and then hybridised with a DIG labelled A) *LpCAD1*, and B) *LpCAD2* hybridisation
25 probe.

Figure 18 shows the nucleotide sequence of the *LpOmt1* promoter (Sequence ID No: 13).

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Figure 19 shows a plasmid map of plant transformation vector carrying the reporter β -glucuronidase (GUS) gene (*gusA*) under control of the perennial ryegrass *LpOmt1* promoter.

Figure 20 (upper image) shows PCR analysis of transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter (upper figure). PCR reactions using *gusA*-specific primers were performed. Figure 20 (lower images) show histochemical GUS assays, demonstrating xylem-specific *gusA* expression (A and B) and *gusA* expression in glandular leaf trichomes (C and D) in transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter.

Figure 21 shows the isolation of the *LpCCR1* genomic clone 1. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *XbaI*, *NcoI*, *SaII*, *XhoI*, *XhoI/SaII* DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CCR1 probe. B) Map showing the genomic gene organisation of *LpCCR1* clone 1 based on sequence results. C) Comparison of plant CCR exon size and number in different plant species (*Lolium perenne*, *Lp.*, *Eucalyptus gunni*, *Eg.*, *Eucalyptus saligna*, *Es.*, *Populus balsamifera*, *Pb.*)

Figure 22 shows the isolation of the *LpCCR1* genomic clone 2. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *XbaI*, *NcoI*, *SaII*, *XhoI*, *XhoI/SaII* DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with 200bp of the CCR1 promoter (Figure 21B). B) Map showing the promoter region of *LpCCR1* clone 2 based on sequence results.

Figure 23 shows the isolation of an *Lp4CL* genomic clone. A) Southern hybridisation analysis of 4CL genomic clone λ Lp4CL2 digested with *BamHI*, *KpnI* or *SaII*. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 hybridisation probe. B) 10 μ l of a standard PCR reaction using forward and reverse oligonucleotides

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designed to positions outlined on C). The PCR products were separated on a 0.8% agarose gel and stained with ethidium bromide. C) Map showing the genomic gene organisation of λ Lp4CL2 based on sequence and PCR results.

Figure 24 shows the isolation of an *Lp4CL* genomic clone. A) Southern hybridisation analysis of 4CL genomic clone λ Lp4CL2 digested with BamHI, KpnI, Sall. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 probe. B) Map showing the genomic gene organisation of *Lp4CL2* clone 1 and the promoter region of clone 2.

Figure 25 shows plasmid map of plant transformation vector carrying the *gusA* gene under control of the perennial ryegrass *Lp4CL2* promoter (*Lp4CL2::gusA*).

Figure 26 shows nucleotide (Sequence ID No: 14) and amino acid (Sequence ID No: 15) sequences of genomic clone CAD2 cv Barlano (Intron 1 and first 111 bp of the coding region are missing).

Figure 27 shows nucleotide (Sequence ID No: 16) and amino acid (Sequence ID No:15) sequences of coding sequence deduced from genomic clone CAD2 cv Barlano (region in bold is missing from the genomic clone).

Figure 28 shows the isolation of *LpCAD2* genomic clone. A) Southern hybridization analysis of CAD genomic clone λ LpCAD2 digested with BamHI, EcoRI, KpnI, Sall or XbaI. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CAD2 hybridisation probe. B) Map showing the genomic gene organisation of λ LpCAD2 based on sequence results.

Figure 29 shows A) Sense and antisense *Lp4CL1*, *Lp4CL2* and *Lp4CL3* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *Lp4CL1*, *Lp4CL2* and *Lp4CL3* transformation vectors under control

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of the maize ubiquitin promoter.

Figure 30 shows A) Sense and antisense *LpCCR1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCCR1* transformation vectors under control of the maize ubiquitin promoter.

- 5 Figure 31 shows A) Sense and antisense *LpCAD1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCAD1* transformation vectors under control of the maize ubiquitin promoter.

- Figure 32 shows molecular analysis of *Lp4CL1*-transgenic tobacco. A) Plasmid map of transformation vector carrying a chimeric sense *Lp4CL1* gene.
10 B) PCR analysis of independent transgenic tobacco clones using *Lp4CL1* specific primers. C) Southern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe. D) Northern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe.

- 15 Figure 33 shows molecular analysis of *LpCCR1*-transgenic tobacco. A) Plasmid map of transformation vectors carrying a chimeric sense and antisense *LpCCR1* gene. B) PCR analysis of independent sense transgenic tobacco clones using *LpCCR1* specific primers.

- Figure 34 shows protocol for suspension culture-independent
20 production of transgenic perennial ryegrass plants. A) Isolated zygotic embryos, plated on MSM5 medium, day 0; B) Embryogenic callus formation and proliferation, 6 - 8 weeks after embryo isolation; C) Embryogenic calli arranged on high osmotic MSM3Plus medium prior to biolistic transformation; D) Histochemical GUS assay showing GUS expressing foci 3 - 4 days post-
25 bombardment of chimeric *gusA* gene; E) Selection of embryogenic calli on MSM3 medium containing 100 mg/l paromomycin (Pm), 2 weeks after microprojectile bombardment; F) Regeneration of Pm resistant shoots on MSK medium containing 100 mg/l Pm, 4 weeks after microprojectile bombardment; G) *In vitro* plant regeneration from PM resistant embryogenic calli, 6 weeks

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after microprojectile bombardment; H) Transgenic perennial ryegrass plants 28 weeks after embryo isolation.

Figure 35 shows molecular analysis of transgenic perennial ryegrass plants carrying sense and antisense *LpOmt1* transgenes. Plasmid maps of vectors used for the co-transformation of perennial ryegrass embryogenic calli; pHP23 carrying a chimeric neomycin phosphotransferase (*npt2*) selectable marker gene; pUbiomt1 carrying a maize ubiquitin promoter driven sense *LpOmt1* gene; pUbitmo1 carrying a maize ubiquitin promoter driven antisense *LpOmt1* gene (top). PCR analysis using *npt2*-specific primers of 5 independent transgenic perennial ryegrass plants from biolistic transformation with sense and antisense *LpOmt1* vectors (upper centre). Southern hybridization analysis with an *omt1* hybridization probe of 7 independent perennial ryegrass plants co-transformed with sense (lanes 1-3) and antisense (lanes 4-7) *LpOmt1* vectors (lower centre left). Southern hybridisation analysis with an *npt2* hybridisation probe of independent perennial ryegrass plants (lower centre right). Northern hybridisation analysis of perennial ryegrass plants co-transformed with antisense *LpOmt1* vector (bottom). C = negative control untransformed perennial ryegrass; P = positive plasmid control.

Figure 36 shows biochemical analysis of *LpOmt1*-transgenic perennial ryegrass. OMT activity of leaf samples from selected independent *LpOmt1*-transgenic perennial ryegrass plants (Ell8, Ell11, Ell14 and Ell15) was determined and compared to untransformed perennial ryegrass negative control plant *L. perenne* cv. Ellett (wild type). Mean values and standard deviations of replicate assays are shown.

Figure 37 shows PCR screening of transgenic ryegrass plants. PCR analysis using *npt2*-specific primers of 8 independent transgenic perennial ryegrass plants from biolistic transformation with antisense *LpUbi4CL2* vector.

Figure 38 shows the nucleotide sequence of genomic clone 4CL2 from perennial ryegrass (Sequence ID No: 17).

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Figure 39 shows the nucleotide sequence of genomic clone CCR1 from perennial ryegrass (Sequence ID No: 18).

Figure 40 shows the map location of *Lp4CL1*, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* (in bold) within the genetic linkage
5 map of perennial ryegrass.

EXAMPLE 1

Isolation and characterisation of three 4-Coumarate CoA-Ligase (4CL) cDNAs from *Lolium perenne*

Materials and Methods

10 *Plant material*

Plants and embryogenic cell suspensions of perennial ryegrass (*Lolium perenne* L.) cv Ellet and tall fescue (*Festuca arundinacea* Schreb.) cv Triumph were established and maintained as previously described (Heath et al., 1998). Wounding experiments were performed with 10-day-old seedlings of perennial
15 ryegrass (cv Ellet) as previously described (Heath et al., 1998).

Screening of a cDNA library

A cDNA library prepared with RNA isolated from perennial ryegrass seedlings (Heath et al., 1998) was screened with a [³²P]dCTP-labelled rice partial 4CL probe. The rice 4CL probe and consisted of a 844 bp 4CL specific
20 sequence inserted into PUC119. This insert has 93 % sequence identity with a rice 4CL cDNA sequence (Genbank, L43362, bases 453-1300). cDNA inserts were excised and recircularized using the ExAssist helper phage with SOLR strain (Stratagene) as described by the manufacturer.

DNA sequencing

25 cDNA clones were digested with 8 restriction enzymes (*Bam*HI, *Eco*RI, *Kpn*I, *Not*I, *Pst*I, *Sal*I, *Xba*I, *Xho*I) and selected clones were sequenced on both

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strands by the dideoxy chain termination method using M13 forward and reverse primers. For sequencing the internal regions of *Lp4CL1*, *Lp4CL2* and *Lp4CL3* synthetic oligonucleotide primers were designed from the DNA sequences previously determined. Sequencing was performed using the ABI dye terminator kit and automatic sequencer. Nucleotide sequences were aligned using the SeqEd program (ABI) and further analysis was performed using the HIBIO DNASIS vs2 program (Hitachi Software Engineering).

Genomic DNA blot analysis

Genomic DNA was isolated from single genotype-derived cell suspensions of perennial ryegrass and tall fescue according to Lichtenstein and Draper (1985). Ten µg of perennial ryegrass DNA and 20 µg of tall fescue DNA was digested with each of the restriction enzymes *HindIII* and *XbaI*, separated on 1 % agarose gels, and transferred to Hybond N⁺ membranes according to the manufacturer's instructions (Amersham). Probes consisted of BamHI/KpnI fragments of *Lp4CL1* (1771 bp), *Lp4CL2* (2034 bp) or *Lp4CL3* (2080 bp) labelled using the Megaprime labelling kit (Amersham) and [³²P]dCTP. Hybridization was performed at 65 °C in 5 X SSPE, 5 X Denhardt's solution, 0.5 % (w/v) SDS, and 200 µg/mL denatured herring sperm DNA. Membranes were washed three times in 2 X SSPE, 0.1 % SDS for 10 min at 25 °C and then twice in 0.1 X SSPE, 0.1 % SDS for 20 min at 65 °C.

RNA blot analysis

Total RNA (10 µg) was separated on 1.2 % formaldehyde gels and transferred to Hybond N (Amersham) membranes according to the manufacturers instructions. Membranes were stained with 0.2 % methylene blue to confirm correct loading and transfer of RNA. Hybridisation was performed at 42 °C in 5 X SSPE, 5 X Denhart's solution, 0.5 % SDS, 50 % deionized formamide, 200 µg/mL denatured herring sperm DNA. Preparation of probes and washing of membranes was as for DNA blot analysis except for the tall fescue Northern blot when the final two washes were performed with 0.1 X SSPE, 0.1 % SDS for 10 min at 42°C.

Results

Isolation and sequence analysis of perennial ryegrass 4CL cDNAs

A cDNA library prepared from RNA extracted from perennial ryegrass seedlings was screened with a rice 4CL hybridization probe and ten cDNAs were isolated from 2×10^5 pfu. The cDNAs were characterised by restriction analysis with 8 restriction enzymes. All clones were full length (approximately 2.0-2.2 kb) with poly(A) tails and could be separated into three groups: *Lp4CL1* (four clones) *Lp4CL2* (five clones) and *Lp4CL3* (one clone). Plasmid maps for *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 1). *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were fully sequenced (Figures 2, 3 and 4, respectively).

Lp4CL1 is 2284 bp long with an open reading frame (ORF) of 1710 bp, a 5' noncoding region of 322 bp and a 3' noncoding region of 252 bp including a poly(A) tail. *Lp4CL2* is 1992 bp long with an ORF of 1668 bp, a 5' noncoding region of 61 bp and a 3' noncoding region of 263 bp including a poly(A) tail. *Lp4CL3* is 2038 bp long with an ORF of 1671 bp, a 5' noncoding region of 112 bp and a 3' noncoding region of 255 bp including a poly(A) tail.

Within the coding region, *Lp4CL1* has 70 % nucleic acid sequence identity with both *Lp4CL2* and *Lp4CL3*, while *Lp4CL2* has 79 % sequence identity with *Lp4CL3*. There is little sequence homology in the 3' noncoding regions between clones (52-55 %).

Amino acid sequence comparisons

The putative proteins encoded by the three cDNAs consist of 570 amino acids [60290 u (Da)] for *Lp4CL1*, 556 amino acids (59238 u) for *Lp4CL2* and 557 amino acids (59735 u) for *Lp4CL3*. The deduced amino acid sequences of *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 5). *Lp4CL2* and *Lp4CL3* share 79 % amino acid sequence identity, *Lp4CL1* and *Lp4CL2* have 61 % amino acid sequence identity, while *Lp4CL1* and *Lp4CL3* have only 58 % amino acid sequence identity. Regions of high sequence homology are more prevalent in the central and c-terminal regions of the enzyme. For example the

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sequence identity between amino acids 208 to 568 of each enzyme is 85 % for *Lp4CL2* and *Lp4CL3*, 72 % for *Lp4CL1* and *Lp4CL2* and 67 % for *Lp4CL1* and *Lp4CL3*.

Lp4CL1, *Lp4CL2* and *Lp4CL3* share several common regions with other
5 plant 4CLs. In particular, they contain the putative AMP-binding domain and
the conserved GEICIRG motif, except for *Lp4CL3* where the second isoleucine
has been replaced with valine (Figure 5). It has been proposed that domain II
is associated with the catalytic activity of 4CL. Also, four Cys residues
conserved in plant 4CLs are conserved in *Lp4CL1*, *Lp4CL2* and *Lp4CL3*
10 (Figure 5). These results suggest that the *L. perenne* cDNAs encode three
divergent 4CL enzymes that are likely to have originated from three different
4CL genes.

Expression of perennial ryegrass 4CL genes

Lp4CL1, *Lp4CL2* and *Lp4CL3* were used as hybridization probes in
15 Northern blots with RNA prepared from different organs of perennial ryegrass
at two developmental stages. All three probes hybridized to a single mRNA
species of approximately 2.2 - 2.3 kb. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were
expressed at both seedling and mature stages of development and in all
organs tested. For *Lp4CL2* and *Lp4CL3* the strongest signal was found in RNA
20 samples from seedling roots and mature stems (Figure 6).

Lp4CL1, *Lp4CL2* and *Lp4CL3* were also used as hybridization probes in
Northern blots with RNA prepared from tall fescue. All three probes hybridized
to a similar mRNA species (2.3 kb) as that in perennial ryegrass (Figure 6).
The strongest signal was found in RNA samples from mature stems with
25 weaker signals in RNA from roots and seedling shoots. No expression of
Lp4CL1, *Lp4CL2* or *Lp4CL3* was observed in leaves. The three probes varied
in their ability to hybridize to the corresponding homologues in tall fescue, with
Lp4CL3 resulting in the highest signal and *Lp4CL1* hybridizing only weakly.

To determine whether 4CL could be induced under stress conditions,

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leaves of perennial ryegrass seedlings were wounded. No increase in the transcript level upon wounding was observed with *Lp4CL1*, *Lp4CL2* or *Lp4CL3* (Figure 7).

Genomic organization of perennial ryegrass 4CL genes

5 Perennial ryegrass DNA was digested with two restriction enzymes, *HindIII* or *XbaI*. Restriction sites for these enzymes are not present in the cDNA sequence of *Lp4CL1*, *Lp4CL2* or *Lp4CL3*. When *Lp4CL1*, *Lp4CL2* or *Lp4CL3* was used as a probe, several DNA hybridizing fragments of varying intensity were revealed (Figure 8). Each probe hybridized to a unique set of
10 fragments, suggesting that *Lp4CL1*, *Lp4CL2* and *Lp4CL3* represent three different genes. Furthermore, *Lp4CL1* and *Lp4CL2* hybridized to 2 to 3 major fragments per digest which may represent either alleles of the same gene or indicate the presence of more than one gene in each class. The *Lp4CL1*, *Lp4CL2* and *Lp4CL3* probes also revealed several different size hybridizing
15 DNA fragments in genomic Southern blots from tall fescue under high stringency conditions (Figure 8), suggesting that three similar 4CL genes are present in *F. arundinacea*.

EXAMPLE 2

Isolation and characterisation of a Cinnamoyl CoA Reductase (CCR)

cDNA from *Lolium perenne*

20 A total of 500,000 phage were screened from a cDNA library constructed from ten-day-old etiolated *L. perenne* seedlings using a maize CCR probe. Ninety-three positive plaques were observed in the primary screen and five were subsequently analysed by restriction enzyme digestion.
25 Four out of the five were identical. One of the four identical cDNAs, *LpCCR1*, was selected for further analysis (Figure 9).

Nucleic acid sequence analysis of perennial ryegrass CCR cDNA

The full nucleotide sequence of *LpCCR1* was obtained and the amino

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acid sequence predicted (Figure 10). *LpCCR1* is a 1395 bp cDNA with 149 bp of 5' non-coding region and 160 bp of 3' non-coding region. An open reading frame of 1086 bp encodes a protein of 362 amino acids. The composition of the coding region was found to be 68% G+C rich. Codon usage was also
5 examined and found to be biased towards *XXC/G* codons (94%), with *XCG* and *XUA* codons accounting for only 9% and 0.55% respectively. G+C richness and bias towards G and C in the third position of a codon triplet are previously reported characteristics of monocot genes.

Genomic organization of perennial ryegrass CCR gene

10 The number of CCR genes present in the ryegrass genome was determined by Southern blot analysis of genomic DNA from double haploid plants, using as probe a fragment of the *LpCCR1* cDNA (*LpCCR531*, Figure 9). Double haploid DNA reduces the complexity associated with allelic variation. Genomic DNA was cut with enzymes that do not cut the cDNA
15 internally; *DraI*, *BamHI*, *EcoRI*, *EcoRV*, *HindIII* and *XbaI*, and the membrane was hybridised and washed under medium-stringency conditions. A single strongly hybridising band was evident in each lane (Figure 11) indicating that there is a single copy of the *LpCCR1* gene in the perennial ryegrass genome.

Expression of perennial ryegrass CCR gene

20 To investigate the expression profile of the CCR gene in ryegrass, northern hybridisation analysis was carried out with total RNA extracted from roots and shoots at seedling growth stages (0.5-1cm and 4-6cm shoots) and roots, stem and leaves at mature growth stages (6 and 10 weeks). Seedlings were grown on filter paper in the dark at 25°C and then transferred to soil and
25 glasshouse conditions (25°C) until the 6 and 10-week stages. Whole seedling total RNA from *Festuca* and *Phalaris* was included in the northern analysis. Hybridisation with *LpCCR531* (Figure 9) was performed at medium-stringency and the membrane was then washed at high-stringency. A transcript of approximately 1.5 kb was detected in all tissues, the level of expression
30 varying with maturity and from one tissue type to another (Figure 12). The *LpCCR1* transcript appears to be more abundant in roots and stem than

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shoots and leaves. In the stem, transcript abundance increases from 6-weeks to 10-weeks; indicating that transcription in stem tissue is up-regulated as the plant matures. Expression was found predominantly in tissues such as stems and roots that are forming secondary cell walls indicating that *LpCCR1* is
5 constitutively involved in lignification.

EXAMPLE 3

Isolation and characterisation of Cinnamyl Alcohol Dehydrogenase (CAD) cDNAs from *Lolium perenne*

A 558 bp cinnamyl alcohol dehydrogenase (CAD) fragment was
10 amplified from cDNA synthesised from total RNA prepared from perennial ryegrass seedlings. The conserved amino acid domains between *Pinus radiata*, *Medicago sativa*, *Aralia cordata*, *Eucalyptus botryoides* and *Arabidopsis thaliana* CADs were used to design oligonucleotides for the amplification of the perennial ryegrass CAD. The forward oligonucleotide was
15 designed to the conserved amino acid domain CAGVTVYS and the reverse oligonucleotide to the conserved domain DVRYRFV. The 551 bp PCR fragment was cloned and sequenced to confirm that it corresponded to a perennial ryegrass CAD PCR fragment. A cDNA library prepared from RNA
20 extracted from perennial ryegrass seedlings was screened with the 551bp PCR fragment specific for perennial ryegrass CAD. Eight cDNAs were isolated and separated into six groups by restriction digest analysis. One representative clone each from two groups (*LpCAD1*, *LpCAD2*) were selected for further characterisation.

Nucleic acid sequence analysis of perennial ryegrass CAD cDNAs

25 The complete sequence of the perennial ryegrass CAD homologue *LpCAD1* was determined (Figure 13). The 1325 bp clone had a poly (A) tail, typical start and stop codons and the open reading frame (ORF) of this clone coded for a putative protein of 408 amino acids.

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The complete nucleotide sequence of the perennial ryegrass CAD homologue *LpCAD2* was also determined (Figure 14).

Expression of perennial ryegrass CAD genes

A northern hybridisation analysis with RNA samples isolated from perennial ryegrass at different developmental stages hybridised with the full length *LpCAD1* 1325 bp cDNA (Figure 15) was performed to determine patterns of organ and developmental expression. The probe hybridised to a single mRNA species of approximately 1.6 kb. The *LpCAD1* transcript was expressed in all tissue tested: roots, shoots, stem and leaves (Figure 16A). The *LpCAD1* transcript was most abundant in root tissue and the mature stem, this expression pattern is typical of a gene involved in the lignification of plant cell walls. Intergeneric homologies were revealed in *Festuca* and *Phalaris*.

A similar northern hybridisation analysis was performed with *LpCAD2* (Figure 16B), however the transcript was found to be most abundant in mature stem tissue and the shoots.

Genomic organization of perennial ryegrass CAD genes

A Southern hybridisation analysis using DNA samples isolated from a perennial ryegrass double haploid plant digested with *DraI*, *BamHI*, *EcoRI*, *EcoRV*, *HindIII* and *XbaI* and hybridised with a 500 bp *LpCAD1* probe was performed. The hybridisation pattern at high stringency revealed the presence of two prominent bands for most digests indicating that *LpCAD1* belongs to a small gene family and exists as a multicopy gene in perennial ryegrass (Figure 17A).

A similar Southern hybridization analysis was performed with *LpCAD2* (Figure 17B) the hybridisation pattern at high stringency revealed the presence of one or two prominent bands for most digests indicating that *LpCAD2* exists as a single copy gene or a member of a small gene family in perennial ryegrass (Figure 17B).

EXAMPLE 4

Isolation and characterisation of genomic clones and promoters for *O*-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) from *Lolium perenne*

Genomic clones and promoters of *O*-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) were isolated from a perennial ryegrass genomic library using the corresponding cDNAs as hybridisation probes.

10 *Isolation and characterisation of genomic clones and promoters for perennial ryegrass O-methyltransferase (OMT)*

A perennial ryegrass genomic library was screened with the cDNA clone, *LpOmt1*, (Heath *et al.* 1998) encoding *O*-methyltransferase (OMT). The sequence of the 5' untranslated region and the coding region was found to be
15 identical to that of the *LpOmt1* cDNA previously isolated. The entire 4.8 kb genomic clone was fully sequenced (Figure 18).

To further characterise the promoters, transcriptional fusions of the promoter sequence to the β -glucuronidase (GUS) coding sequence (*gusA*) have been generated (Figure 19). Direct gene transfer experiments to tobacco
20 protoplasts were performed with the corresponding chimeric genes to transgenically express them in a heterologous system for *in planta* expression pattern analysis by histochemical GUS assays. A set of transgenic tobacco plants carrying a chimeric *gusA* gene under the control of the 5' regulatory region of the *LpOmt1* promoter was generated to assess the potential use of
25 the *LpOmt1* promoter for xylem-specificity and targeted downregulation of genes encoding key lignin biosynthetic enzymes.

The transgenic tobacco plants generated using the *LpOmt1* promoter driven chimeric *gusA* transformation vector were screened by PCR and histochemical GUS assays.

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A PCR screening was undertaken using *gusA* specific primers for the initial identification of transgenic tobacco plants (Figure 20). PCR positive tobacco plants were screened by histochemical GUS assays for *in planta* expression pattern analysis (Figure 20).

5 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamoyl-CoA reductase (CCR)***

A CCR genomic clone from perennial ryegrass was isolated containing 6.5 kb of promoter and the entire gene organisation (intron/exon boundaries). The CCR promoter can be used for targeted expression of foreign genes in
10 transgenic plants.

A perennial ryegrass genomic library was screened with the cDNA clone *LpCCR1* which codes for the lignin biosynthetic enzyme, cinnamyl-CoA reductase (CCR). Four different genomic clones were identified based on restriction digest analysis. Clone 6.1.1a was selected for further analysis. A
15 6.42 kb *XhoI* fragment from clone 6.1.1a, which hybridized strongly to the *LpCCR1* cDNA probe, was subcloned into pBluescriptSK (Figure 21A). Sequence analysis revealed that the 6.42 kb *XhoI* fragment contained the entire *LpCCR1* gene and 200 bp of promoter region. The intron/exon boundaries are illustrated in figure 21B, the location and the size of the exons
20 appear to be conserved in other CCRs from different species (Figure 21C).

To isolate the promoter region of *LpCCR1*, the Southern blot containing digested phage genomic DNA isolated from clone λ Lp6.1.1a was reprobed with the 200bp promoter region. The probe hybridized strongly to a 6.5 kb *SaI* fragment. This genomic fragment *LpCCR1* clone 2, was subcloned into
25 pBluescriptSK and sequenced (Figure 22A). Sequence results revealed that the 6.5 kb *SaI* fragment contained 6.5 kb of promoter (Figure 22B). The full sequence of *LpCCR1* genomic clone containing the promoter and entire gene sequence (exons and introns) was obtained and is shown on Figure 39.

Isolation and characterisation of genomic clones and promoters for perennial ryegrass 4 coumarate CoA-ligase (4CL)

A 4CL2 genomic clone from perennial ryegrass was isolated containing 2.5 kb of promoter and partial gene organisation (intron/exon boundaries). The
5 4CL2 promoter can be used for targeted expression of foreign genes in transgenic plants. The 2.5 kb promoter has been fused to the reporter gene *gusA* for expression analysis.

A perennial ryegrass genomic library was screened with an *Lp4CL* cDNA probe. After tertiary screening positive 4CL genomic clones were
10 obtained and characterised by restriction digest and Southern hybridisation analysis (Figure 23A).

Sequence analysis revealed that the isolated 4CL genomic clone (4CL2) from perennial ryegrass had 100% nucleotide identity to the *Lp4CL2* cDNA clone. To further characterise this 5 kb λ *Lp4CL2* genomic clone and to
15 confirm that it corresponds to the cDNA of *Lp4CL2*, a number of PCR reactions using primers designed to the cDNA were used. PCR results confirmed that the 5 kb genomic fragment was a partial genomic clone corresponding to the *Lp4CL2* cDNA (Figure 23B). Using primer combinations F1 and R1 the entire 4.8kb genomic fragment was amplified. To determine the
20 location of introns additional PCR reactions using the primer combinations F1 / R2 and F2 / R1 were performed, a 1 kb and 3.5 kb bands were amplified respectively. The location and size of the introns could be determined from these results, and further confirmed by sequence analysis. This large 5 kb genomic fragment contains 4 small exons representing the coding sequence of
25 *Lp4CL2* between 508 bp and 1490 bp (Figure 23C).

The genomic clone 1, *Lp4CL2* contained no promoter region. To isolate the promoter region of *Lp4CL2*, the Southern blot containing digested phage genomic DNA isolated from clone λ *Lp4CL2* was reprobbed with a 300 bp EcoRI/BglII isolated from the 5' end of the cDNA clone *Lp4CL2*. The 300 bp
30 probe hybridised strongly to a 2.5 kb BamHI fragment. This genomic fragment

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Lp4CL2 clone 2, was subcloned into pBluescriptSK and sequenced (Figure 24A). Sequence results revealed that the 2.5 kb *Bam*HI fragment contained the 508 bp of the 5' ORF of *Lp4CL2* missing from genomic clone 1 and 2.0 kb of promoter region (Figure 24B). The full sequence of the *Lp4CL2* genomic clone containing the promoter and partial gene sequence (exons and introns) was obtained and is shown on Figure 39.

The promoter from *Lp4CL2* was thus isolated and used for the production of a chimeric *gusA* reporter gene (Figure 25).

Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamyl alcohol dehydrogenase (CAD)

A CAD genomic clone from perennial ryegrass was isolated containing the gene organisation (intron/exon boundaries) minus intron 1 containing the first 111 bp of the CAD coding region. The genomic clone has allowed the identification of a G at position 851 bp in the coding region of the CAD2 genomic clone isolated from perennial ryegrass cv. Barlano which is absent in the CAD2 cDNA clone isolated from perennial ryegrass cv. Ellett. The SNP (single nucleotide polymorphism) found to exist between the 2 cultivars has the potential utility as a molecular marker for herbage quality, dry matter digestibility, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

Results below show the isolation of the genomic clone and sequence analysis of deduced coding sequence from the genomic clone CAD2 from perennial ryegrass cv. Barlano compared to the truncated cDNA CAD2 from the cv Ellett. The missing G in the perennial ryegrass cv. Ellett has been highlighted (Figures 26 and 27).

A perennial ryegrass genomic library was screened with a probe corresponding to the 5' end of the *LpCAD2* cDNA clone, which codes for the lignin biosynthetic enzyme cinnamyl alcohol dehydrogenase. Ten positive plaques were identified and isolated in the primary library screening. After a secondary and tertiary screening, two positive plaques were obtained and

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corresponding positive genomic clones were further characterised by restriction digest and Southern hybridization analyses. Both genomic clones were found to be identical based on restriction digest analyses. One clone, named λ LpCAD2 was chosen for further Southern hybridization analyses. A
5 4.5 kb *Bam*HI fragment which hybridized strongly to the *Lp*CAD2 cDNA probe was subcloned into pBluescriptSK and sequenced (Figure 28A). Sequence analysis revealed that the 4.5 kb *Bam*HI fragment was a partial genomic clone of *Lp*CAD2. This large 4.5 kb genomic fragment contains 4 small exons representing the coding sequence of *Lp*CAD2 between 213 bp and the stop
10 codon at 1213 bp, and the location of the intron/exon boundaries are illustrated in Figure 28B.

EXAMPLE 5

Development of transformation vectors containing chimeric genes with 4CL, CCR and CAD cDNA sequences from perennial ryegrass

15 To alter the expression of the key enzymes involved in lignin biosynthesis 4CL, CCR and CAD, through antisense and/or sense suppression technology and for over-expression of these key enzymes in transgenic plants, a set of sense and antisense transformation vectors was produced. Transformation vectors containing chimeric genes using perennial ryegrass
20 4CL, CCR and CAD cDNAs in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoter were generated (Figures 29, 30 and 31).

EXAMPLE 6

Production and characterisation of transgenic tobacco plants expressing 25 chimeric 4CL, CCR and CAD genes from perennial ryegrass

A set of transgenic tobacco plants carrying chimeric 4CL, CCR and CAD genes from perennial ryegrass were produced and analysed.

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Transformation vectors with *Lp4CL1*, *Lp4CL2* and *Lp4CL3* full length cDNA sequences in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoters were generated. Transformation vectors with *LpCCR1* cDNA in both sense and antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated. Transformation vectors with 1325 bp full length *LpCAD1* cDNA in sense and 1051 bp partial *LpCAD1* cDNA in antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated.

- 10 Direct gene transfer experiments to tobacco protoplasts were performed using these transformation vectors.

The production and molecular analysis of transgenic tobacco plants carrying the perennial ryegrass *Lp4CL1* and *LpCCR1* cDNAs under the control of the constitutive CaMV 35S promoter is described here in detail.

- 15 A set of transgenic tobacco plants generated using the *Lp4CL1* sense transformation vector was screened by PCR and subjected to Southern and northern hybridization analyses.

- 20 A PCR screening was undertaken using *npt2* and *Lp4CL1* specific primers for the initial identification of transgenic tobacco plants. Independent transgenic tobacco plants were identified to be co-transformed with both the selectable marker *npt2* and the *Lp4CL1* chimeric genes (Figure 32).

- 25 Southern hybridisation analysis was performed with DNA samples from PCR positive transgenic tobacco plants to demonstrate the integration of the chimeric *Lp4CL1* transgene in the tobacco plant genome. Independent transgenic tobacco plants carried between 1 and 5 copies of the *Lp4CL1* transgene. No cross-hybridization was observed between the endogenous tobacco 4CL gene and the perennial ryegrass hybridization probe used (Figure

32).

Northern hybridization analysis using total RNA samples prepared from the transgenic tobacco plants carrying the chimeric sense *Lp4CL1* transgene and probed with the *Lp4CL1*-specific hybridization probe revealed the presence of a 1.2 kb *Lp4CL1* transcript strongly expressed in one *Lp4CL1*-transgenic tobacco plant analysed (Figure 32).

The sense and antisense transformation vectors of *LpCCR1* under the control of the CaMV 35S promoter were introduced into tobacco protoplasts via direct gene transfer. A set of transgenic tobacco plants was generated and screened by PCR with specific primers to identify transgenic tobacco plants carrying chimeric *LpCCR1* transgene. The molecular analysis of *LpCCR1*-transgenic tobacco plants is shown (Figure 33).

EXAMPLE 7

Production and characterisation of transgenic perennial ryegrass plants expressing chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass

An improved transformation method was developed for the production of transgenic perennial ryegrass plants by biolistic transformation of embryogenic cells. Transgenic perennial ryegrass plants were generated using chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass and the improved transformation method.

Improved method for the production of transgenic perennial ryegrass plants

This improved procedure utilises embryogenic calli produced from mature seed-derived embryos as direct targets for biolistic transformation without requiring the establishment of embryogenic cell suspensions. The protocol relies on a continuous supply of isolated zygotic embryos for callus induction. Transgenic ryegrass plants can be regenerated 24 – 28 weeks after

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embryo isolation (Fig. 34). Isolated embryos are plated onto MSM5 medium to produce embryogenic calli suitable as targets for biolistic transformation within 8 weeks. The embryogenic calli, treated on high-osmoticum medium MSM3 Plus prior to microprojectile bombardment, are selected on MSM3 medium
5 containing 100 mg/l paromomycin (Pm) for 2 weeks before being transferred onto MSK with 100 mg/l Pm for further 4 weeks until differentiation of Pm resistant shoot appear. Regenerated shoots are transferred on to fresh selective media MSK with 100 mg/l Pm for a further 4 weeks (Figure 34).

***Production of transgenic perennial ryegrass plants expressing chimeric
10 OMT, 4CL, CCR and CAD genes from perennial ryegrass***

Transgenic perennial ryegrass (*Lolium perenne*) plants were generated using chimeric ryegrass OMT, 4CL, CCR and CAD genes by biolistic transformation of embryogenic calli. Examples of the production and detailed molecular analysis of these transgenic ryegrass plants are described.

15 Transgenic perennial ryegrass plants for OMT down-regulation were produced using biolistic transformation of embryogenic calli and plant transformation vectors pUbiomt1 and pUbitmo1 carrying *LpOmt1* cDNA sequence in sense and antisense orientation under control of the constitutive maize ubiquitin promoter. These transgenic perennial ryegrass plants for
20 down-regulated OMT activity were regenerated from paromomycin resistant calli obtained from biolistic transformation using microprojectiles coated with two plasmids; pHP23 (carrying the chimeric *npt2* gene as the selectable marker) and either the sense or antisense *LpOmt1* transformation vector driven by the maize *Ubi* promoter.

25 Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants obtained from biolistic transformation of embryogenic calli – generated from approximately 60,000 isolated mature seed-derived embryos - using *LpOmt1* sense (pUbiomt1) and
30 *LpOmt1* antisense (pUbitmo1) transformation vectors were identified [16 pUbiomt1 transgenic plants and 27 pUbitmo1 transgenic plants] (Figure 35).

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Southern hybridization analysis was performed with undigested and *HindIII*-digested DNA samples prepared from the PCR positive transgenic perennial ryegrass plants, to demonstrate their transgenic nature and the integration of the chimeric *npt2* and *LpOmt1* transgenes. Independent
5 transgenic perennial ryegrass plants co-transformed with both, the selectable marker *npt2* gene and *LpOmt1* chimeric genes, were identified (Figure 35). In most instances, the transgenic perennial ryegrass plants recovered contained multiple copies of the selectable marker gene including rearranged transgene copies. No *npt2*-hybridizing bands were detected in the untransformed
10 negative control.

Samples of *HindIII*-digested genomic DNA were included in the analysis when the *LpOmt1* gene-specific hybridization probe (*omt1*) was used. The *omt1* probe hybridized to a number of bands in DNA samples corresponding to both, the transgenic plants and the untransformed negative control. The *omt1*-
15 hybridizing bands shared in all samples correspond to endogenous *LpOmt1* gene sequences represented as a small multigene family in the perennial ryegrass genome (Heath et al. 1998). The different *omt1*-hybridizing bands evident in the samples from the transgenic plants and absent in the untransformed negative control sample correspond to antisense (*tmo1*) and
20 sense (*omt1*) *LpOmt1* transgene integration events (Figure 35).

Northern hybridization analysis using strand-specific *LpOmt1* probes allowed the identification of transgenic perennial ryegrass plants expressing the antisense *LpOmt1* transgene (Figure 35).

The OMT activity of selected antisense and sense *LpOmt1* transgenic
25 perennial ryegrass plants was determined. Biochemical assays for OMT activity were initially established in untransformed plants (such as tobacco and perennial ryegrass). The assays utilise radiolabelled S-adenosylmethionine as the methyl donor for the OMT-catalysed conversion of caffeic acid into ferulic acid. The production of radioactive ferulic acid is measured and allows the
30 OMT activity to be determined.

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The OMT activity of selected *LpOmt1*-transgenic perennial ryegrass plants (*L. perenne* cv. Ellett) was determined. Significantly altered OMT activity in individual transformation events was observed (Figure 36). The manipulation of OMT activity in transgenic perennial ryegrass plants due to the expression of the chimeric ryegrass *LpOmt1* gene was thus demonstrated.

Transgenic perennial ryegrass plants were recovered, using biolistic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzyme, 4CL. The plant transformation vectors pUbi4CL2 and pUbi2LC4 carrying chimeric *Lp4CL2* cDNA sequences in sense and antisense orientation, respectively, driven by the constitutive maize ubiquitin (*Ubi*) promoter were used. Perennial ryegrass plants for 4CL manipulation were regenerated from Pm-resistant calli obtained from biolistic transformation of embryogenic calli using microprojectiles coated with the plasmids pHP23, carrying a chimeric *npt2* gene as selectable marker gene and the antisense pUbi2LC4.

Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants were obtained from biolistic transformation of embryogenic calli (Figure 37).

Transgenic perennial ryegrass plants were also recovered, using biolistic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzymes, CCR and CAD.

EXAMPLE 8

Genetic mapping of perennial ryegrass OMT, 4CL, CCR and CAD genes

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* clones were PCR amplified and radio-labelled for use as probes to detect

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restriction fragment length polymorphisms (RFLPs). RFLPs were mapped using 110 progeny individuals of the p150/112 perennial ryegrass reference population restricted with the enzymes described in the table below.

Clones	Polymorphic in p150/112	Enzyme mapped with	Locus	Linkage group
<i>Lp4CL1</i>	Y	<i>DraI</i>	<i>Lp4CL1</i>	2
<i>Lp4CL3</i>	Y	<i>EcoRV</i>	<i>Lp4CL3</i>	6
<i>LpCAD1</i>	Y	<i>EcoRV</i>	<i>LpCAD1</i>	2
<i>LpCAD1.2.1</i>	Y	<i>EcoRI</i>	<i>LpCAD2a</i> <i>LpCAD2b</i> <i>LpCAD2c</i>	7 - 2
<i>LpCCR1</i>	Y	<i>EcoRI</i>	<i>LpCCR1</i>	7
<i>LpOMT1</i>	Y	<i>DraI</i>	<i>LpOMT1</i>	7
<i>LpOMT2</i>	Y	<i>EcoRV</i>	<i>LpOMT2</i>	6

5

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* loci mapped to the linkage groups as indicated in the table and in Figure 40. These gene locations can now be used as candidate genes for quantitative trait loci for lignin biosynthesis associated traits such as herbage quality, dry matter digestibility, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

10

REFERENCES

Heath *et al* (1988) cDNA cloning and differential expression of three caffeic acid O-methyltransferase homologues from perennial ryegrass (*Lolium perenne*). Journal of Plant Physiology 153:649-657

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Lichtenstein, C, And J. Draper (1985) Genetic engineering of plants. In: D.M. Glover (ed.), DNA Cloning, Vol. 2, pp. 67-119, IRL Press, Washington.

Finally, it is to be understood that various alterations, modifications and/or additions may be made without departing from the spirit of the present invention as outlined herein.

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It will also be understood that the term “comprises” (or its grammatical variants) as used in this specification is equivalent to the term “includes” and should not be taken as excluding the presence of other elements or features.

Documents cited in this specification are for reference purposes only
5 and their inclusion is not an acknowledgement that they form part of the common general knowledge in the relevant art.

CLAIMS

1. A substantially purified or isolated nucleic acid or nucleic acid fragment encoding an enzyme selected from the group consisting of 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl
5 alcohol dehydrogenase (CAD), from a ryegrass (*Lolium*) or fescue (*Festuca*) species.

2. A nucleic acid or nucleic acid fragment according to claim 1 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).

10 3. A nucleic acid or nucleic acid fragment according to claim 1 encoding 4CL and including a nucleotide sequence selected from the group consisting of (a) sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in
15 Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

4. A nucleic acid or nucleic acid fragment according to claim 1 encoding CCR and including a nucleotide sequence selected from the group
20 consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

25 5. A nucleic acid or nucleic acid fragment according to claim 1 encoding CAD and including a nucleotide sequence selected from the group consisting of (a) the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9,

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11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

5 6. A vector including a nucleic acid or nucleic acid fragment according to claim 1.

7. A vector according to claim 6 further including a promoter and a terminator, said promoter, nucleic acid or nucleic acid fragment and terminator being operatively linked.

10 8. A plant cell, plant, plant seed or other plant part, including a vector according to claim 6.

9. A method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment according to claim 1 and/or a vector according to claim 6.

15 10. Use of a nucleic acid or nucleic acid fragment according to claim 1, and/or nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof as a molecular genetic marker.

20 11. A substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the enzymes 4CL, CCR and CAD.

12. A polypeptide according to claim 11 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).

25 13. A polypeptide according to claim 11 wherein said polypeptide is 4CL and includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 2, 4 and 6, respectively); and functionally active fragments and variants thereof.

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14. A polypeptide according to claim 11 wherein said polypeptide is CCR and includes an amino acid sequence selected from the group consisting of the sequence shown in Figure 10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

5 15. A polypeptide according to claim 11 wherein said polypeptide is CAD and includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

10 16. A lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part according to claim 8.

15 17. An isolated regulatory element capable of causing expression of an exogenous gene in plant cells, wherein said regulatory element is isolated from a nucleic acid or nucleic acid fragment encoding a protein selected from the group consisting of: O-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

18. A regulatory element according to claim 17 wherein said regulatory element includes an O-methyltransferase promoter.

20 19. A regulatory element according to claim 17 wherein said regulatory element includes a 4 coumarate CoA-ligase promoter.

20. A regulatory element according to claim 17 wherein said regulatory element includes a cinnamoyl CoA-reductase promoter.

25 19. A regulatory element according to claim 17 from a ryegrass (*Lolium*) or Fescue (*Festuca*) species.

20. A regulatory element according to claim 17 including the first

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approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

21. A regulatory element according to claim 17 including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto
5 (Sequence ID No: 17); or a functionally active fragment or variant thereof.

22. A regulatory element according to claim 17 including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

23. A vector including a regulatory element according to claim 17.

10 24. A vector according to claim 23 further including an exogenous gene and a terminator, said regulatory element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells.

25. A plant cell, plant, plant seed or other plant part, including a
15 vector according to claim 23.

26. A method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element according to claim 17 and/or a vector according to claim 23.

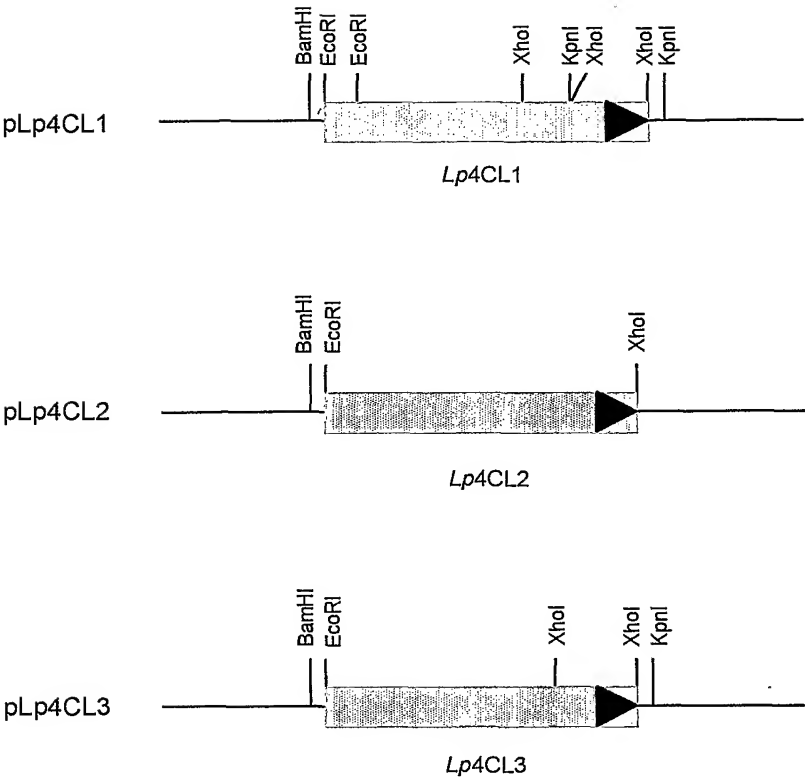


FIGURE 1

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1 CGGCACGAGTGGACTTTCCGACGCCGGAGTCGCCGATGATGACCGCCTTGAGGAGGTAGT 60
-----+-----+-----+-----+-----+-----+
61 CGTAGTCGTCCTCCGCCCTGTACGCGCCGCTGCCCGCCATTTCC'TCCTCGCCTCGCGGG 120
-----+-----+-----+-----+-----+-----+
121 TCCTCCTCCCCGACCTGCGCTAGGCTCTGGATCTCGCGGGGTTTGGGCGCGGCGTCTCG 180
-----+-----+-----+-----+-----+-----+
181 CTGTGAGCTCGTGCCGAATTCGGCACGAGCCACCTTCGAGGCGTGCACTGGTACGAGCTC 240
-----+-----+-----+-----+-----+-----+
241 GCGAGCCAT'TGTCAGTGCAGTGTAGGCTCTGCTACTCGTTGGCCATTCCAAGAAGCTCTC 300
-----+-----+-----+-----+-----+-----+
301 TGCTCCCTGAAACCAGAGGATCATGATCACGGTGGCGGCGCCCGAGGTGCAGCAGCCGCA 360
-----+-----+-----+-----+-----+-----+
M I T V A A P E V Q Q P Q
361 GATCGCGGCGGCTGCTGCGGCCGTGGAGGCGGCGGCACCGGAGGCGACGACGATCTTCCG 420
-----+-----+-----+-----+-----+-----+
I A A A A A A V E A A A P E A T T I F R
421 GTCCAGGCTCCCGGACATCGACATCCCGACCCACATGCCCTGCACGACTATTGCTTCGC 480
-----+-----+-----+-----+-----+-----+
S R L P D I D I P T H M P L H D Y C F A
481 GACGGCAGCCTCGGCCCCGGACGCGCCGTGCC'TCATCACCGCGGCCACGGGGAAGACCTA 540
-----+-----+-----+-----+-----+-----+
T A A S A P D A P C L I T A A T G K T Y
541 CACGTTCCGCCGAGACGCACCTGCTGTGCCGCAAGGCCGCGGCGGCGCTGCACGGGCTCGG 600
-----+-----+-----+-----+-----+-----+
T F A E T H L L C R K A A A A L H G L G
601 CGTGCGCCACGGGGACCGGATCATGCTGCTGCTCCAGAACTCCGTGGAGTTCGCGCTCGC 660
-----+-----+-----+-----+-----+-----+
V R H G D R I M L L L Q N S V E F A L A
661 CTTCTTCGGCGCGTCCATGCTCGGCGCCGTCAGCACGGCGGCGAACCCGTTCTGCACGCC 720
-----+-----+-----+-----+-----+-----+
F F G A S M L G A V S T A A N P F C T P
721 GCAGGAGATCCACAAGCAGCTCGTGGCCTCCGGCGCGAAGCTGGTCGTCACGCAGTCCGC 780
-----+-----+-----+-----+-----+-----+
Q E I H K Q L V A S G A K L V V T Q S A

FIGURE 2

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```

781  CTACGTCGACAAGCTCCGGGCACGAGGCCTTCCCCCGAATCGGCGAGGCCCTCACCCTGAT      840
      +-----+-----+-----+-----+-----+-----+
      Y V D K L R H E A F P R I G E A L T V I

841  CACCATCGACGAGGACGACGGCACCCCGACGGCTGCCAGCCGTTCTGGGCCCTCGTGTC      900
      +-----+-----+-----+-----+-----+-----+
      T I D E D D G T P D G C Q P F W A L V S

901  AGCCGCCGACGAGAACAGCGTCCCGGAGTCTCCCATCTCGCCGGACGACGCGGTGGCGCT      960
      +-----+-----+-----+-----+-----+-----+
      A A D E N S V P E S P I S P D D A V A L

961  GCCCTACTCGTCGGGCACGACGGGGCTGCCAAGGGCGTGCTGCTGACGCACGGGGGGCT      1020
      +-----+-----+-----+-----+-----+-----+
      P Y S S G T T G L P K G V V L T H G G L

1021 GGTGTCGAGCGTGGCGCAGCAGGTGGACGGCGAGAACCCGAACCTGCACATGCGGGCGGG      1080
      +-----+-----+-----+-----+-----+-----+
      V S S V A Q Q V D G E N P N L H M R A G

1081 GGAGGACGTGGTGCTCTGCGTGCTGCCGCTCTTCCACATCTTCTCGCTCAACTCGGTGCT      1140
      +-----+-----+-----+-----+-----+-----+
      E D V V L C V L P L F H I F S L N S V L

1141 GCTGTGCGCGCTGCGGGCGGGCGCCCGCTGATGCTGATGCCTAGGTTGAGATGGGGGC      1200
      +-----+-----+-----+-----+-----+-----+
      L C A L R A G A A V M L M P R F E M G A

1201 CATGCTGGAGGGCATCGAGCGGTGGCGCGTACGGTGGCGGCCGTTGGTGCCGCCGCTGGT      1260
      +-----+-----+-----+-----+-----+-----+
      M L E G I E R W R V T V A A V V P P L V

1261 GCTCGCGCTCGCAAGAACCCCGGGGTGGAGAAGCACGACCTCAGCTCCATTCGGATCGT      1320
      +-----+-----+-----+-----+-----+-----+
      L A L A K N P G V E K H D L S S I R I V

1321 GCTCTCCGGCGCCGCGCCGCTCGGCAAGGAGCTCGAGGACGCGCTACGTGGCCGCGCTGCC      1380
      +-----+-----+-----+-----+-----+-----+
      L S G A A P L G K E L E D A L R G R L P

1381 GCAGGCCATCTTCGGACAGGGCTACGGGATGACGGAGGCCGGGCCGGTGCTGTCCATGTG      1440
      +-----+-----+-----+-----+-----+-----+
      Q A I F G Q G Y G M T E A G P V L S M C

1441 CCCGGCGTTTCGCGCGGGAGCCGACCCGGCCAAGTCCGGCTCGTGCGGCACCGTGGTGCG      1500
      +-----+-----+-----+-----+-----+-----+
      P A F A R E P T P A K S G S C G T V V R

```

FIGURE 2 CONTINUED

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```
CAACGCCCAGCTCAAGGTGGTCGACCCCGACACCGGCGTCTCCCTCGGCCGCAACCTCCC
1501 -----+-----+-----+-----+-----+-----+-----+ 1560
      N A Q L K V V D P D T G V S L G R N L P

CGGCAGATCTGCATCCGCGGCCCGCAGATCATGAAAGGATACTTGAATGATCCCGTGGC
1561 -----+-----+-----+-----+-----+-----+-----+ 1620
      G E I C I R G P Q I M K G Y L N D P V A

CACGCCGCGACCATCGACGTCGAGGGGTGGCTCCACACCGGCGACATCGGCTACGTCTGA
1621 -----+-----+-----+-----+-----+-----+-----+ 1680
      T A A T I D V E G W L H T G D I G Y V D

CGACGACGACGAGGTCTTCATCGTCGACCGCGTCAAGGAGCTCATCAAGTTCAAGGGCTT
1681 -----+-----+-----+-----+-----+-----+-----+ 1740
      D D D E V F I V D R V K E L I K F K G F

CCAGGTACCGCCGCGGAGCTCGAGGCTCTGCTCATCGCGCATCCGTCCATCGCCGACGC
1741 -----+-----+-----+-----+-----+-----+-----+ 1800
      Q V P P A E L E A L L I A H P S I A D A

GGCCGTCGTCCCGCAAAAGGATGATGCCGCCGCGGAGGTCCCGGTTGCCTTCGTGGTCCG
1801 -----+-----+-----+-----+-----+-----+-----+ 1860
      A V V P Q K D D A A G E V P V A F V V R

CGCCGCGGACTCCGACATCGCCGAGGAGGCCATCAAGGAGTTCTGATCCAAGCAGGTGGT
1861 -----+-----+-----+-----+-----+-----+-----+ 1920
      A A D S D I A E E A I K E F V S K Q V V

GTTCTACAAGAGGCTGCACAAGGTCTACTTCACCCACGCGATACCCAAGTCGGCGTCGGG
1921 -----+-----+-----+-----+-----+-----+-----+ 1980
      F Y K R L H K V Y F T H A I P K S A S G

GAAGATACTCAGGAAAGAACTCAGAGCTAAACTCGCCGCCCCGGCCACTGCCTGAAGAGT
1981 -----+-----+-----+-----+-----+-----+-----+ 2040
      K I L R K E L R A K L A A P A T A * R V

GGTTCATGGCTTCATGCTAATCATTTTCGATCAGAAAGGCACTTCTAGCATATATGTTCCA
2041 -----+-----+-----+-----+-----+-----+-----+ 2100
      V H G F M L I I S I R K A L L A Y M F H

CCTTTTGTTTCATTTGGAAGATTGTATTCCAGCTAGTGGCCAGTGAAGTAAGGGATG
2101 -----+-----+-----+-----+-----+-----+-----+ 2160
      L L F H L E D C I P A S G Q *

GGGATAAAAGTTTTGTCTACGTTTTCTTTTACGCTACTCTCTCATTTGGGGAGTACAATG
2161 -----+-----+-----+-----+-----+-----+-----+ 2220

TATCAGGGGATTTCGTGATTGAAGTTAATCAAGATTGGTTCAATTATAAAAAAAAAAAAAA
2221 -----+-----+-----+-----+-----+-----+-----+ 2280

AAAA
2281 ---- 2284
```

FIGURE 2 CONTINUED

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1 CGGCACGAGCGCCATTCTCCACCTTCAGCTCCGGCCAAAGATTTCCATCCGGCGAGATC 60
-----+-----+-----+-----+-----+-----+
61 CATGGGCTCCATCGCGGCGGACGCGCCTCCCGCGGAGCTGGTGTTCGGGTCCAAGCTCCC 120
-----+-----+-----+-----+-----+-----+
M G S I A A D A P P A E L V F R S K L P
121 GGACATCGAGATCCCGACCCACCTGACGCTGCAGGACTACTGCTTCCAGCGCCTGCCGGA 180
-----+-----+-----+-----+-----+-----+
D I E I P T H L T L Q D Y C F Q R L P E
181 GCTCTCCGCGCGCGCCTGCCTCATCGACGGCGCCACGGGCGCCGCGCTCACCTACGGCGA 240
-----+-----+-----+-----+-----+-----+
L S A R A C L I D G A T G A A L T Y G E
241 GGTGGACGCCCTGTCCCGCGCTGCGCCGCGGGGCTGCGCCGCGCTCGGCGTCGGCAAGGG 300
-----+-----+-----+-----+-----+-----+
V D A L S R R C A A G L R R L G V G K G
301 CGACGTCGTCATGGCGCTCCTCCGCAACTGCCCCGAGTTTCGCCTTCGTGTTCCTCGGCGC 360
-----+-----+-----+-----+-----+-----+
D V V M A L L R N C P E F A F V F L G A
361 GGCCCGGCTCGGCGCCGCCACCACCACCGCCAACCCGTCTCTACACGCCCCACGAGATCCA 420
-----+-----+-----+-----+-----+-----+
A R L G A A T T T A N P F Y T P H E I H
421 CCGCCAGGCCACCGCCGCGGGGCCAGGGTCATCGTCACCGAGGCCTGCGCCGTCGAGAA 480
-----+-----+-----+-----+-----+-----+
R Q A T A A G A R V I V T E A C A V E K
481 GGTGCGCGCCTTCGCGCCGAGAGAGGGATTCCCGTCGTCTCCGTCGACGAGGGCGTCGA 540
-----+-----+-----+-----+-----+-----+
V R A F A A E R G I P V V S V D E G V D
541 CGGCGGCTGCCTCCCGTTCCGCCGAGACTCTGCTCGGGGAAGAAAGCGGGGAGCGGTTTCGT 600
-----+-----+-----+-----+-----+-----+
G G C L P F A E T L L G E E S G E R F V
601 CGACGAGGCGGTTCGACCCCGACGACGTGGTGGCGCTGCCGTACTCGTCCGGCACCACCGG 660
-----+-----+-----+-----+-----+-----+
D E A V D P D D V V A L P Y S S G T T G
661 CCTGCCCAAGGGCGTCATGCTCACCCACCGCAGCCTCGTCACCAGCGTCGCCCAGCAGGT 720
-----+-----+-----+-----+-----+-----+
L P K G V M L T H R S L V T S V A Q Q V
721 GGACGGTGAGAACCCGAACCTGCACTTCAGCTCGTCGGACGTGCTGCTGTGCGTGCTGCC 780
-----+-----+-----+-----+-----+-----+
D G E N P N L H F S S S D V L L C V L P

FIGURE 3

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781	GCTGTTCCACATCTACTCGCTCAACTCGGTGCTGCTCGCCGGTCTCCGCGCCGGGTGCGC -----+-----+-----+-----+-----+-----+-----+-----+-----+ L F H I Y S L N S V L L A G L R A G C A	840
841	GATCGTGATCATGCGCAAGTTCGACCACGGCGCGCTGGTGGACCTGGTGCGCACGCACGG -----+-----+-----+-----+-----+-----+-----+-----+-----+ I V I M R K F D H G A L V D L V R T H G	900
901	CGTCACCGTGGCGCCATTTCGTGCCGCCCATCGTGGTGGAGATCGCCAAGAGCGCGCGGGT -----+-----+-----+-----+-----+-----+-----+-----+-----+ V T V A P F V P P I V V E I A K S A R V	960
961	GACCGCCGCGGACCTGGCGTCCATCCGGCTGGTTCATGTCGGGGGCGGCGCCCATGGGCAA -----+-----+-----+-----+-----+-----+-----+-----+-----+ T A A D L A S I R L V M S G A A P M G K	1020
1021	GGAGCTGCAGGACGCGTTTCATGGCCAAGATCCCCAACGCCGTGCTCGGCCAGGGATATGG -----+-----+-----+-----+-----+-----+-----+-----+-----+ E L Q D A F M A K I P N A V L G Q G Y G	1080
1081	GATGACCGAGGCCGGCCCTGTGCTGGCGATGTGCCTGGCCTTCGCCAAGGAGCCGTTTCGC -----+-----+-----+-----+-----+-----+-----+-----+-----+ M T E A G P V L A M C L A F A K E P F A	1140
1141	GGTCAAGTCCGGTTCCTGCGGCACCGTCGTCAGGAACGCCGAGCTCAAGATCGTCGACCC -----+-----+-----+-----+-----+-----+-----+-----+-----+ V K S G S C G T V V R N A E L K I V D P	1200
1201	CGACACCGGCGCCTCCCTCGGCCGCAACCTGCCGGGGGAGATCTGCATCCGCGGCAAGCA -----+-----+-----+-----+-----+-----+-----+-----+-----+ D T G A S L G R N L P G E I C I R G K Q	1260
1261	GATCATGAAAGGTTACCTAAATGATCCGGTGGCCACAAAGAACACCATTGACAAGGACGG -----+-----+-----+-----+-----+-----+-----+-----+-----+ I M K G Y L N D P V A T K N T I D K D G	1320
1321	TTGGCTGCATACTGGTGACATTGGTTATGTGATGATGACGACGAGATCTTTATTGTCTGA -----+-----+-----+-----+-----+-----+-----+-----+-----+ W L H T G D I G Y V D D D D E I F I V D	1380
1381	CAGACTGAAGGAGATAATTAAATATAAGGGATTCCAAGTACCTCCGGCGGAACTTGAAGC -----+-----+-----+-----+-----+-----+-----+-----+-----+ R L K E I I K Y K G F Q V P P A E L E A	1440
1441	CCTTCTCATTTACACACCCTGAAATCAAGGATGCTGCTGTCGTATCGATGCAAGACGAACT -----+-----+-----+-----+-----+-----+-----+-----+-----+ L L I T H P E I K D A A V V S M Q D E L	1500
1501	TGCTGGTGAAGTTCCGGTTGCGTTTGTGTGCGGACTGAGGGTTCAGAGATCAGCGAAAA -----+-----+-----+-----+-----+-----+-----+-----+-----+ A G E V P V A F V V R T E G S E I S E N	1560

FIGURE 3 CONTINUED

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```
1561  CGAGATCAAGCAGTTCGTTGCAAAAGAGGTTGTTTCTACAAGAGGATCTGCAAAGTGTT 1620
      -----+-----+-----+-----+-----+-----+
      E I K Q F V A K E V V F Y K R I C K V F

1621  CTTGCGGGATTCCATTCCAAGAGTCCATCTGGCAAGATCCTCAGGAAGGACCTGAGAGC 1680
      -----+-----+-----+-----+-----+-----+
      F A D S I P K S P S G K I L R K D L R A

1681  AAAGCTCGCCGCAGGCATTCCCAGCAGTAATACCACACAGTCCAAAAGCTAAGTCAGATA 1740
      -----+-----+-----+-----+-----+-----+
      K L A A G I P S S N T T Q S K S *

1741  TATTGTTTCCCAACCTTACACACCTCTGTCCAACACCATGTAATGTTCTTAATATAAACG 1800
      -----+-----+-----+-----+-----+-----+

1801  GAAATTATTACATATAGAAGGGCTGATTCTTTTTTACTAGATGTGTCCAACATATGATATG 1860
      -----+-----+-----+-----+-----+-----+

1861  CTTGTTAGCCGATGATGTGTAACCTGTCATGTATAGATACCGCCTTTTTTTGACAAGAA 1920
      -----+-----+-----+-----+-----+-----+

1921  AGGCTGATTATAATGTATACCGTGAAGTGAATATTTGTTTCAGGGAGATCAAAAAAAAAA 1980
      -----+-----+-----+-----+-----+-----+

1981  AAAAAAAAAA
      -----+-- 1992
```

FIGURE 3 CONTINUED

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1 CGGCACGAGATCTCCACGACTAATTTAGAAGAAGATTTACTTAGTCTCTGCTTCTCGCT 60
-----+-----+-----+-----+-----+-----+
61 CGATCGCCGCGCCGGTGAGGTAGCTAGCTAGCTACTCGTACTAGACCATTACCATGGGTTTC 120
-----+-----+-----+-----+-----+-----+
M G S
121 CGTGCCGAGGAGTCAAGTGGTGGCGGTGGCACC GCGGAGACGGTGTTCGGTCTGAAGCT 180
-----+-----+-----+-----+-----+-----+
V P E E S V V A V A P A E T V F R S K L
181 CCCCACATCGAGATCAACAACGAGCAGACGCTGCAGAGCTACTGCTTCGAGAAGATGGC 240
-----+-----+-----+-----+-----+-----+
P D I E I N N E Q T L Q S Y C F E K M A
241 CGAGGTCGCGTCCCGCCCCCTGCATCATCGACGGCCAGACGGGCGCCTCCTACACCTACAC 300
-----+-----+-----+-----+-----+-----+
E V A S R P C I I D G Q T G A S Y T Y T
301 GGAGGTCGACTCCCTGACCCGTCGCGCCGCGGGGCTGCGCCGCATGGGCGTGGGGAA 360
-----+-----+-----+-----+-----+-----+
E V D S L T R R A A A G L R R M G V G K
361 GGGCGACGTGGTGATGAACCTGCTGCGCAACTGCCCCGAGTTTCGCCTTCTCCTTCTGGG 420
-----+-----+-----+-----+-----+-----+
G D V V M N L L R N C P E F A F S F L G
421 CGCGGCGCGGCTGGGCGCCGCCACCACCACCGCCAACCCGTTCTACACCCCGCACGAGAT 480
-----+-----+-----+-----+-----+-----+
A A R L G A A T T T A N P F Y T P H E I
481 CCACCGCCAGGCGGAGGCGGCGGCCAAGCTGATCGTGACCGAGGCCTGCGCCGTGGA 540
-----+-----+-----+-----+-----+-----+
H R Q A E A A G A K L I V T E A C A V E
541 GAAGGTGCTGGAGTTCGCGGCGGGGCGGGGCGTGGCGTACCGTCGACGGGAGGCG 600
-----+-----+-----+-----+-----+-----+
K V L E F A A G R G V P V V T V D G R R
601 CGACGGGTGCGTGGACTTCGCGGAGCTGATCGCCGCGAGGAGCTGCCCAGGCGGACGA 660
-----+-----+-----+-----+-----+-----+
D G C V D F A E L I A G E E L P E A D E
661 GGCCGGGGTCTCTCCCGACGACGTCGTCGCCCTGCCCTACTCTCCGGCACCACCGGGCT 720
-----+-----+-----+-----+-----+-----+
A G V L P D D V V A L P Y S S G T T G L
721 CCCCAGGGCGTCATGCTCACCCACCGCAGCCTCGTCACCAGCGTCGCCCAGCTGGTCGA 780
-----+-----+-----+-----+-----+-----+
P K G V M L T H R S L V T S V A Q L V D

FIGURE 4

	CGGGTCGAACCCCTAACGTGTGCTTCAACAAGGACGCACGCGCTGCTGTGCCTGCTGCCGCT	
781	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	840
	G S N P N V C F N K D D A L L C L L P L	
	CTTCCACATCTACTCGTGTCACACGGTGC'TGCTGGCGGGGCTCCGCGCTCGGCGCCGCCAT	
841	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	900
	F H I Y S L H T V L L A G L R V G A A I	
	CGTCATCATGCGCAAGTTTCGACGTCGGCGCGCTGGTGGACC'TCGTCCGCGCGCACCCGAT	
901	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	960
	V I M R K F D V G A L V D L V R A H R I	
	CACCATCGCGCCATTTCGTGCCGCCCATCGTCGTGGAGATCGCCAAGAGCGACCGCGTCGG	
961	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1020
	T I A P F V P P I V V E I A K S D R V G	
	CGCCGACGACCTCGCATCCATCCGCATGGTGTCTC'CGGCGCCGCGCCCATGGGCAAGGA	
1021	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1080
	A D D L A S I R M V L S G A A P M G K D	
	CCTCCAGGACGCCTTCATGGCCAAGATCCCCAACGCCGTGCTCGGACAGGGGTACGGGAT	
1081	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1140
	L Q D A F M A K I P N A V L G Q G Y G M	
	GACCGAGGCTGGGCGCGGTGCTGGCCATGTGCCTGGCGTTCGCCAAGGAGCCGTTCAAGGT	
1141	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1200
	T E A G P V L A M C L A F A K E P F K V	
	CAAGTCCGGGTCTGTGCGGAACCGTGGTGC'GCAACGCCGAGCTCAAGGTCTGACCCCCGA	
1201	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1260
	K S G S C G T V V R N A E L K V V D P D	
	CACCGGCGCATCCCTCGGCCGGAACAGCCTGGCGAGATTTGCGTCCGGGGGAAGCAGAT	
1261	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1320
	T G A S L G R N Q P G E I C V R G K Q I	
	CATGATAGGTTACCTGAACGACCCAGAGTCGACCAAGAACACCATCGACAAGGACGGCTG	
1321	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1380
	M I G Y L N D P E S T K N T I D K D G W	
	GCTGCACACCGGAGACATCGGCTTGGTGGATGACGACGAGATCTTCATCGTCGACAG	
1381	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1440
	L H T G D I G L V D D D D E I F I V D R	
	GCTCAAGGAGATCATCAAGTACAAGGGCTTCCAAGTGGCGCCGGCGGAGCTCGAGGCCCT	
1441	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1500
	L K E I I K Y K G F Q V A P A E L E A L	
	CCTCCTCACGAACCCGGAGGTCAAGGACGCCGCGCTCGTAGGGGTGAAGGATGATCTCTG	
1501	- - - - + - - - - + - - - - + - - - - + - - - - + - - - - + - - - - +	1560
	L L T N P E V K D A A V V G V K D D D L C	

FIGURE 4 CONTINUED

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```
1561  CGGCGAAGTCCCGGTCGCCTTCATTAAGAGGATCGAAGGATCTGAGATCAACGAGAACGA 1620
      -----+-----+-----+-----+-----+-----+
      G E V P V A F I K R I E G S E I N E N E

1621  GATCAAGCAATTCGTCTCAAAGGAGGTTGTTTTCTACAAGAGGATCAACAAGGTCTACTT 1680
      -----+-----+-----+-----+-----+-----+
      I K Q F V S K E V V F Y K R I N K V Y F

1681  CACCGACTCCATTCCCAAGAACCCTTCCGGCAAGATCCTAAGGAAGGACTTGAGAGCCAG 1740
      -----+-----+-----+-----+-----+-----+
      T D S I P K N P S G K I L R K D L R A R

1741  GCTCGCCGCTGGCATCCCCACCGAAGTTGCCGCGCCGAGAAGCTAAGGGCCGCTTCTCAG 1800
      -----+-----+-----+-----+-----+-----+
      L A A G I P T E V A A P R S *

1801  GAACGCAGTCACCCATGGTGCTGTTTAGGTGCTGTTATAGACCACACCAAATGGGGAAAG 1860
      -----+-----+-----+-----+-----+-----+

1861  AAACACGGGAGGGGATCATATTATTGTTGCAGGAGATATCAGTTTGTGATTGCCCCCTG 1920
      -----+-----+-----+-----+-----+-----+

1921  CTTGTGTAATGTTGATAAAATGAAATGATATAATAGATGTGTTGTTTATTTTTTGACCA 1980
      -----+-----+-----+-----+-----+-----+

1981  TGTAAGAACAAGGCTGTTTTATACACTACTTATTTTTTGAAAAAAAAAAAAAAAAAAAA 2038
      -----+-----+-----+-----+-----+-----+
```

FIGURE 4 CONTINUED

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FIGURE 5

	10	20	30	40	50	60
Lp4CL1	MITVAAPEVQQPQIAAAAAVEAAPEATTIERSRLPDIDITPTHMPHDYCFATAASAPD					
Lp4CL2	MGSIAADAPPAEL. .VFRSLPDIEIPTHITLQDYCFORLPELSA					
Lp4CL3	MGSVPEESVAVAPAETVFRSLPDIEINNEQTLQSYCFEKMAEVAS					
	70	80	90	100	110	120
Lp4CL1	APCLITAAATGKTYTFAETHLLCRKAAAALHGLCVRHGDRIMLLONSVEFALAFFGASML					
Lp4CL2	RACLDGATGAALTYGEVDALSRCAAGLRRLCVGKGDVVMALLRNCPEFAFVELGAARL					
Lp4CL3	RPCIIDGQTCASYTYTEVDSLIRRAAAGLRRCVVGKGDVVMNLLRNCPEFAFSFLGAARL					
	130	140	150	160	170	180
Lp4CL1	GAVSTAAFPFCTPQETHKOLVASGAKLVVTQSAYVDKLRHEAFPRIGEALTIVITIDEDDG					
Lp4CL2	GAATTTANPFYTPETHRQATAAGARVIVTEACAVEKVRFAFAERGIPVVSV.DE					
Lp4CL3	GAATTTANPFYTPETHRQAEAAGAKLIVTEACAVEKVFLEFAAGRGVPVTV.DG					
	190	200	210	220	230	240
Lp4CL1	TPDGCQPFWALVSAADENSVPESPIS. .PDDAVALPYSSGTTGLPKGVVLTHGGLVSSVA					
Lp4CL2	GVDGSCLPFAETLLGEESSGERFVDEAVDPDDVVALPYSSGTTGLPKGVMLTHRSLSVTSVA					
Lp4CL3	RRDGCVDFAELIAGEELPEADEAGVL. PDDVVALPYSSGTTGLPKGVMLTHRSLSVTSVA					
	250	260	270	280	290	300
Lp4CL1	QQVDGENPNLHMRAGEDVVLCLVLPFHIFSLNSVLLCALRAGAAVMMPRFEMGAMLEGI					
Lp4CL2	QQVDGENPNLHFSS. SDVLLCLVLPFHIIYSLSNVLLAGLRAGCAIVIMRKFDHGALVDLV					
Lp4CL3	QLVDGSENPNVCFNK. DDALLCLLPFHIIYSLHTVLLAGLRVGAAIVIMRKFDVGALVDLV					
	310	320	330	340	350	360
Lp4CL1	ERWRVTVAAVVPPIVLALAKNPGVEKEDLSIRIVLSGAAPLKGKELDALRGRLEQALFG					
Lp4CL2	RTHGVTVAPFVPIVVEIAKSARVTAADLASIRIVMSGAAPMGKELQDAFMAKIEPNAVLG					
Lp4CL3	RAHRTTIAPEFVPIVVEIAKSDRVGADDLASIRIVLSGAAPMGKELQDAFMAKIEPNAVLG					
	370	380	390	400	410	420
Lp4CL1	QGYGMTEAGPVLAMCPAFAREPTPAKSGSCGTVVRNAOLKVVDPDGTGVSLSGRNLPGEICL					
Lp4CL2	QGYGMTEAGPVLAMCLAFAREPFAVKSGSCGTVVRNAELKIVDPDTGASLSGRNLPGEICL					
Lp4CL3	QGYGMTEAGPVLAMCLAFAREPFVKSGSCGTVVRNAELKVVDPDGTGASLSGRNLPGEICV					
	430	440	450	460	470	480
Lp4CL1	RGKQIMKGYLNDEPVATAATIDVEGWLHTGDIGYVDDDDDEIFIVDRVKELIKFKGFQVPPA					
Lp4CL2	RGKQIMKGYLNDEPVATKNTIDKDGWLHTGDIGYVDDDDDEIFIVDRVKELIKFKGFQVPPA					
Lp4CL3	RGKQIMKGYLNDEPSTKNTIDKDGWLHTGDIGYVDDDDDEIFIVDRVKELIKFKGFQVAPA					
	490	500	510	520	530	540
Lp4CL1	ELEALLIAHPISIADAAVVPQKDDAAGEVPVAFVVRADSDIAEEATKEFVSKQVVFYKRL					
Lp4CL2	ELEALLITHPEIKDAAVVSMQDELAGEVPVAFVVRTEGSEISENEIKQFVAKEVVFYKRI					
Lp4CL3	ELEALLLTNPEVKDAAVVGKDDLCGEVPVAFIKRIEGSEINENEIKQFVSKEVVFYKRI					
	550	560	570			
Lp4CL1	HKVVFTHATPKSASGKILRKELRAKLAA PATA					
Lp4CL2	CKVVFADSIKSPSGKILRKDLRAKLAA GIPSSNTTQSKS					
Lp4CL3	NKVVFETDSIKNPSSGKILRKDLRAKLAA GIPTEVAAPRS					

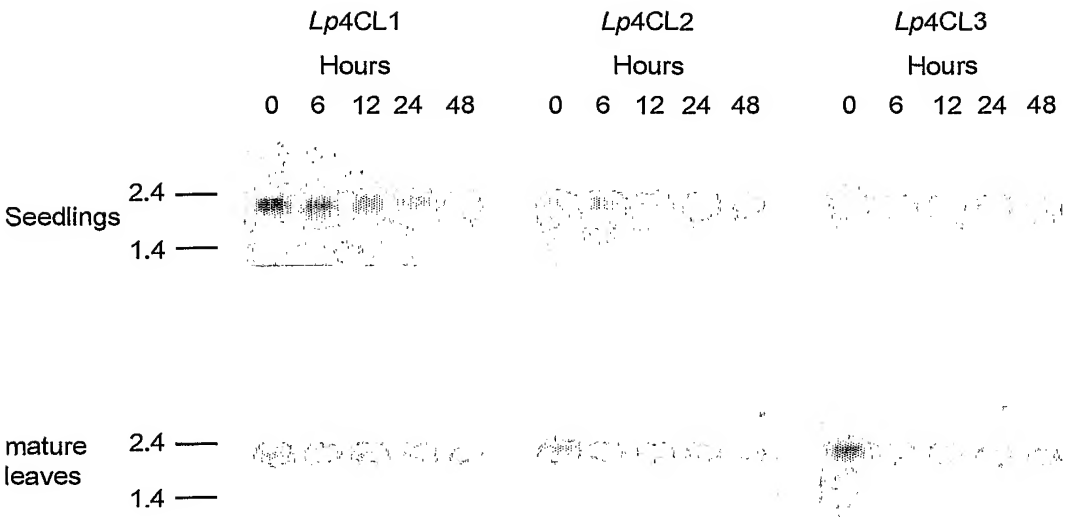


FIGURE 7

FIGURE 8

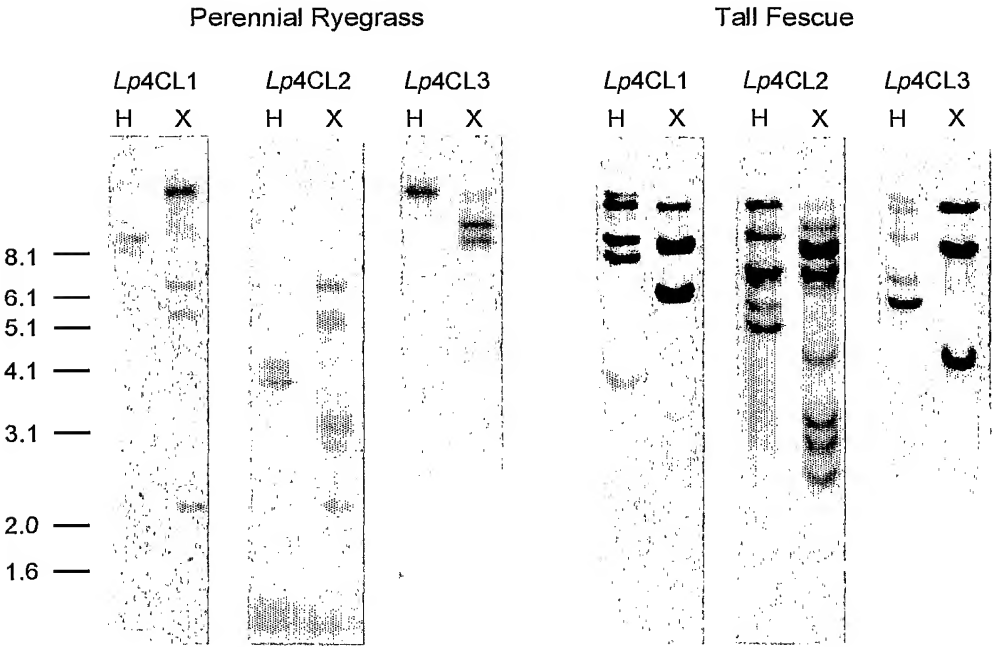
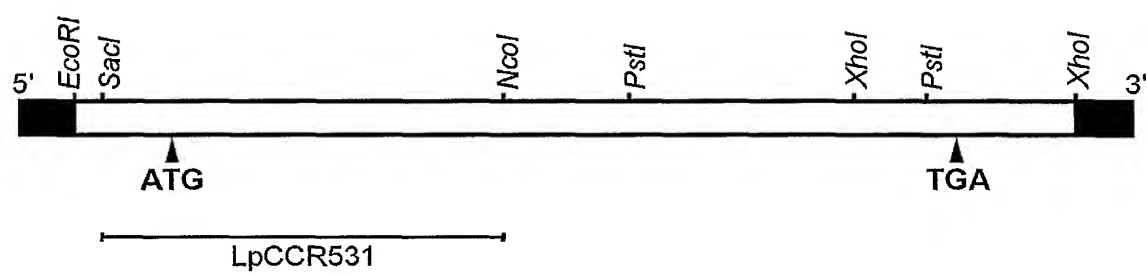


FIGURE 9



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1  GGCACGAGGAATCCTACCAAACCGAGCTACCAGATCCTTCTCTACTAATCGAGCTCCCTA 60
   -----+-----+-----+-----+-----+-----+
61  CGCTGCTCCGCCTGTCTTCGTTTCCGCCTCACCGCCGGCCGGTTCTCCGCTCCAAGCTAC 120
   -----+-----+-----+-----+-----+-----+
121 GTCCGTCCGTCCACATATATAGCATCGACATGACCATCGCCGAGGTCGTGGCTGCCGGAG 180
      -----+-----+-----+-----+-----+-----+
      M T I A E V V A A G D
181  ACACCGCCGCCCGCGGTGGTGCAGCCCGCCGGAACGGGCAGACCGTGTGCGTGACCGGCG 240
      -----+-----+-----+-----+-----+-----+
      T A A A V V Q P A G N G Q T V C V T G A
241  CCGCCGGGTACATCGCGTCTGGCTCGTCAAGCTGCTGCTGGAGAAGGGGTACACCGTCA 300
      -----+-----+-----+-----+-----+-----+
      A G Y I A S W L V K L L L E K G Y T V K
301  AGGGCACCGTCAGGAACCCAGACGACCCGAAGAACGCGCACCTGAGGGCGCTCGACGGCG 360
      -----+-----+-----+-----+-----+-----+
      G T V R N P D D P K N A H L R A L D G A
361  CCGCCGACCGGCTGGTCCTCTGCAAGGCCGACCTCCTCGACTACGACGCCATCCGCCGCG 420
      -----+-----+-----+-----+-----+-----+
      A D R L V L C K A D L L D Y D A I R R A
421  CCATCGACGGCTGCCACGGCGTCTTCCACACCGCGTCCCCCGTCACCGACGACCCCGAGC 480
      -----+-----+-----+-----+-----+-----+
      I D G C H G V F H T A S P V T D D P E Q
481  AAATGGTGGAGCCGGCGGTGAGGGGCACGCAGTACGTATAGACGCGGCGGCGGAGGCCG 540
      -----+-----+-----+-----+-----+-----+
      M V E P A V R G T Q Y V I D A A A E A G
541  GCACGGTGCGGCGGATGGTGTCTACCTCCTCCATCGGCGCCGTCACCATGGACCCCAACC 600
      -----+-----+-----+-----+-----+-----+
      T V R R M V L T S S I G A V T M D P N R
601  GCGGGCCGGACGTGGTTCGTCGACGAGTCGTGCTGGAGCGACCTCGACTTCTGCAAGAAAA 660
      -----+-----+-----+-----+-----+-----+
      G P D V V V D E S C W S D L D F C K K T
661  CCAGGAAC TGGTACTGCTACGGGAAGGCGGTTGCGGAGCAGGCGGCATCGGAGTTGGCGC 720
      -----+-----+-----+-----+-----+-----+
      R N W Y C Y G K A V A E Q A A S E L A R
721  GGCAGCGCGGCGTGGACCTTGTGGTGGTGAACCCGGTGTGGTGATCGGCCCCCTGCTGC 780
      -----+-----+-----+-----+-----+-----+
      Q R G V D L V V V N P V L V I G P L L Q

```

FIGURE 10

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781 AGCCGACGGTGAACGCCAGCATCGGCCACATCCTCAAGTACCTGGACGGGTCGGCCAGCA 840
-----+-----+-----+-----+-----+-----+
P T V N A S I G H I L K Y L D G S A S K

841 AGTTCGCCAACGCCGTGCAGGCGTACGTGGACGTCCGCGACGTGGCCGACGCCACCTCC 900
-----+-----+-----+-----+-----+-----+
F A N A V Q A Y V D V R D V A D A H L R

901 GCGTCTTCGAGTGCGCCGCCGCGTCCGGCCGCCACCTCTGCGCCGAGCGCGTCTCCACC 960
-----+-----+-----+-----+-----+-----+
V F E C A A A S G R H L C A E R V L H R

961 GCGAGGACGTTCGTGCGCATCCTCGCCAAGCTCTTCCCCGAGTACCCCGTCCCCACCAGGT 1020
-----+-----+-----+-----+-----+-----+
E D V V R I L A K L F P E Y P V P T R C

1021 GCTCTGATGAGACGAACCCGAGGAAGCAGCCATACAAGATGTGCAACCAGAAGCTCCAGG 1080
-----+-----+-----+-----+-----+-----+
S D E T N P R K Q P Y K M S N Q K L Q D

1081 ACCTCGGACTCGAGTTCAGGCCGGTGAGCCAGTCCCTGTACGAGACGGTGAAGAGCCTCC 1140
-----+-----+-----+-----+-----+-----+
L G L E F R P V S Q S L Y E T V K S L Q

1141 AGGAGAAGGGCCACCTTCCGGTGCTCAGCGAGCAGGCAGAGGCGGACAAGGAAACCTAG 1200
-----+-----+-----+-----+-----+-----+
E K G H L P V L S E Q A E A D K E T L A

1201 CTGCCGAGCTGCAGGCAGGGGTTACCATCCGAGCATGAGGAACAAGAAATCAACCATGTC 1260
-----+-----+-----+-----+-----+-----+
A E L Q A G V T I R A *

1261 CATACTGCTACTGTCATGTAAACCAGCTGTTGAATGCCTAAAACTAAGTTCTTGTAAATA 1320
-----+-----+-----+-----+-----+-----+
CTGTGTTGTTTCATGTGGACTAGATTGATCGAATAAACATCTCTACACAAGGTTGCTAAA

1321
-----+-----+-----+-----+-----+-----+
AAAAAAAAAAAAAAAAA

1381
-----+-----+-----+-----+-----+-----+ 1395

FIGURE 10 CONTINUED

FIGURE 11

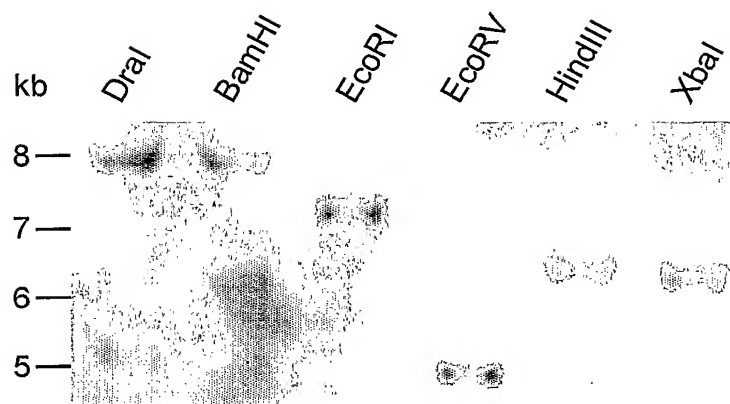
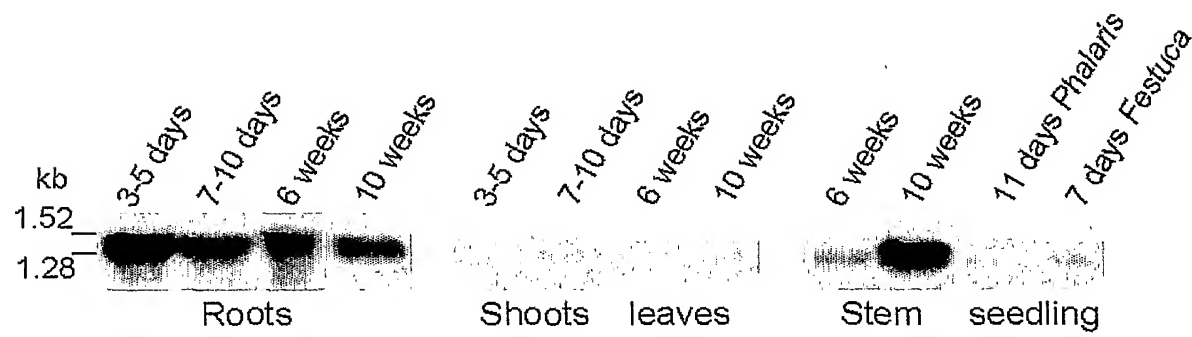


FIGURE 12



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1 GGCACGAGCAACAAGTCATCAATGGCGGAAGGCTTGCCGGCGCTCGGTTGGGCTGCGAGG 60
-----+-----+-----+-----+-----+-----+
M A E G L P A L G W A A R

61 GACGCCTCCGGTCACCTCTCCCCTTACAGCTTCTCGAGAAGCGTTCCGAAGGACGACgAT 120
-----+-----+-----+-----+-----+-----+
D A S G H L S P Y S F S R S V P K D D D

121 GTGACGATCAAGGTGCTCTTCTGCGGGATCTGCCACACTGACCTCCACATCATCAAGAAC 180
-----+-----+-----+-----+-----+-----+
V T I K V L F C G I C H T D L H I I K N

181 GACTGGGGCAACGCCCTCTACCCCATCGTCCCAGGGCATGAGATCGTGGGCGTCGTCGCC 240
-----+-----+-----+-----+-----+-----+
D W G N A L Y P I V P G H E I V G V V A

241 AGCGTCGGCAGCGGCGTCAGCAGCTTCAAGGCCGGCgACACGGTGGGCGTGGGCTACTTC 300
-----+-----+-----+-----+-----+-----+
S V G S G V S S F K A G D T V G V G Y F

301 CTCGACTCCTGCCGCACCTGCTACAGCTGCAGCAAGGGGTACGAGAACTTCTGCCCCACC 360
-----+-----+-----+-----+-----+-----+
L D S C R T C Y S C S K G Y E N F C P T

361 CTGACGCTCACCTCCAACGGCGTCGACGGCGGCGGCCACCACCCAGGGCGGCTTCTCC 420
-----+-----+-----+-----+-----+-----+
L T L T S N G V D G G G A T T Q G G F S

421 GACGTCCTCGTCGTCACAAGGACTACGTCATCCGCGTCCCGGACAACCTGCCCCTGGCC 480
-----+-----+-----+-----+-----+-----+
D V L V V N K D Y V I R V P D N L P L A

481 GGCGCGGCACCTCTCCTCTGCGCCGGCGTCACAGTCTACAGCCCTATGGTGGAGTACGGC 540
-----+-----+-----+-----+-----+-----+
G A A P L L C A G V T V Y S P M V E Y G

541 CTCAACGCCCCcgGGAAGCACyTCGGcGTCGTCGGCCTGGGCGGGCTCGGCCACGTCGcC 600
-----+-----+-----+-----+-----+-----+
L N A P G K H X G V V G L G G L G H V A

601 GTCAAGTTCGGCAAGGCCTTCGGGATGACCGTCACCGTCATCAGCTCCTCGGACAGGAAG 660
-----+-----+-----+-----+-----+-----+
V K F G K A F G M T V T V I S S S D R K

661 CGCGACGAGGCGCTCGGCCGCCCTCGGCGCCGACGCcTTCTCTGTCAGCAGCGACCCGAG 720
-----+-----+-----+-----+-----+-----+
R D E A L G R L G A D A F L V S S D P E

FIGURE 13

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```

721 CAGATGAAGGCGGCGGGCGGGCACCATGGACGGCATCATCGACACGGTGTCCGCGGGCCAC 780
-----+-----+-----+-----+-----+-----+
Q M K A A A G T M D G I I D T V S A G H

781 CCGATCGTGCCGCTGCTCGACCTGCTCAAGCCCATGGGGCAGATGGTTCGTGGTGGGCGCG
-----+-----+-----+-----+-----+-----+ 840
P I V P L L D L L K P M G Q M V V V G A

841 CCCAGCAAGCCGCTCGAGCTCCCGGCCTTCGCCATCATCGGCGGGCGGCAAGCGCCTCGCC
-----+-----+-----+-----+-----+-----+ 900
P S K P L E L P A F A I I G G G K R L A

901 GGGAGCGGCACCGGCAGCGTCGCACACTGCCagGCCATGCTCGACTTCGCGGGCAAGCAC
-----+-----+-----+-----+-----+-----+ 960
G S G T G S V A H C Q A M L D F A G K H

961 GGCATCACCGCCGACGTCGAGGTCGTCAAGATGGACTACgGTCAACACCGCCATCGAGCG
-----+-----+-----+-----+-----+-----+ 1020
G I T A D V E V V K M D Y G Q H R H R A

1021 GCTAGAGAAGAACGACGTCAGGTACCGCTTCGTTCATCGACGTCGCCGGCAGCCACCTGCA
-----+-----+-----+-----+-----+-----+ 1080
A R E E R R Q V P L R H R R R R Q P P A

1081 GGGCACCGCCGCTTAACTTGTGCTACACAATGTGGACGCGCGCTCGTTTGGTCCAGAAAA
-----+-----+-----+-----+-----+-----+ 1140
G H R R L T C A T Q C G R A L V W S R K

1141 AGGTTCCGCGGCTCACAGCCACATGAACAAGTCAATGAGTCGTTGGTGTGTGTTTATCT
-----+-----+-----+-----+-----+-----+ 1200
R F A G S Q P H E Q V N E S L V C C L S

1201 TCATTCCACATATGGGACGCAGTTCAGATTTTCATGTCAAATAATTGCGTCGTGTGCGG
-----+-----+-----+-----+-----+-----+ 1260
S F H I W D A V P D F H V K

1261 TTGTCAAGACTCAAATAGGAGAAAAAAAGACTCGTGATTTCGTTTTGCAAAAAAAAAAAAA
-----+-----+-----+-----+-----+-----+ 1320

1321 AAAAA 1325
-----
```

FIGURE 13 CONTINUED

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GGCACGAGTCGCCTCCAACGCTCTCCCTTAACCGGCCGTCCCTACGCTTGCACCACCACC
1 -----+-----+-----+-----+-----+-----+-----+ 60

ACGCACAGACAGAGCAGTTTCCCAGCCCCCGCCGAACCGGATGGCACCCACGGCGGCGG
61 -----+-----+-----+-----+-----+-----+-----+ 120
M A P T A A E

AGCAGACGGAGCACCACCAGCACACCAGGAAGGCGGTGGGGCTGGCGGCGCGCGACGACG
121 -----+-----+-----+-----+-----+-----+-----+ 180
Q T E H H Q H T R K A V G L A A R D D A

CCGGCCACCTCTCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA
181 -----+-----+-----+-----+-----+-----+-----+ 240
G H L S P L A I T R R S T G D D D V V I

TAAAGATTTTGTACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA
241 -----+-----+-----+-----+-----+-----+-----+ 300
K I L Y C G I C H S D L H A L K N D W K

AGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTCACGGAGGTGG
301 -----+-----+-----+-----+-----+-----+-----+ 360
N S R Y P M I P G H E I A G E V T E V G

GCAAGAACGTGAGCAAGTTCAAGGCCGCGACCGCGTGGGCGTGGGTGCATGGTGAAC
361 -----+-----+-----+-----+-----+-----+-----+ 420
K N V S K F K A G D R V G V G C M V N S

CGTGCCGGTCTGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCCTGCCCGGGCATGATCC
421 -----+-----+-----+-----+-----+-----+-----+ 480
C R S C E S C D K G F E N H C P G M I L

TCACCTACAACCTCGGTGCGAGCTGCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATGG
481 -----+-----+-----+-----+-----+-----+-----+ 540
T Y N S V D V D G T V T Y G G Y S S M V

TGGTGGTGCACGAGCGGTTCGTGGTCCGGTTCCCCGACGCCATGCCGCTGGACAAGGGCG
541 -----+-----+-----+-----+-----+-----+-----+ 600
V V H E R F V V R F P D A M P L D K G A

CGCCGCTGTGTCGCGCCGCATCACCGTGTACAGCCCCATGAAGTACCACGGGCTCAACG
601 -----+-----+-----+-----+-----+-----+-----+ 660
P L L C A G I T V Y S P M K Y H G L N V

TTCCCGGGCTGCACCTCGGCGTGTGGGGCTGGGCGGGCTGGGCCACGTTGCGGTCAAGT
661 -----+-----+-----+-----+-----+-----+-----+ 720
P G L H L G V L G L G G L G H V A V K F

TCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGAAGAAGGAGG
721 -----+-----+-----+-----+-----+-----+-----+ 780
G K A F G M K V T V I S S S P G K K E E

FIGURE 14

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```

781  AGGCCCTGGGGCGGCTGGGCGCCGACGCGTTTCATCGTCAGCAAGGACGCCGACGAGATGA      840
      -----+-----+-----+-----+-----+-----+
      A L G R L G A D A F I V S K D A D E M K

841  AGGCTGTGATAGCACCATGGATGGCATCANTAAACACGGTATCTGCAAACATCCCCCTGA      900
      -----+-----+-----+-----+-----+-----+
      A V I A P W M A S X N T V S A N I P L T

901  CCCCTCTCTTCGGGCTGCTCAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGA      960
      -----+-----+-----+-----+-----+-----+
      P L F G L L K P N G K M I M V G L P E K

961  AGCCCATCGAGATTTCCTCCCTTCGCTCTAGTTGCCACGAATAAGACCCTGGCCGGGAGCA      1020
      -----+-----+-----+-----+-----+-----+
      P I E I P P F A L V A T N K T L A G S I

1021 TCATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGGCGAAGCACGGCGTGA      1080
      -----+-----+-----+-----+-----+-----+
      I G G M S D T Q E M L D L A A K H G V T

1081 CGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCCTTGGAGCGCCTTGCCA      1140
      -----+-----+-----+-----+-----+-----+
      A D I E V V G A E Y V N T A L E R L A K

1141 AGAACGACGTCAGGTATCGCTTCGTTCATCGACATCGGCAACACCCTCGACAATGTTGCGG      1200
      -----+-----+-----+-----+-----+-----+
      N D V R Y R F V I D I G N T L D N V A A

1201 CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC      1260
      -----+-----+-----+-----+-----+-----+
      T T E *

1261 TCCGTAGTAAACAATAAACGATCAAAACTCTTGTCATCTGGTGCATTGGTGTAGACATGG      1320
      -----+-----+-----+-----+-----+-----+

1321 TTGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAAAAAAAA      1378
      -----+-----+-----+-----+-----+-----+

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FIGURE 14 CONTINUED

FIGURE 15

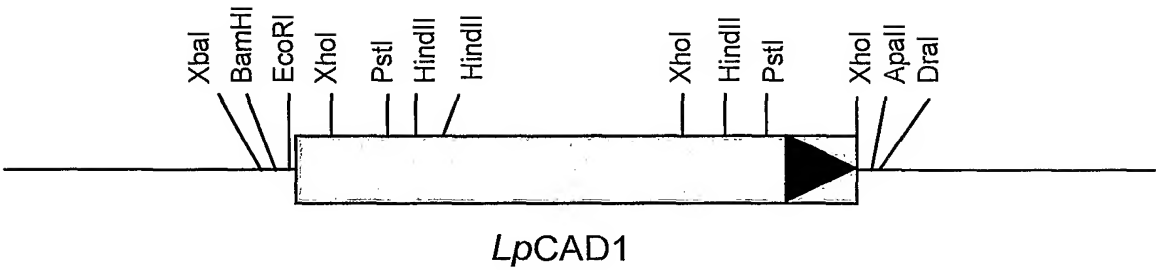


FIGURE 16

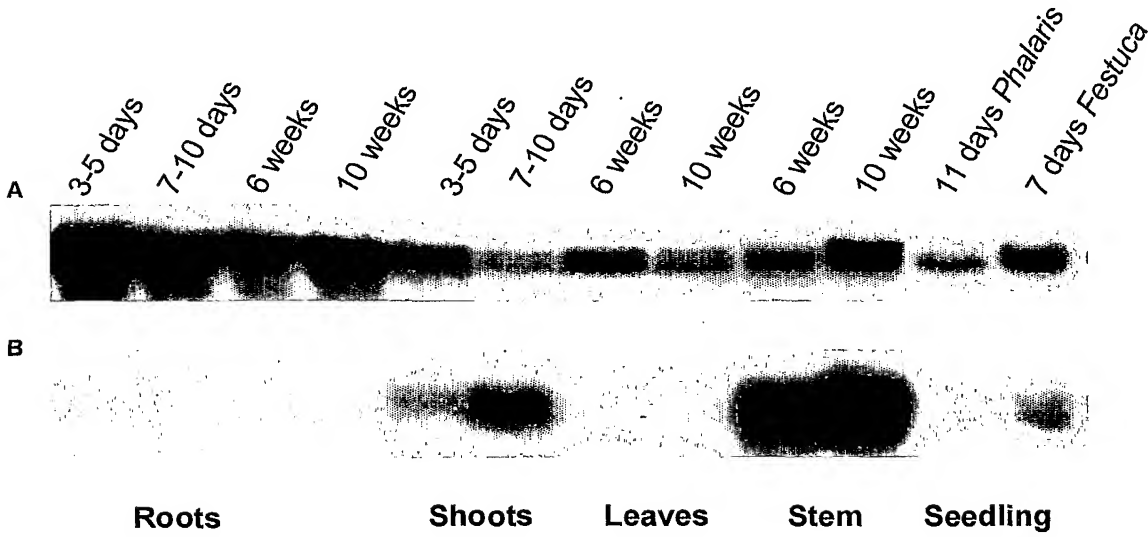
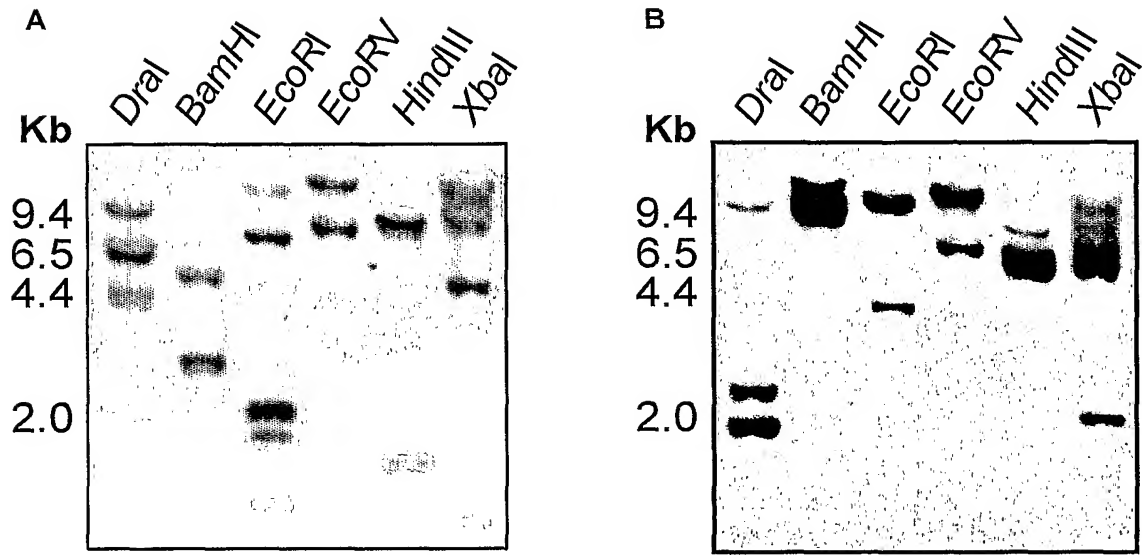


FIGURE 17



[illegible]

FIGURE 18

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```

-3500 AATACTTTTGCACACCCATGCATGGGTGTGGGTGTTCCGTCACCGTCTATGTATTTCTC -3441
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3440 GAAATTCATGCCACCATGGTAGATAAAAAATATTTTTCCTCTCTCTCTTTTATTCAA -3381
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3380 ATCTCAAAGCAtAAkrArTGGTGACAGAACGATAAGATTCCCTACCTAGCTTTCTGAGATC -3321
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3320 CCACTAGTTTATCTTCAAGCTGGTGATTGAAGGATTAACCATGCTTGAATTAGATTGGCT -3261
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3260 TCAAACCTGGTAGTAGCTTGTTCATACTTTGATTACTTTGGTATGGTTAGTTGGTTTGA -3201
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3200 GATTTTGGTCAATGTAGAATCAGATTTGAGAGCGATTGTCAGCTTGAATTGCCGCAGTTT -3141
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3140 TAGCACATACTAGTTTGGATAGATGAACAGTTTGGAGAGACAAATAATGTCTATACGAGC -3081
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3080 TCATCGGATAATATTAGTCTATGGCTTTTGCTTCGGTGTCCCTCTGCAAACCTTACCCC -3021
      +-----+-----+-----+-----+-----+-----+-----+-----+
-3020 TCTGTAGATGGTAGGATTTTCTGATATCCTTTCATGGTTTAAGGGTGTGCGTGTAAAGAA -2961
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2960 CGGGAGATACCGGATCACACCTTTTCGTCTACACTTTACAAGCATGTAACACCTAAGATT -2901
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2900 GATTGATATCTAGGCTTACACCCCAATGGAGGTAACTAATATTATGTAAATGCGACTTT -2841
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2840 TCAAAAGTCCCAATATAACCTTGACGATGATCTTACAAC TACTCGCGCCAGTCTTGTATG -2781
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2780 ATATCAGATTGGCCGAGGATCGTGGGTACCTTTGTAGTGGACTATGATGCTCATGGAGGTT -2721
      +-----+-----+-----+-----+-----+-----+-----+-----+
      KpnI
-2720 GTATGGACATGTGTAAATGCTGGTTTCTCTAGGTTTTTCTAATCAACTTGGCATTCTT -2661
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2660 CTCTTAACACATAATAAGAGGGAATACCTCCATACATTATCTGAAAAAGCATGGCCA -2601
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2600 ACAATGAAACAGAAACAAGTACGACAGTCTATACCCGACCCAAACAATGGCTCAGGTCTT -2541
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2540 TCACGATGCATAGTTTGTTAGCATGTATTTTATAGTAGGAACTAAAAATTAAAGACAAC -2481
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2480 TGCnAAAACAATTTGTCTCTTGAGTGTTTTAAAGGATGCGGCATTTATCGATTATACA -2421
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2420 TTACATATGTGATTGGATtAGCCAACCTTTTGTCTTCCgATGATCATATGAAAGGGTGT -2361
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2360 ATCTTAGGGCATCTCCAATGGGnAGACTCAAATGCAAAAAAATnGTCCGTTTGGGTCTTC -2301
      +-----+-----+-----+-----+-----+-----+-----+-----+
-2300 CnGGACAAAACCTGCTCCCAACGGGGCAACCCAACTTAAAAACGGACAGGTGCAGCGTCC -2241
      +-----+-----+-----+-----+-----+-----+-----+-----+

```

FIGURE 18 CONTINUED

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-2240 GGChTGACCCAAAAC TGACGCAAATTTGGnAnATTTTGGGGCnAGCCAGACGAACGCGG -2181
 +-----+-----+-----+-----+-----+-----+-----+-----
 -2180 GCGTCCACTGTATCCGACTATGTCCGCATCCTGGCCCATCTGACAGTGACACAAAATACA -2121
 +-----+-----+-----+-----+-----+-----+-----+-----
 -2120 ACCACATGCGCCCCCACCCTTCTCTCTCCTCCGTCGCTTTTCCCATGGAA nCnGTCC -2061
 +-----+-----+-----+-----+-----+-----+-----+-----
 -2060 TCGCTCCTCGCCGAATTGATCTCGCCTAACCATGCTCCGCCGCCACCcTCGcCTkAAGG -2001
 +-----+-----+-----+-----+-----+-----+-----+-----
 -2000 CCCCAgCCGCCGCTACcTCC'TTTTGTGTCAGCCCTAT'TgGAAGTCGCCGgAGTTGAAACGA -1941
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1940 GCGCCGCCAGCCTcGACACCGCCGAGCAAGACGAAGACTGGGGCGGAGCTCGCCGAGACGG -1881
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1880 GACGGGGACGGAGCTCGCCATGCGTGCCTCGCAGGGGCGCGATGGGGGCGGAGCTCGCCG -1821
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1820 *Pst.I*
 TGGCTGGCTGCGAGCACCTCGGGCCGCTGCTAGCCGTGCCACGACGCGAGCATGCGCCTCG -1761
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1760 ACGCCGCCCCGTGCTACCTCGTCGCGCGCCAGGGCCGCCCCGCCCCGCGGACCGGcGcGg -1701
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1700 CGgAgACGCGAcCTTCGCGgACGTGCCCGGCGGCAGAGACGCGTCCTTCGCGACAGCGCC -1641
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1640 CTCCTCGATCTCCGTGAGCGCATACGCgGcTAgGAgGGACGCGGGCGTCCCCGGTGTC -1581
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1580 GGCCTCCGTGTGTGGCGCATCGCGGGCGCGGCTCCGTGAGGcGCATCGCGGGCGTGGCC -1521
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1520 TCGTGGCGCAGCCTGCCCTGATTGCGTCTGAGGCGCGGCGCGGAGCTTCCTCGCGGCGGC -1461
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1460 GCGGGCGGAGCCTCCTCGCTGCGGCGCGACCTGCTCTGCCGCGGTCCGAGACGCGGCGCG -1401
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1400 GGCAGAGCTTCCTCGCGGCGGCTCGGGCGCGGCTTCCTCGCGGCGATGGCGCTTCAGGC -1341
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1340 TCGCACGCGGCTCCGGCGTGGCGCAGCGAGAGCGCAGCTCCGGTGAGTTAGGCACAGG -1281
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1280 CGCGACACGACATCCCCGGCCTCGGCCTCCGGCGTGGCGCAGCGCGAGCGCGAGCTAGCC -1221
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1220 TAGGTGGCAACTAGTAcTACGAGGAAGAAAGAGGAGAAACAATTATTTGGGTACACGCG -1161
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1160 TTGGGCGTACTGTGCGATCCAAACGGACACCCgGACGCGAaACGATGTCAGCGTGTCCGC -1101
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1100 GTGGcGACCCAAACGACCCGAAACGGACGTCCGT'TTGGGTGCGTTCGTTGGAGATGCCCT -1041
 +-----+-----+-----+-----+-----+-----+-----+-----
 -1040 TACTCCCCATCCTCAAATGAGTCTAATTATATATCTTGTTGTAAGTTTAAAAAAGTTAA -981
 +-----+-----+-----+-----+-----+-----+-----+-----

FIGURE 18 CONTINUED

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-980 ACTTTGATCAACATTAGTAATGATAGTAGCAACGAATACAAAATTAAATTGTAAAAATAT -921
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -920 ATTATGAAACTTTATTTTAAGATGGATCTAGTTATACTAATTTTCTGCGGATGGAGGAAG -861
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -860 TAGCTAAATATTGTTAATTTCTAAATAAAAAATTAAACTTTAACTTAAACAAAAGTTA -801
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -800 CAAGCATAATTATCTGtGGATGGAGGAAGtAGCTAAGATACACCAATCCTCTCTCTACAT -741
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -740 TACCTAGCATGCCACATCAGGAAACTATTTAGGATAAGCTCCAAGGAACCACCCAGAACA -681
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -680 ACAATTTACATGGCCTGGCTACCTAATGACAATTTCCGAGCAACTGGTGGTGGTGGTAC -621
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -620 GCGTTCCTTGTTCAATTGTCTCTATTACAAGAGTGGCCCTGTATAGGTAAAAAAAATAA -561
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -560 *HindIII* CAAGCTTCCAAGGACGGCCATGTTCTTGTTCTGCAGGCTGCACGTACTCACGACGAAG *PstI* -501
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -500 TGTATCTCGTGTCTGGACATTTGTCTCGCGCATTTTGTAAACATGAAATTAAAAATGTG -441
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -440 GTGGCCTGCTATATCTGTATGGGGGTATCATGCACTCCTTCGCAGAGGAATCCAGACGAC -381
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -380 GATTTACACGTGTTTCCACCTTAGCTTTTTTAAGTGTGTGTGAAGGAACGATCATATA -321
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -320 ACTGCCCTGAATGCTGCATATATATAAACCGACTCCATCATGTACTCGAGACAAGGTCG *XhoI* -261
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -260 TCAAGAAAAACAACTATGCCTATCTCACTAGCAATGATTTGAGAGTACAGCTTTTCCGG -201
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -200 TGCCATATTTTTCCTATATATCTTTTCTGAAGAACAAGAAAAAAAACAGTTGGTGT -141
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -140 GGTGGTTGGTGAAGCGAGAAAGCCCCATATAAGCCCTGCTCACCTCCCCGCAAAGCACA -81
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -80 ACTCATAGCTCGGGTCTCTCGCTCACACCAAAATCGCCCACCAGCACCAGCATCTCTCGA *PvuI* -21
 +-----+-----+-----+-----+-----+-----+-----+-----+
 -20 TCGGCAGACGCATAGATCGATGGGCTCCACCGCCCGACATGGCCGCGTCCGCGGACGA 39
 +-----+-----+-----+-----+-----+-----+-----+-----+
 40 GGACGCGTGCATGTTTCGCCCTCCAGCTCGCTTCTCGTGGTCCCTCCCGATGACGCTGAA 99
 +-----+-----+-----+-----+-----+-----+-----+-----+
 100 GAACGCCATCGAGCTTGGCCTCCTGGAGATCCTGGTGGCCGCGCGGCAAGTCGCTGAC 159
 +-----+-----+-----+-----+-----+-----+-----+-----+
 160 CCCGACCGAGGTGGCCGCAAGCTCCCGTCCGCGGCGAACC CGGAAGCGCGGACATGGT 219
 +-----+-----+-----+-----+-----+-----+-----+-----+
 P T E V A A K L P S A A N P E A P D M V

FIGURE 18 CONTINUED

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```

220  GGACCGCATACTCCGGCTGCTCGCGTCGTACAACGTCGTGACGTGCCTGGTGGAGGAGGG 279
      +-----+-----+-----+-----+-----+-----+
      D R I L R L L A S Y N V V T C L V E E G

280  CAAGGACGGCCGCCTCTCCCGGAGCTACGGCGCCGCGCCCGTGTGCAAGTTCCTCACCCC 339
      +-----+-----+-----+-----+-----+-----+
      K D G R L S R S Y G A A P V C K F L T P

340  CAACGAGGACGGCGCTCTCCATGGCGCGCTCGCGCTCATGAACCAGGACAAGGTCCTCAT 399
      +-----+-----+-----+-----+-----+-----+
      N E D G V S M A A L A L M N Q D K V L M

      Intron/exon boundary
400  GGAGAGCTG↓GTGAGTCTCTCAGTGGAGCTAGTTACTGTAGATCCGAATTCGTTCCCTTTA 459
      +-----+-----+-----+-----+-----+-----+
      E S

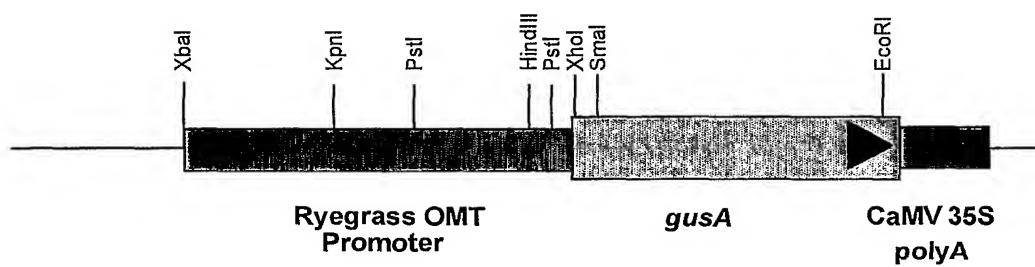
      SalI pBluescript
460  GTGAGGGTTAATTCGCGGCCGCTCGACCTCGAGGGGGGGCCCGGTACCCAATTCGCCC
      +-----+-----+-----+-----+
      TATAGTGAGTCGTATTACGCGCGCTCACTGGCCGTCGTTTACAACGTCGTGACTGGGAA
      AACCTGGCGTTACCCAACCTAATCGCCTTGCAGCACATCCCCCTTTCGCCAGCTGGCGT
      AATAGCGAAGAGGCCCGACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAA
      TGGGACGCGCCCTGTAGCGGCGCATTAAGCGCGCGGGTGTGTG 744

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FIGURE 18 CONTINUED

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FIGURE 19



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FIGURE 20

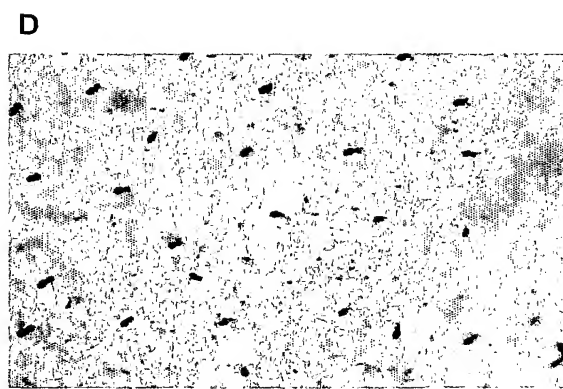
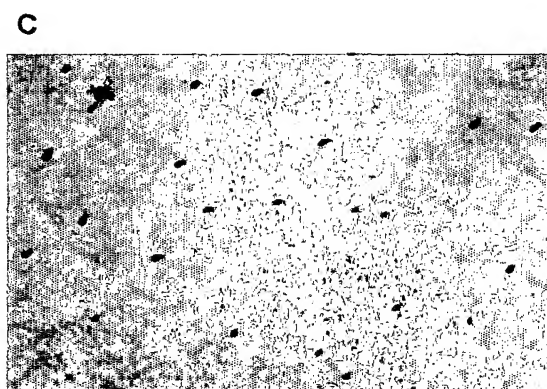
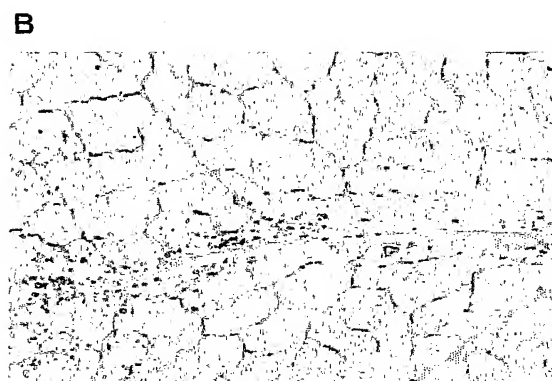
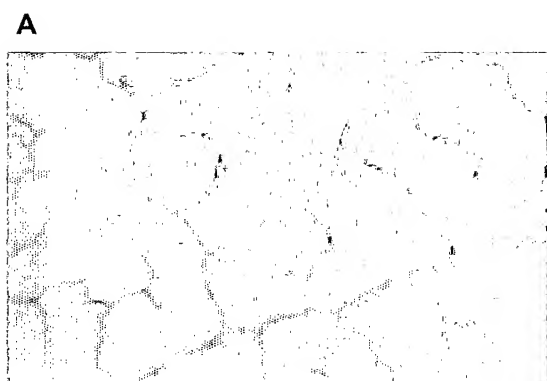


FIGURE 21

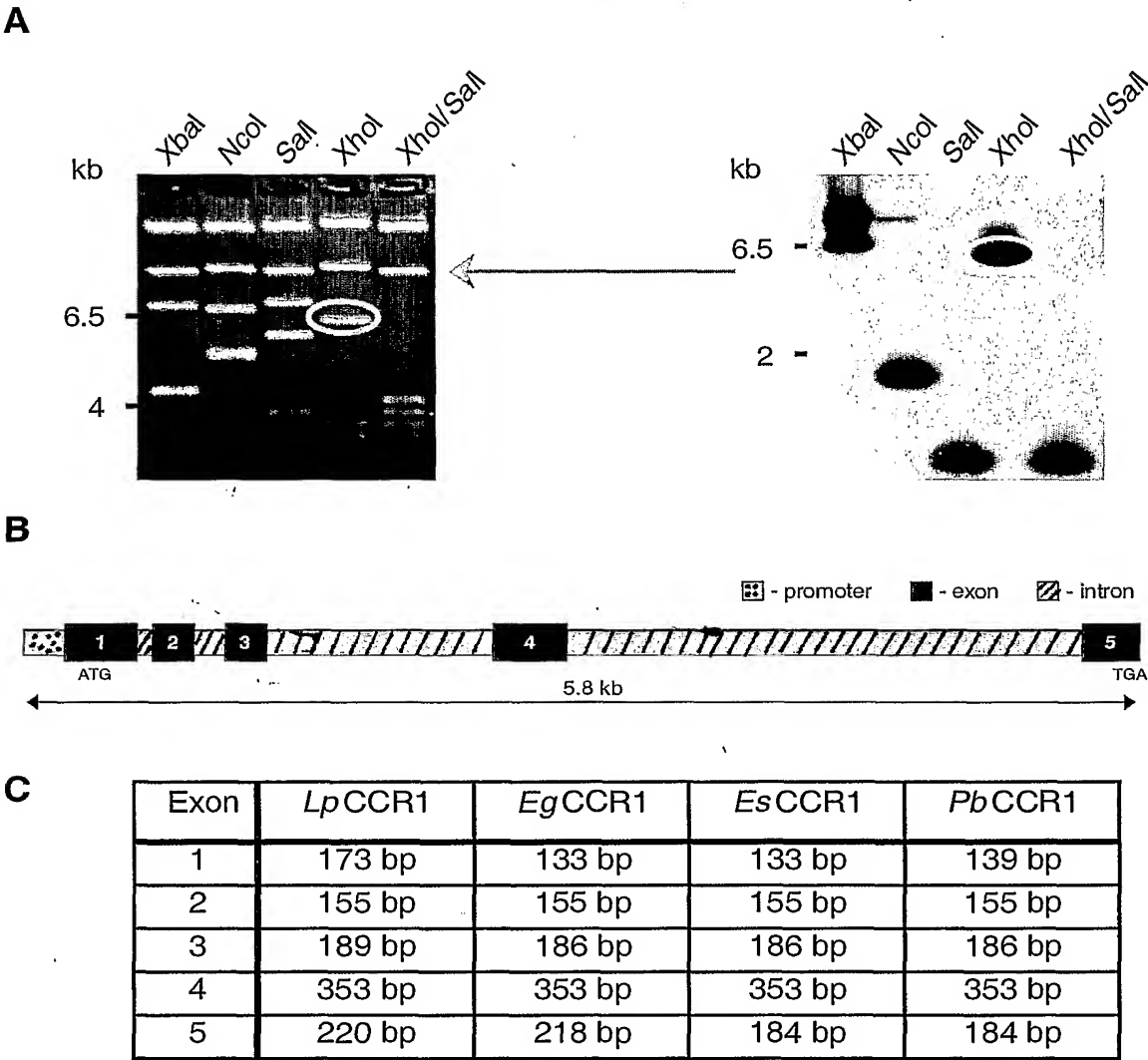
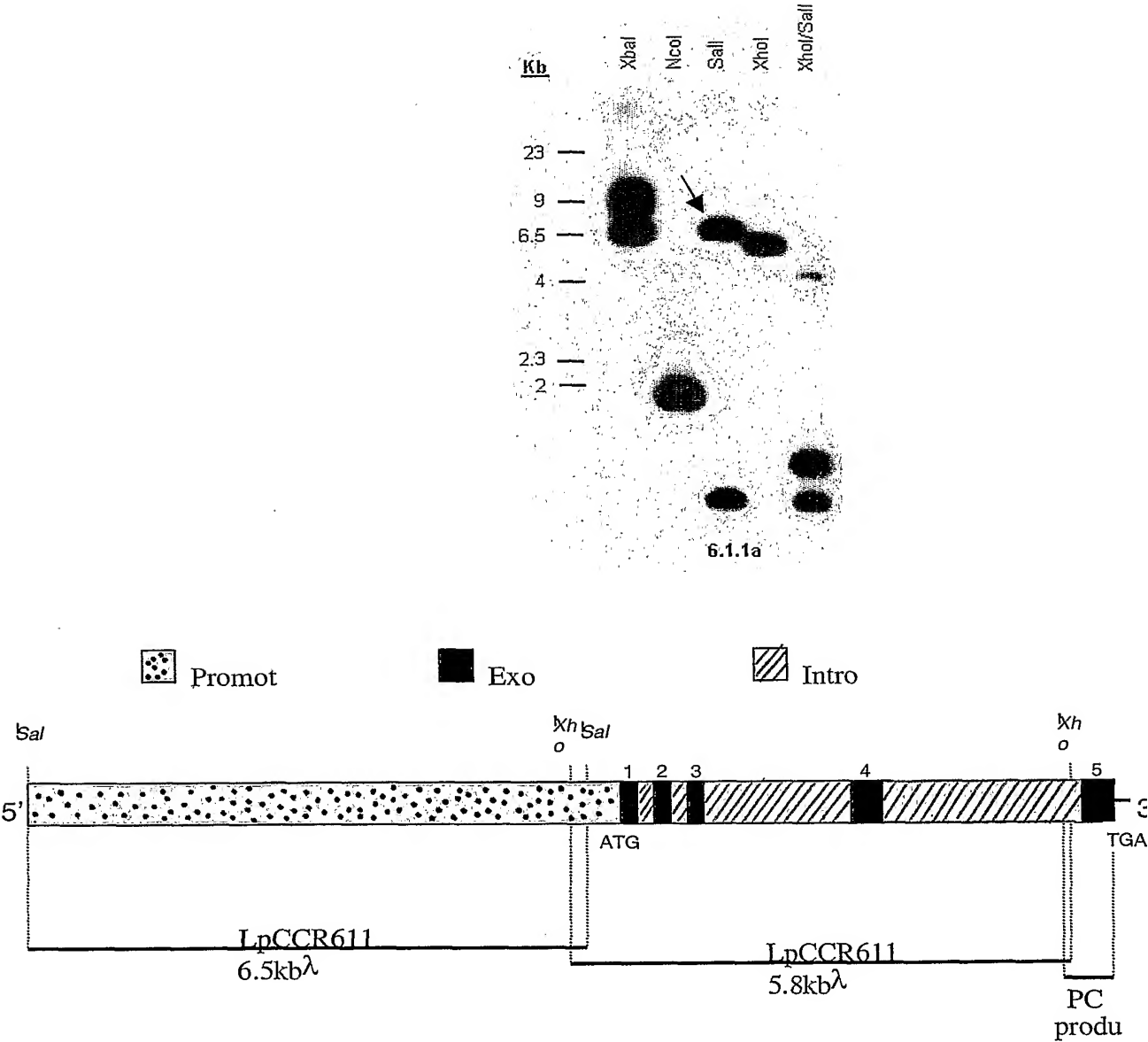


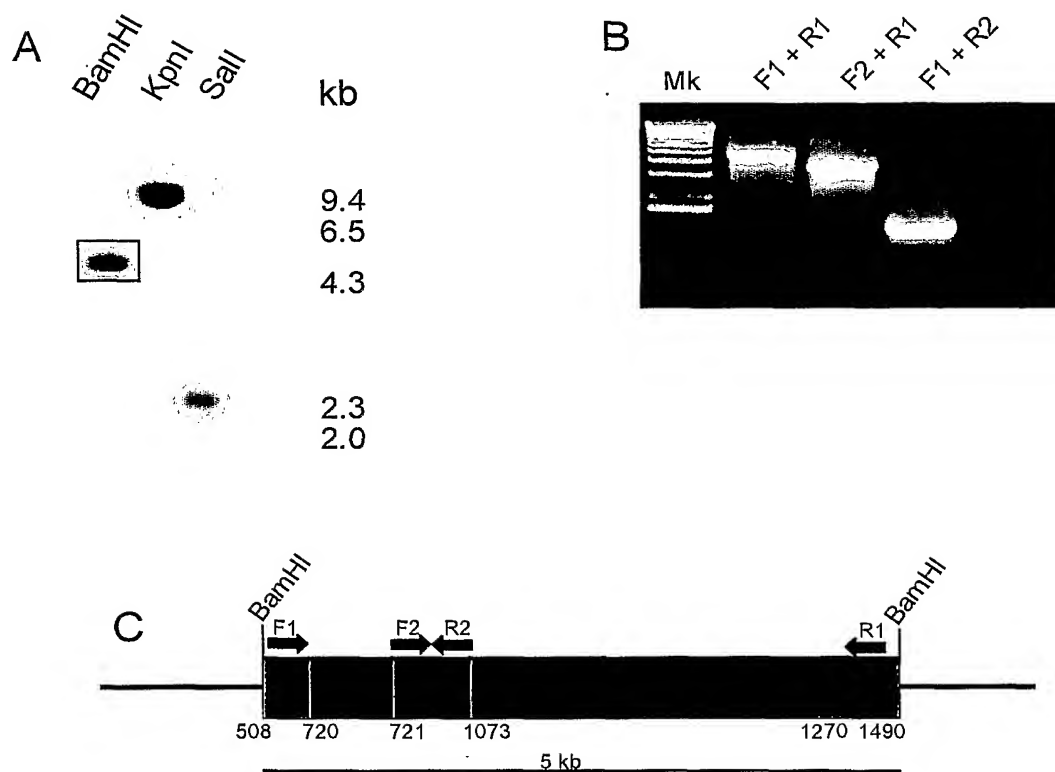
FIGURE 22

A



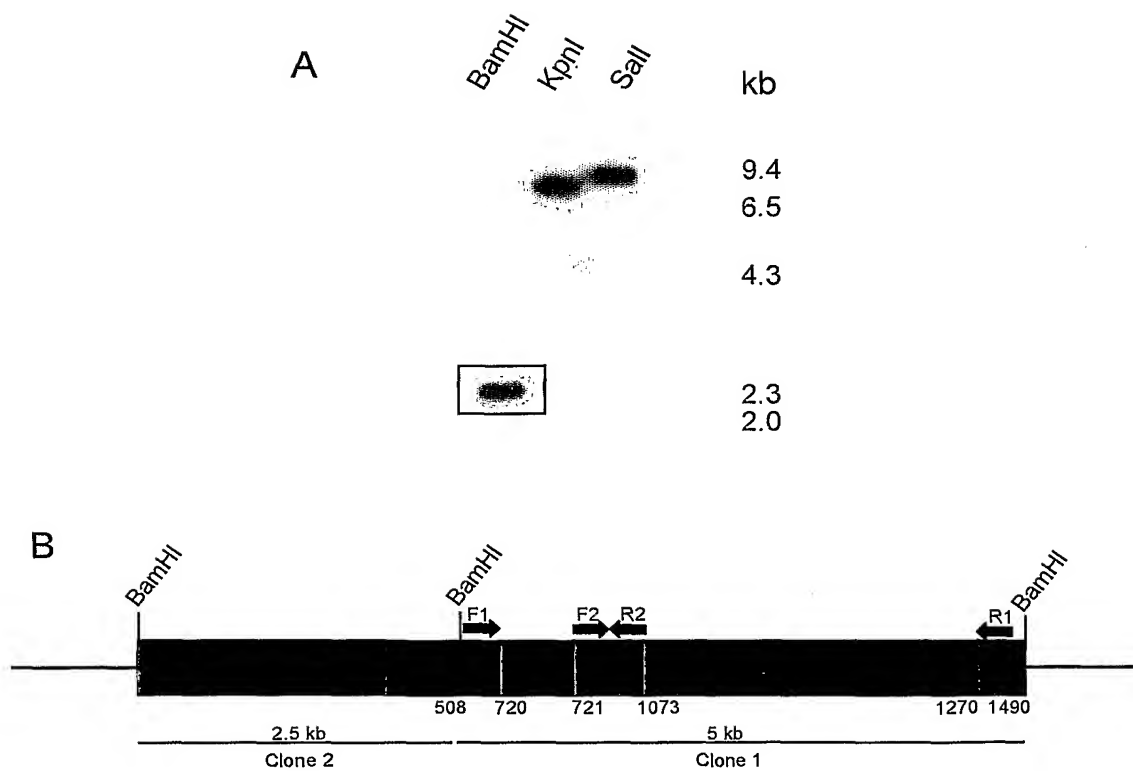
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FIGURE 23



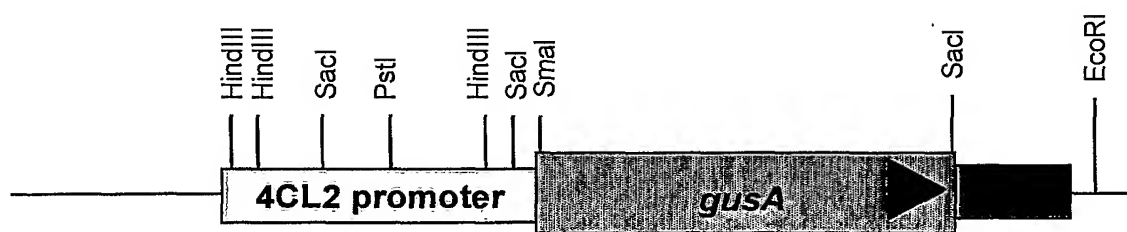
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FIGURE 24



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FIGURE 25



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```

1  TCCCGTATCTTCAACGTGACACCCCTACACTTCCTGCTTGTCTTGGAGATTACACACAC      60
   -----+-----+-----+-----+-----+-----+
61  ACGGCAATTACCAGGAGTATCTTCCTAGATTATTTTTCGATAAGGATCTTCCAGATAT      120
   -----+-----+-----+-----+-----+-----+
121 AGCATGTGAATCTCTGTACTACTACTGTTTGTCAAGCAAAATTAACATTGACATCAGTGT      180
   -----+-----+-----+-----+-----+-----+
181 TTTTGTGTGGGGGCAGCGGAATCTTTGACGCCTCTTCTTGCCTCTCAAGACATGTCACCCCT      240
   -----+-----+-----+-----+-----+-----+
241 CACTAGTTAGTGTGCCAGCTGGTAGTACTACGTACGATGCTCCCTCCCTCCGTAATTATT      300
   -----+-----+-----+-----+-----+-----+
301 CAACCTTTTGTCTCTCTCTTTTATAAAGTCAAACCTTTTAAATCTGACCAGATATCTGC      360
   -----+-----+-----+-----+-----+-----+
361 TAAAAAATTAGCAGACATGCATACATCAAAGCAGTAGTCCCTCCCTCCGTTTAAAAATTACC      420
   -----+-----+-----+-----+-----+-----+
421 TGGGTTTATTCAAATAAAGTCAAACCTCTGTAAATTCAAATTAATATTTAGAAAAATCTA      480
   -----+-----+-----+-----+-----+-----+
481 ACAGCACCTGTAGTATAAAAGTATGCTCCCTCTGTTTGTAAAAAGCTAAGCAACTTTTT      540
   -----+-----+-----+-----+-----+-----+
541 TGAGATACGGATAAAATCTTTAGCTAAAAATGTCTATATACCTTTGTATCTAGATAAAGT      600
   -----+-----+-----+-----+-----+-----+
601 TGGAAAGCTTTTGTAGAAACAGACAAAGTATGTGTTTGACATTATGAATGTTGAGTATTT      660
   -----+-----+-----+-----+-----+-----+
661 TTCCTCTAATCTTGATCAAATTTTACAAATTTTGGCTTGAATAGAGGGACCATTTAGT      720
   -----+-----+-----+-----+-----+-----+
721 ATGAAACTACATAAAATTTGTAAACACTCAACATAATTTACGATGGGTCAGTGATAGCAC      780
   -----+-----+-----+-----+-----+-----+
781 TAACTTAGCTTTTCATAAATGCCACTGCTTTTCAATAGAGCATGAAGCAGGACAAATTTA      840
   -----+-----+-----+-----+-----+-----+
841 TTCGTGTGACTTGAATAGAGGGAGCCTGTTCTGGTTCAACTCACCTGCATGTGTGTCTT      900
   -----+-----+-----+-----+-----+-----+
901 CATCCCTTTTGTCTTTCCTATCTGTGGTGTCAATTGAGTGTCCACGTGCATGTGGGCCGA      960
   -----+-----+-----+-----+-----+-----+

```

FIGURE 26

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961 AACTTGAACCTAGAAATTGACATGCTCCCACTGCCCCGAGCGGAGTATCTTTGTGCTTTG 1020
-----+-----+-----+-----+-----+-----+-----+
1021 TTACCCCTTATTGTTGCTACGTACTACAGTGTTTAGATTGGAACCTCATAATCAAAAGAAC 1080
-----+-----+-----+-----+-----+-----+-----+
1081 TTAGTTTCCTACAATTTTTTGCTAAGCAATATAATGAGCAATCAAACCTCTATATCTGTG 1140
-----+-----+-----+-----+-----+-----+-----+
1141 GCAAATAACTAATCCATTATAGTTACAGTTTAGATGCAGACGCCAGTGTTTCTTCCCTT 1200
-----+-----+-----+-----+-----+-----+-----+
1201 TTCGGAAGGAGCTATTCCATAATAAGTGTTGGAAATTTAATAAATGGGTACTACGAATT 1260
-----+-----+-----+-----+-----+-----+-----+
1261 TGAAAAAGGAGTGTCAAAAATCACTAAGAAAGTACGTAGTACAAATTTAACTAAGAT 1320
-----+-----+-----+-----+-----+-----+-----+
1321 TCCGACACTTATTAGGATCGGAGAGAGTAAGTAGCAAACCTACTCTCCATCCACCTAAAA 1380
-----+-----+-----+-----+-----+-----+-----+
1381 CACGTGATTTAACTTTGTCTAGATACGGATAGAAAGTTGGGATACATCCGTATCTTAAAA 1440
-----+-----+-----+-----+-----+-----+-----+
1441 AAAAAGGAGCTTATTTTAGACGAAGGAGGGAGTATTTC AACCTTGATTTTAAACGGAATC 1500
-----+-----+-----+-----+-----+-----+-----+
1501 TACAAAGGGAATACATGGATTGTACAAGTGGGCTGACCGTATCCATTATGTACTCGTACT 1560
-----+-----+-----+-----+-----+-----+-----+
1561 TTGCAGTTTGAAAGCAAAGGCTAGTGTAATTTGTAGGTGGTTCTAGGCGTCTAGCTGTTT 1620
-----+-----+-----+-----+-----+-----+-----+
1621 CATGGCGTTATCACAGCCGTGCCAGTGTGCTCAGGGCCGTACATAAGTTGCTTGGTGTAT 1680
-----+-----+-----+-----+-----+-----+-----+
1681 GTGTCGATCTAGGATTTGCCGTCTTACAATTTTGCTTTCCAACCTATTTTCTGTAAAGAG 1740
-----+-----+-----+-----+-----+-----+-----+
1741 ATCGATGTGAACCTTCTCTGTCGAGTAAACTGAAATTGCTCTGAATAAATATAACTCGGCAG 1800
-----+-----+-----+-----+-----+-----+-----+
1801 ATTATGTTTATCGTTTGCATGCGTAACAGGCTACACAAATTGCTCGAGTCAGCAGCGAG 1860
-----+-----+-----+-----+-----+-----+-----+
1861 TTGAGCTCACAACGAATCCATCAGCAAAAATACTATACTATAGTAGCACATCGTTTCTTT 1920
-----+-----+-----+-----+-----+-----+-----+

FIGURE 26 CONTINUED

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1921 TTTTCATGACGTTTCTGTTTCCTCCTAACTTTCCAGGAGCACC GGAGACGACGATGTGGTG 1980
-----+-----+-----+-----+-----+-----+
R S T G D D D V V

1981 ATAAAGATTTTGTACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGG 2040
-----+-----+-----+-----+-----+-----+
I K I L Y C G I C H S D L H A L K N D W

2041 AAGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTCACGGAGGTG 2100
-----+-----+-----+-----+-----+-----+
K N S R Y P M I P G H E I A G E V T E V

2101 GGCAAGAACGTGAGCAAGTTCAAGGCCGGCGACCCGCTGGGCGTCGGGTGCATGGTGAAC 2160
-----+-----+-----+-----+-----+-----+
G K N V S K F K A G D R V G V G C M V N

2161 TCGTGCCGGTCTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACCTGCCCCGGGCATGATC 2220
-----+-----+-----+-----+-----+-----+
S C R S C E S C D K G F E N H C P G M I

2221 CTCACCTACAACCTCGGTGCGACGTCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATG 2280
-----+-----+-----+-----+-----+-----+
L T Y N S V D V D G T V T Y G G Y S S M

2281 GTGGTGGTGCACGAGCGGTTCTGTTCCGGTTCCCCGACGCCATGCCGCTGGACAAGGGC 2340
-----+-----+-----+-----+-----+-----+
V V V H E R F V V R F P D A M P L D K G

2341 GCGCCGCTGCTGTGCGCCGGCATACCGTGTACAGCCCCATGAAGTACCACGGGCTCAAC 2400
-----+-----+-----+-----+-----+-----+
A P L L C A G I T V Y S P M K Y H G L N

2401 GTTCCCCGGGCTGCACCTCGGCGTGCTGGGGCTGGGCGGGCTGGGCCACGTTGCGGTCAAG 2460
-----+-----+-----+-----+-----+-----+
V P G L H L G V L G L G G L G H V A V K

2461 TTCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGGAAGAAGGAG 2520
-----+-----+-----+-----+-----+-----+
F G K A F G M K V T V I S S S P G K K E

2521 GAGGCCCTGGGGCGGCTGGGCGCCGACGCGTTTCATCGTCAGCAAGGACGCCGACGAGATG 2580
-----+-----+-----+-----+-----+-----+
E A L G R L G A D A F I V S K D A D E M

2581 AAGGTAGGCGGACCCGCTGGTTTCAGGTTACTTCCCTGTCCGGTGCAGAAGAAAGAGGAA 2640
-----+-----+-----+-----+-----+-----+
K

FIGURE 26 CONTINUED

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G at 851 bp (coding sequence) missing from cDNA in cv Ellett

▼

```

2641  CTTGAGGGTTCATGTTTGTGTTTGGCGTTGGTGATGTCTTTCAGGCTGTGATGAGCACCAT
-----+-----+-----+-----+-----+-----+-----+
                                         A V M S T M
2700

2701  GGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGACCCCTCTCTTCGGGCTGCT
-----+-----+-----+-----+-----+-----+-----+
      D G I I N T V S A N I P L T P L F G' L L
2760

2761  CAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGAAGCCCATCGAGATTCTCTCC
-----+-----+-----+-----+-----+-----+-----+
      K P N G K M I M V G L P E K P I E I P P
2820

2821  CTTGCTCTAGTTGCCAGTAAGTCTTAGGATCTCTTGCAATAAGGAGAAATCATGCACTG
-----+-----+-----+-----+-----+-----+-----+
      F A L V A
2880

2881  ATCGATCAGAGAAATGAGATAGCATCCTGATGAACATTGTACGTGTGTGCAGCGAATAAG
-----+-----+-----+-----+-----+-----+-----+
                                         N K
2940

2941  ACCCTGGCCGGGAGCATCATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCG
-----+-----+-----+-----+-----+-----+-----+
      T L A G S I I G G M S D T Q E M L D L A
3000

3001  GCGAAGCACGGCGTGACGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCC
-----+-----+-----+-----+-----+-----+-----+
      A K H G V T A D I E V V G A E Y V N T A
3060

3061  TTGGAGCGCCTTGCCAAGAACGACGTCAGGTATCGCTTCGTCATCGACATCGGCAACACC
-----+-----+-----+-----+-----+-----+-----+
      L E R L A K N D V R Y R F V I D I G N T
3120

3121  CTCGACAAGGTTGCGGCCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTT
-----+-----+-----+-----+-----+-----+-----+
      L D K V A A T T E *
3180

3181  GTTCCACTGTTAGTGCTCCGTAGTAAACAATAAACGATCAAACTCTTGTATCTTGGTGC
-----+-----+-----+-----+-----+-----+-----+
3240

3241  ATTGGTGTAGACATGGTTGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAGTTTG
-----+-----+-----+-----+-----+-----+-----+
3300

3301  CTTCTTGCCGTGTTAATGGATTACCTACTTAGCTTCACTGCAATTAACAAATTAAGAAAC
-----+-----+-----+-----+-----+-----+-----+
3360

3361  GACACACCCAAAAGACTTTTCGTCAGTTTCTTGGAATATACAAGTCGTTATGGTTGGGTG
-----+-----+-----+-----+-----+-----+-----+
3420

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FIGURE 26 CONTINUED

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3421	TCAGTGTGTCACAGATAATCATACTATGGTATTTAACCTGGAAGATCGTTTTTTTGGCGG	3480
3481	CAACTCAGTGGGTTTTCCCACTATGTATATTTATAAATATTCAACAAGTCATGAGGTACA	3540
3541	AAGGGTTGTTGCTAGAGGATAGCAACAAGAAGCTAGCCAAAAGATCATAGGCTTAAAAA	3600
3601	GAGAGAAAAGAAAACAAAACCTGCTATAGTTATCGAAATCTCTCAGCTCAAATTTTAAAC	3660
3661	CAGCATAAGACTTTCTAGAAGCCTTATGAACAAGAAGAGCTAGCTCATCTTTAAACCTTT	3720
3721	TCCTGCATCTGTAAAGATTGAGGGTGCAACCCTTGAATATAAAATCATTCCTGTATCCA	3780
3781	GATAGACTATGTAGTCAAAATAGTCATTTCCATGAAGAAGGGCACTTTAAATACATTTT	3840
3841	GAGACTTGGTATGATACTCTGAATGTCAACACCCTGGAAGATCTTTTCACTCCTATGGAA	3900
3901	GGACAAGAAAGCATTTCAACTCCTTTTACTAAGGAAGAGATTGACAAGGTGATTCAGAGA	3960
3961	ATTCTTTTAGACACTATAGAAAGTCACAAGGTGCCAACGGCGCAATCCTGTGCCGACGGC	4020
4021	TTTTTATCGGGGAAGCCAGCATCGGTACCGAGACCGGCAGCCACCAACTAGGCCGTCGG	4080
4081	CACACATCCTCCAGTGTGCGCGGCCAACATCGGCATAAGTTGGCCCGTTGGGCATCAACT	4140
4141	CCCCGTCGGAACAGGTCTAGCGCATGGACCGTCGTGATGGCGGCGGCAACGACGTCATC	4200
4201	CTATGCCGACGGCCTAGCCGTCGGCCTAGCTTGCCAGCGCTATGCCGACGTCACATTGCC	4260
4261	ATCGGCACATGCTAGTTTTTTTTTCTTTTTTCTACATGCCAAATTGTATATGTATATATA	4320
4321	CTCATTTACTTATTACTTCCAATTATTTTAATGTGTATATATTTTGCTCACCAATTGTAC	4380

FIGURE 26 CONTINUED

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GAATTTGTACCCTCCGAGAAATTGCTAAAATGATGGAGTGACCTACAACGAGCCTTGGAT
4381 -----+-----+-----+-----+-----+-----+-----+ 4440

ATGTGAGTTCTTCTTGCCCCATTGCACAAAAATTGTAAATATTAGGGTTTACTGGATCCA
4441 -----+-----+-----+-----+-----+-----+-----+ 4500

A) CTAGTTCTAGAGCGGCCGCCACCGGGGAGCTCCAGCTTTTGTTCCTTTAGTA
-----+-----+-----+-----+-----+-----+-----+ 4555

FIGURE 26 CONTINUED

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1 GGCACGAGTCGCCTCCAACGTCTTCCCTTAACCGGCCGTCCCTACGCTTGCACCACCACC 60
-----+-----+-----+-----+-----+-----+-----+
61 ACGCACAGACAGAGCAGTTTCCCAGCCCCCGCCGGAACCGGATGGCACCCACGGCGGCGG 120
-----+-----+-----+-----+-----+-----+-----+
M A P T A A E
121 AGCAGACGGAGCACCACCAGCACACCAGGAAGGCGGTGGGGCTGGCGGCGCGACGACG 180
-----+-----+-----+-----+-----+-----+-----+
Q T E H H Q H T R K A V G L A A R D D A
181 CCGGCCACCTCTCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA 240
-----+-----+-----+-----+-----+-----+-----+
G H L S P L A I T R R S T G D D D V V I
241 TAAAGATTTTGTACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA 300
-----+-----+-----+-----+-----+-----+-----+
K I L Y C G I C H S D L H A L K N D W K
301 AGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTCACGGAGGTGG 360
-----+-----+-----+-----+-----+-----+-----+
N S R Y P M I P G H E I A G E V T E V G
361 GCAAGAACGTGAGCAAGTTCAAGCCGGCGACCGCGTGGGCGTGGGTGCATGGTGAAC 420
-----+-----+-----+-----+-----+-----+-----+
K N V S K F K A G D R V G V G C M V N S
421 CGTGCCGGTGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCAC 480
-----+-----+-----+-----+-----+-----+-----+
C R S C E S C D K G F E N H C P G M I L
481 TCACCTACAAC 540
-----+-----+-----+-----+-----+-----+-----+
T Y N S V D V D G T V T Y G G Y S S M V
541 TGGTGGTGACGAGCGGTTCGTGGTCCGGTTCCCCGACGCCATGCCGCTGGACAAGGGCG 600
-----+-----+-----+-----+-----+-----+-----+
V V H E R F V V R F P D A M P L D K G A
601 CGCCGCTGCTGTGCGCCGGCATCACCGTG 660
-----+-----+-----+-----+-----+-----+-----+
P L L C A G I T V Y S P M K Y H G L N V
661 TTCCCGGGCTGCACCTCGGCGTGCTGGGGCTGGGCGGGCTGGGCCACGTTGCGGTCAAGT 720
-----+-----+-----+-----+-----+-----+-----+
P G L H L G V L G L G G L G H V A V K F
721 TCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGAAGAAGGAGG 780
-----+-----+-----+-----+-----+-----+-----+
G K A F G M K V T V I S S S P G K K E E

FIGURE 27

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AGGCCCTGGGGCGGCTGGGCGCCGACGCGTTCATCGTCAGCAAGGACGCCGACGAGATGA
 781 -----+-----+-----+-----+-----+-----+-----+ 840
 A L G R L G A D A F I V S K D A D E M K
 G at 851 bp (coding sequence) missing from cDNA in cv Ellett
 ▼
 AGGCTGTGATGAGCACCATGGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGA
 841 -----+-----+-----+-----+-----+-----+-----+ 900
 A V M S T M D G I I N T V S A N I P L T
 CCCCCTCTCTTCGGGCTGCTCAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGA
 901 -----+-----+-----+-----+-----+-----+-----+ 960
 P L F G L L K P N G K M I M V G L P E K
 AGCCCATCGAGATTCTCCCTTCGCTCTAGTTGCCACGAATAAGACCCTGGCCGGGAGCA
 961 -----+-----+-----+-----+-----+-----+-----+ 1020
 P I E I P P F A L V A T N K T L A G S I
 TCATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGGCGAAGCACGGCGTGA
 1021 -----+-----+-----+-----+-----+-----+-----+ 1080
 I G G M S D T Q E M L D L A A K H G V T
 CGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCCTTGGAGCGCCTTGCCA
 1081 -----+-----+-----+-----+-----+-----+-----+ 1140
 A D I E V V G A E Y V N T A L E R L A K
 AGAACGACGTCAGGTATCGCTTCGTCATCGACATCGGCAACACCCTCGACAATGTTGCGG
 1141 -----+-----+-----+-----+-----+-----+-----+ 1200
 N D V R Y R F V I D I G N T L D N V A A
 CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC
 1201 -----+-----+-----+-----+-----+-----+-----+ 1260
 T T E *
 TCCGTAGTAAACAATAAACGATCAAACTCTTGTCATCTGGTGCAATTGGTGTAGACATGG
 1261 -----+-----+-----+-----+-----+-----+-----+ 1320
 TTGTTTGGCGAGGAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAAAAAAAA
 A) -----+-----+-----+-----+-----+-----+-----+ 1378
 GGCACGAGTCGCCTCCAACGTCTTCCCTTAACCGGCCGTCCCTACGCtTGCACCACCACC
 1 -----+-----+-----+-----+-----+-----+-----+ 60
 ACGCACAGACAGAGCAGTTTCCCAGCCCCCGCCGGAACCGGATGGCACCACGGCGGCGG
 61 -----+-----+-----+-----+-----+-----+-----+ 120
 M A P T A A E
 AGCAGACGGAGCACCACCAGCACACCAGGAAGGCGGTGGGGCTGGCGGCGCGCGACGACG
 121 -----+-----+-----+-----+-----+-----+-----+ 180
 Q T E H H Q H T R K A V G L A A R D D A

FIGURE 27 CONTINUED

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CCGGCCACCTCTCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA
181 -----+-----+-----+-----+-----+-----+-----+ 240
G H L S P L A I T R R S T G D D D V V I

TAAAGATTTTGTACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA
241 -----+-----+-----+-----+-----+-----+-----+ 300
K I L Y C G I C H S D L H A L K N D W K

AGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTCACGGAGGTGG
301 -----+-----+-----+-----+-----+-----+-----+ 360
N S R Y P M I P G H E I A G E V T E V G

GCAAGAACGTGAGCAAGTTCAAGGCCGGCGACCGCTGGGCGTCGGGTGCATGGTGAAC
361 -----+-----+-----+-----+-----+-----+-----+ 420
K N V S K F K A G D R V G V G C M V N S

CGTGCCGGTCTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACCTGCCCGGGCATGATCC
421 -----+-----+-----+-----+-----+-----+-----+ 480
C R S C E S C D K G F E N H C P G M I L

TCACCTACAACCTCGGTGCGAGCTGCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATGG
481 -----+-----+-----+-----+-----+-----+-----+ 540
T Y N S V D V D G T V T Y G G Y S S M V

TGGTGGTGCACGAGCGGTTTCGTGGTCCGGTTCCCCGACGCCATGCCGCTGGACAAGGGCG
541 -----+-----+-----+-----+-----+-----+-----+ 600
V V H E R F V V R F P D A M P L D K G A

CGCCGCTGCTGTGCGCCGGCATCACCGTGTACAGCCCCATGAAGTACCACGGGCTCAACG
601 -----+-----+-----+-----+-----+-----+-----+ 660
P L L C A G I T V Y S P M K Y H G L N V

TTCCCGGGCTGCACCTCGGCGTGCTGGGGCTGGGCGGGCTGGGCCACGTTGCGGTCAAGT
661 -----+-----+-----+-----+-----+-----+-----+ 720
P G L H L G V L G L G G L G H V A V K F

TCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGGAAGAAGGAGG
721 -----+-----+-----+-----+-----+-----+-----+ 780
G K A F G M K V T V I S S S P G K K E E

AGGCCCTGGGGCGGCTGGGCGCCGACGCGTTCATCGTCAGCAAGGACGCCGACGAGATGA
781 -----+-----+-----+-----+-----+-----+-----+ 840
A L G R L G A D A F I V S K D A D E M K

G missing at 851 bp in the cDNA isolated from cv Ellett
resulted in a premature stop codon (truncated CAD2)

AGGCTGTGATAGCACCATGGATGGCATCATAAACCGGTATCTGCAAACATCCCCCTGAC
841 -----+-----+-----+-----+-----+-----+-----+ 900
A V I A P W M A S *

FIGURE 27 CONTINUED

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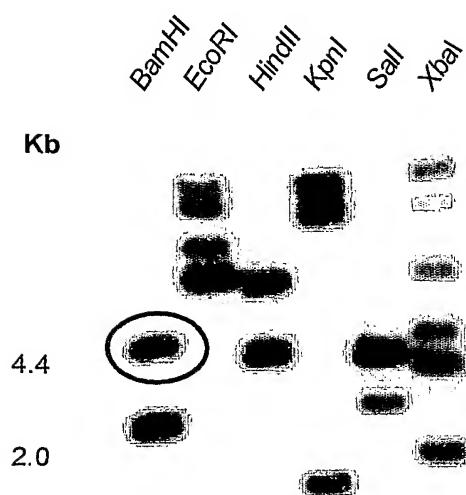
901 CCCTCTCTTCGGGCTGCTCAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGAA 960
-----+-----+-----+-----+-----+-----+
961 GCCCATCGAGATTCTCTCCCTTCGCTCTAGTTGCCACGAATAAGACCCCTGGCCGGGAGCAT 1020
-----+-----+-----+-----+-----+-----+
1021 CATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGGCCAAGCACGGCGTGAC 1080
-----+-----+-----+-----+-----+-----+
1081 GGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCCTTGGAGCGCCTTGCCAA 1140
-----+-----+-----+-----+-----+-----+
1141 GAACGACGTCAGGTATCGCTTCGTCATCGACATCGGCAACACCCCTCGACAATGTTGCGGC 1200
-----+-----+-----+-----+-----+-----+
1201 CACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGCT 1260
-----+-----+-----+-----+-----+-----+
1261 CCGTAGTAAACAATAAACGATCAAACTCTTGTTCATCTGGTGCATTGGTGTAGACATGGT 1320
-----+-----+-----+-----+-----+-----+
A) TGTTTTCGAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAAAAAAAA 1377
-----+-----+-----+-----+-----+-----+

FIGURE 27 CONTINUED

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FIGURE 28

A



B

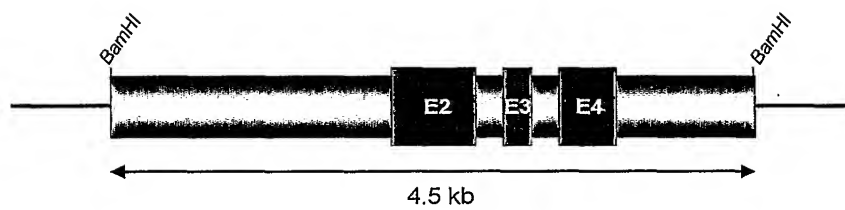


FIGURE 29A

A)

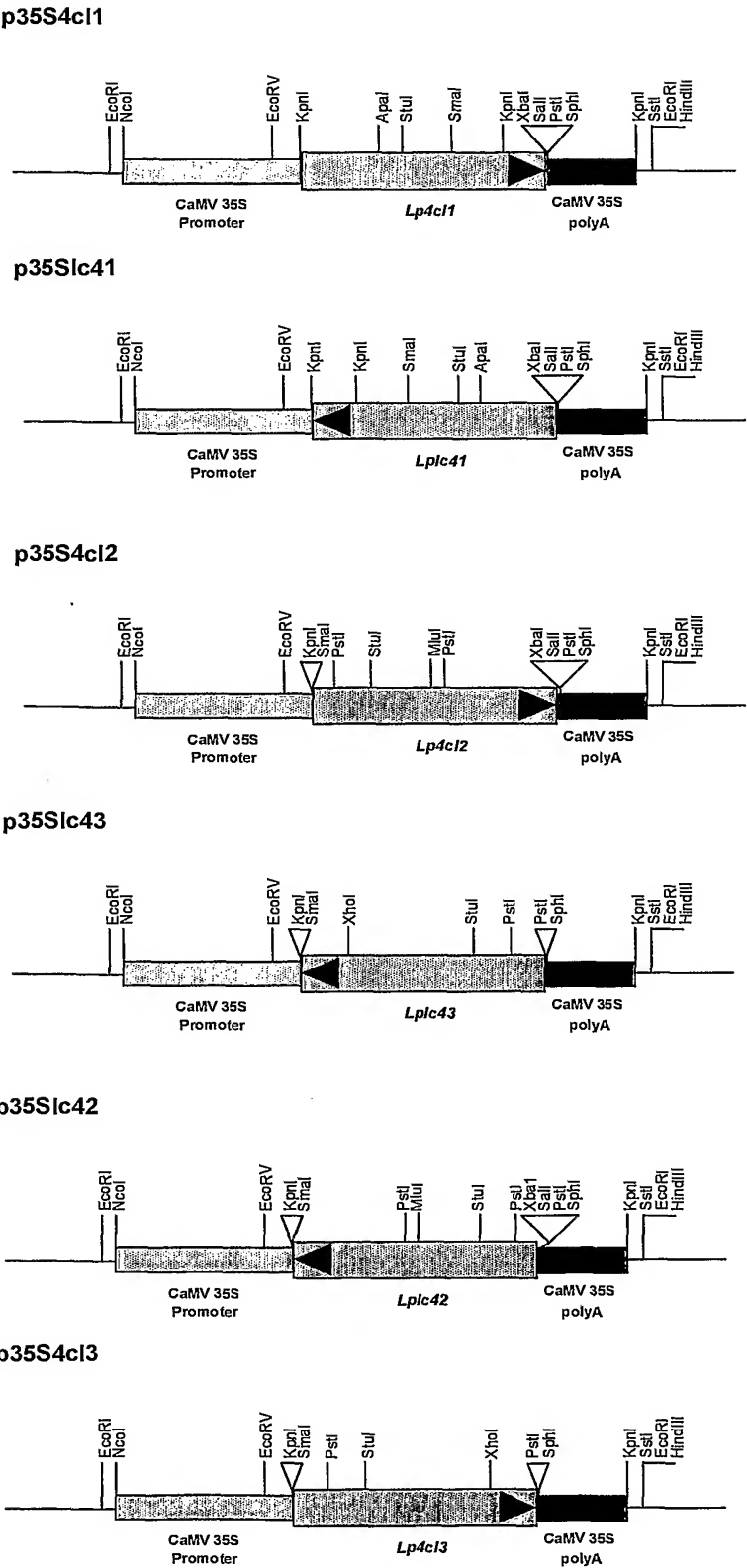


FIGURE 29B

B)

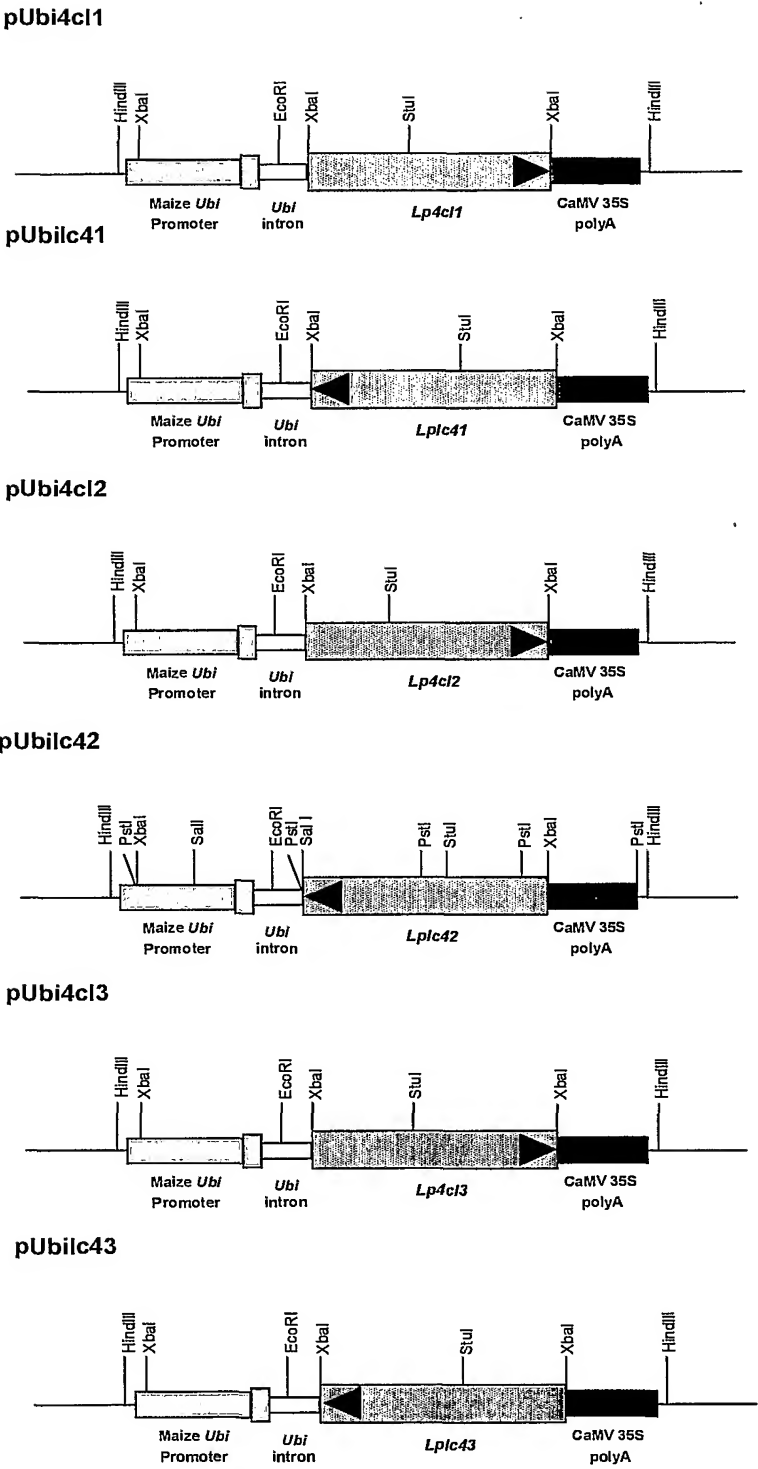
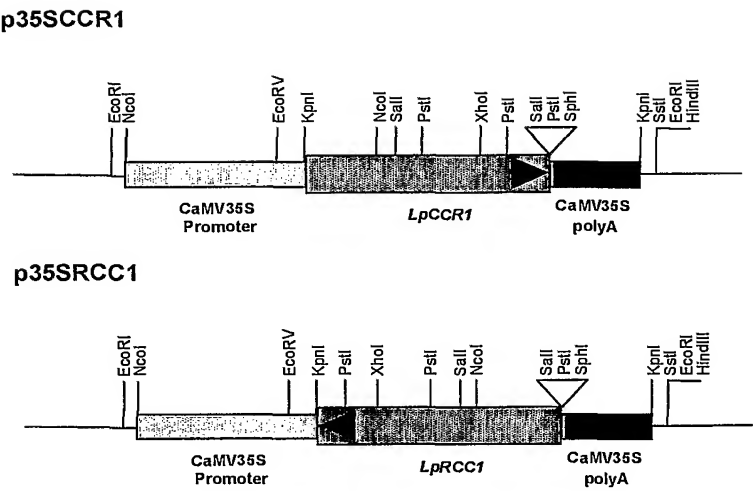


FIGURE 30

A)



B)

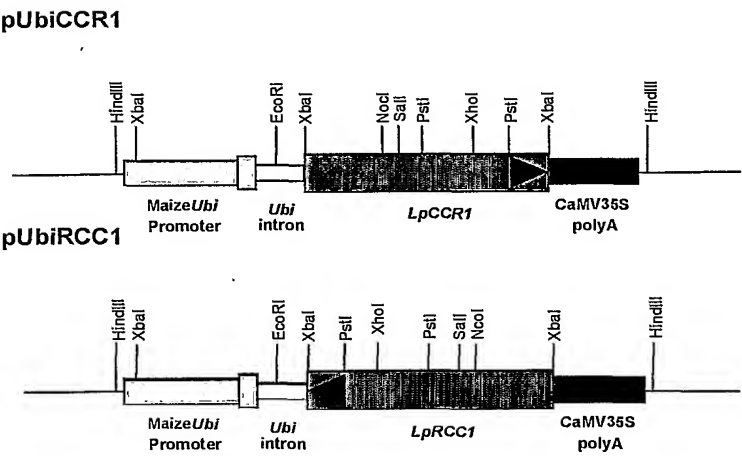
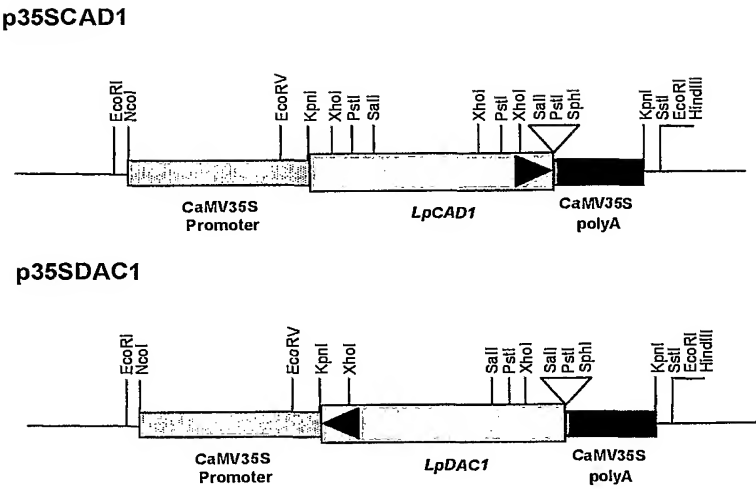


FIGURE 31

A)



B)

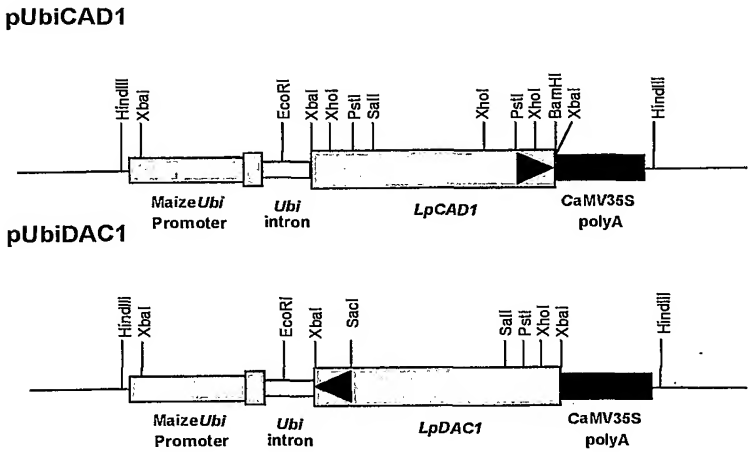


FIGURE 32

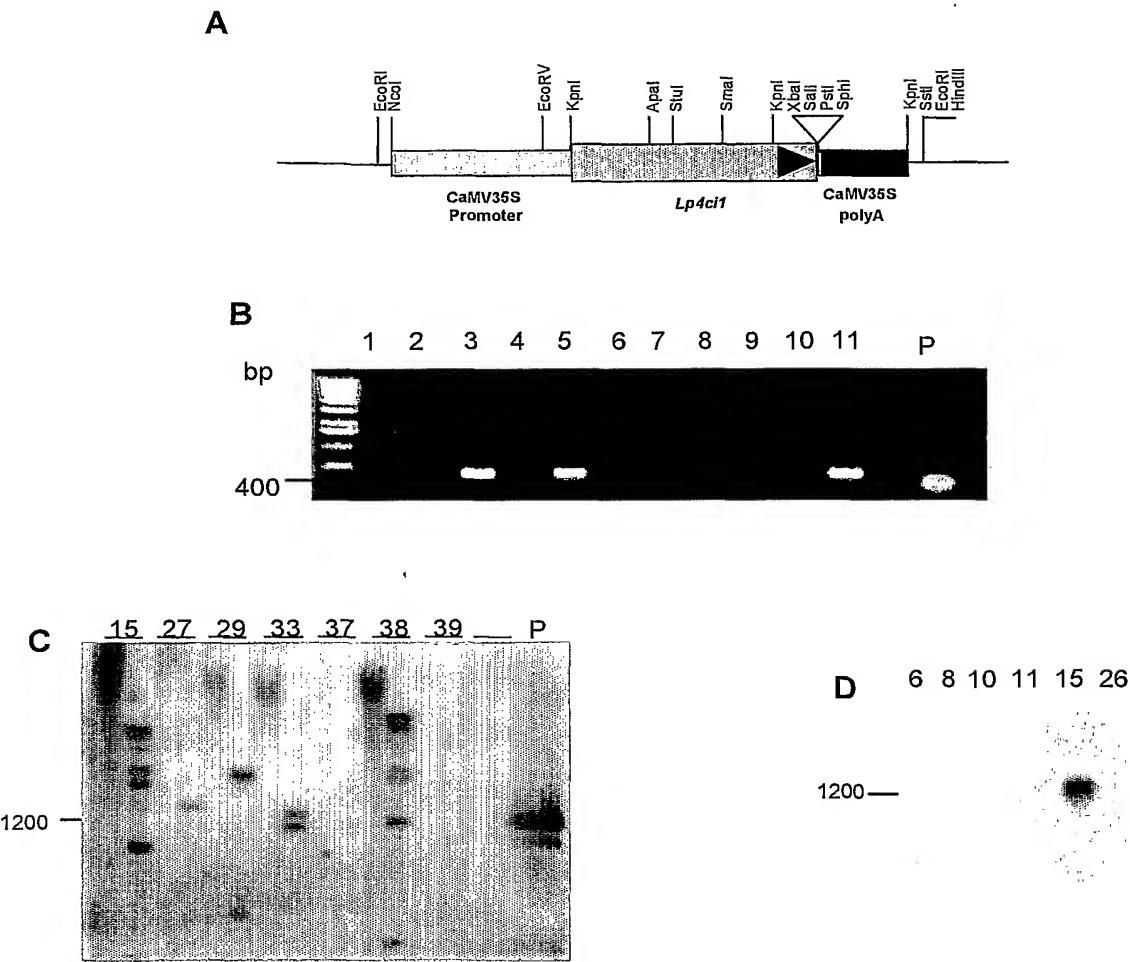
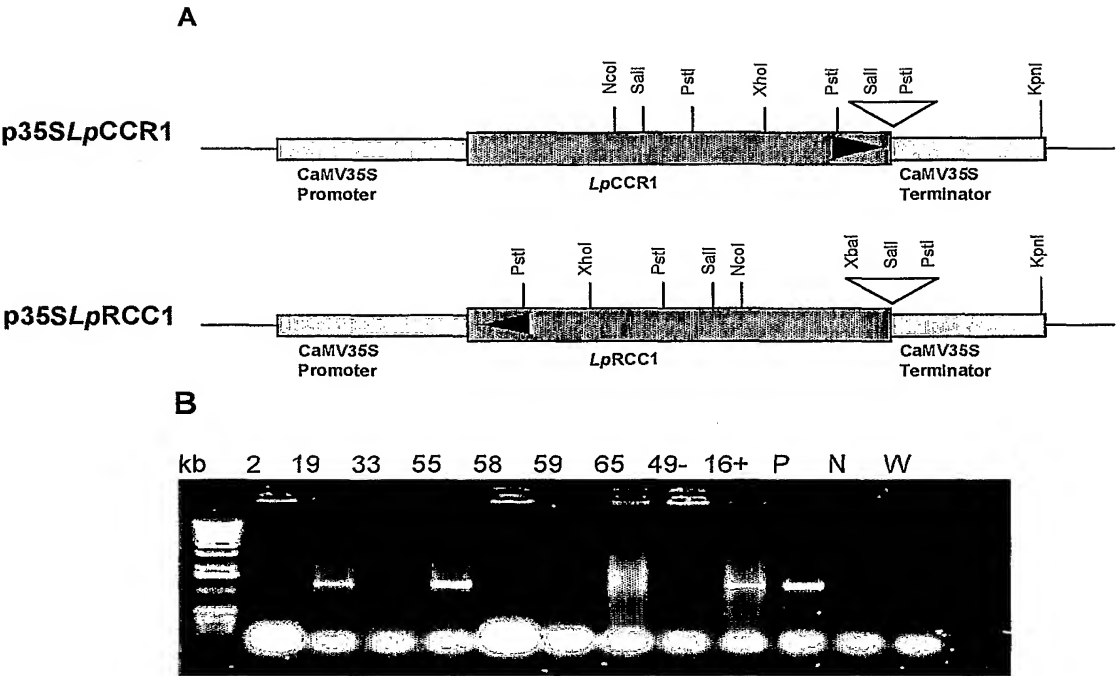


FIGURE 33



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FIGURE 34

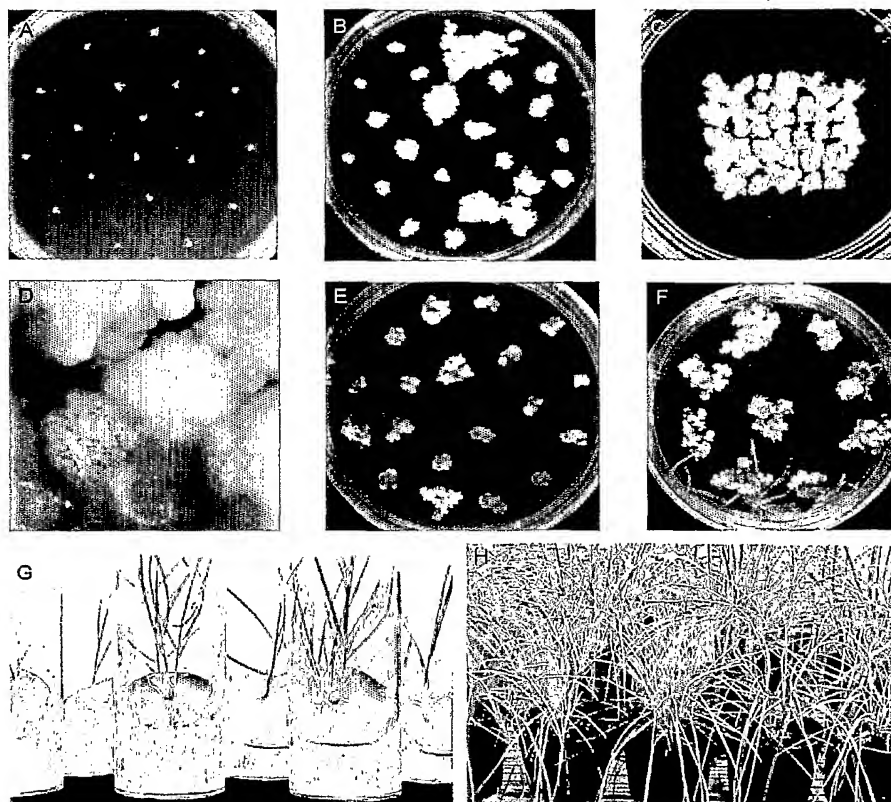
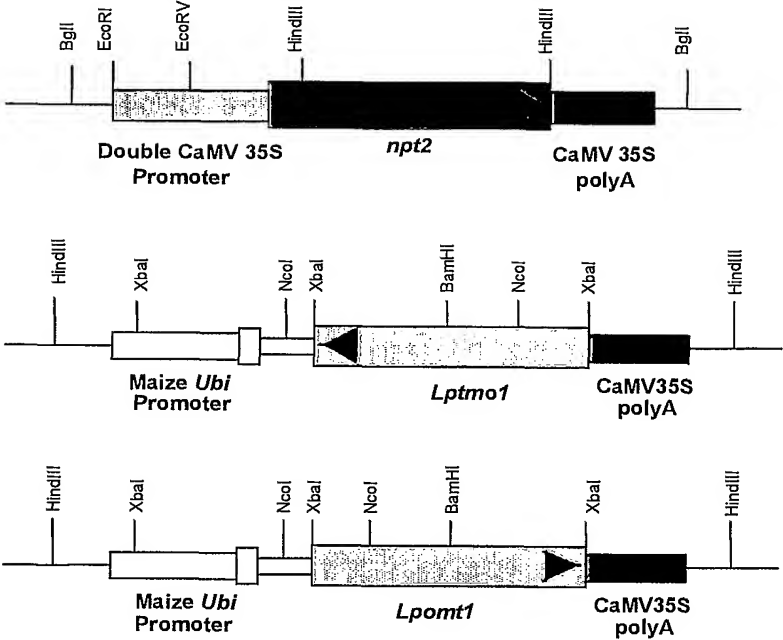
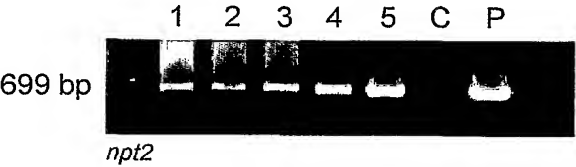


FIGURE 35

A

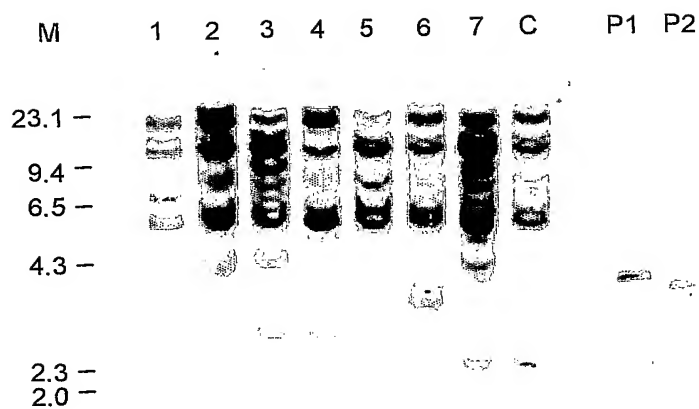


B



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C



D

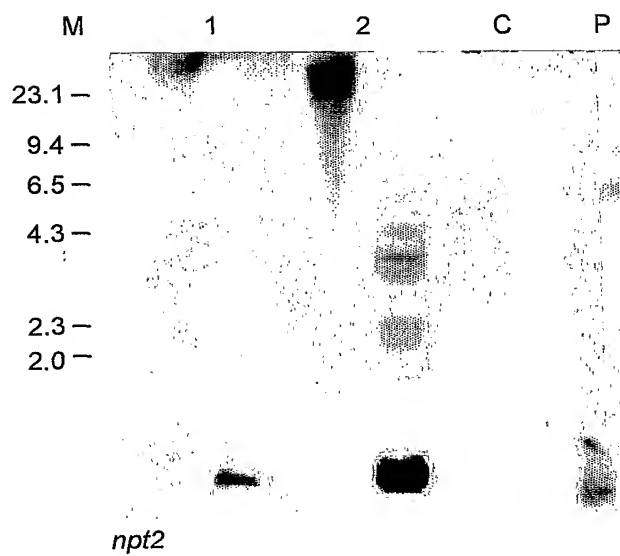
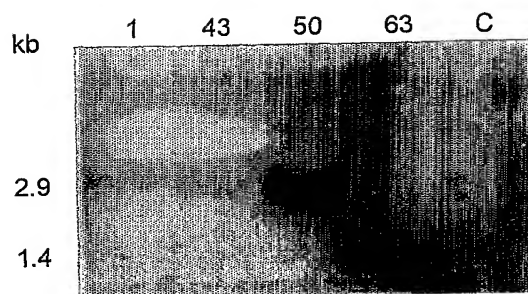


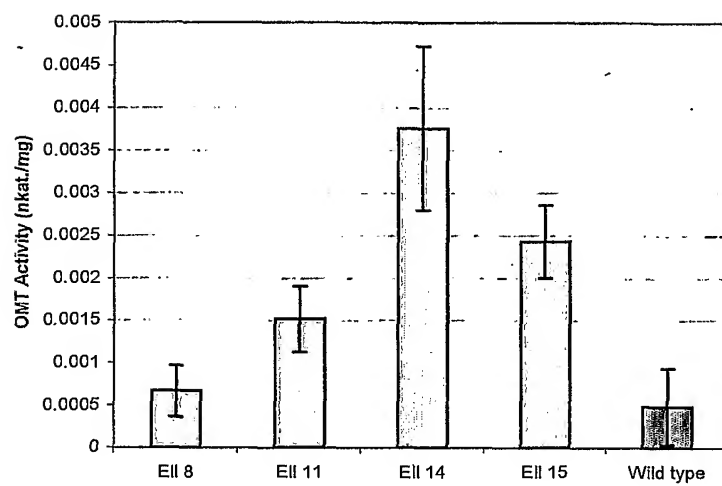
FIGURE 35
CONTINUED

E



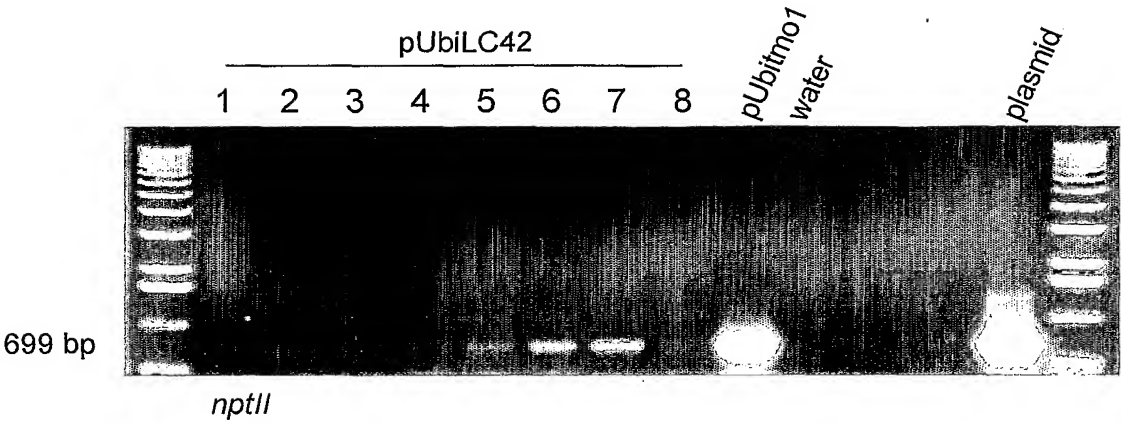
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FIGURE 36



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FIGURE 37



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-2206	CGGGATCAACTTGGATGTCCTTTGCGGGCACGGTTTCAGGAACAACGACACATGCAGCAG -----+-----+-----+-----+-----+-----+-----+-----	-2147
-2146	GGATCTCCTCCAAAGACTCACACAAAGGTGACATGAGCGCCCGCTTTTTTGAAGCCAAGT -----+-----+-----+-----+-----+-----+-----+-----	-2087
-2086	TGGCTAAGAAATCGCAAAGCTTGGTGGAGTCGGCCACCTCAGGATCTGCAACAAAAGGCA -----+-----+-----+-----+-----+-----+-----+-----	-2027
-2026	CCAAGGGAGCTGCCAACACATCAACCACAACATCATGTTCAAACGCAGTCTCCTCAAGCC -----+-----+-----+-----+-----+-----+-----+-----	-1967
-1966	TGGAATGCTCAACCGAAAGAGAGGGCAGAAGCTTCAACAAAAAACTCAGCCAACCCAAAGC -----+-----+-----+-----+-----+-----+-----+-----	-1907
-1906	CCTCGACGTCATCAGAGATTAGGCTCTGAGGACCCGCAGGGAAGCAACCTTGTCACCAAC -----+-----+-----+-----+-----+-----+-----+-----	-1847
-1846	CGCATCCGGCAGAAAAGGAGCAAGACCGGAGCAACCCTCAAGAGGCACACGAAAGACGTC -----+-----+-----+-----+-----+-----+-----+-----	-1787
-1786	GAAGCCAAGAGGAGACGAGTCGCAGGGACGGCGGACAGGCGAGAAGGGGCCGTAGAACTC -----+-----+-----+-----+-----+-----+-----+-----	-1727
-1726	CAAGAGCTCGGCGTCCCTCGACCTAGCATCCGAAGCACTGACCGGGGCACTCAATGCATA -----+-----+-----+-----+-----+-----+-----+-----	-1667
-1666	ACTTTATCTTGATGGCATATGTACTCAAACCCATACAATGTTTACCATGCATTATCTATG -----+-----+-----+-----+-----+-----+-----+-----	-1607
-1606	GAACATTCTTCATATACAACTtCTGAGTGGTCAGTGCATAGGAATTTTCATTAACAACC -----+-----+-----+-----+-----+-----+-----+-----	-1547
-1546	AAAAACATACTTGGGGCCTACACACACTTTCACAGCATGGAAAACCTGTTAGCTTTTTAA -----+-----+-----+-----+-----+-----+-----+-----	-1487
-1486	AGAGTTGCAAAATCTGTCAAGCGAATGTTCTTGTGATAATTGGAACGAAGCATGTTTCCC -----+-----+-----+-----+-----+-----+-----+-----	-1427
-1426	CATTTTCAATGTGTGTCTCTTACCCTAACTAGCACCCGACCAACAAAATCTGACCATCCT -----+-----+-----+-----+-----+-----+-----+-----	-1367

FIGURE 38

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-1366 AGTTATATCATCATAGAGACCCACATGTAGGTTGACCCCCATAACACTTGTGTGGATATC -1307
-----+-----+-----+-----+-----+-----+-----+-----

-1306 ATGGA AAAATGGCCTTGATCAACACTTCTTTCTACTTGGTACAAATGGTTATGGACTTA -1247
-----+-----+-----+-----+-----+-----+-----+-----

-1246 CTC AATTAGTGCTTTAGAGAGCTTTGGCTGCAGACTTTGTAGCTTCCCAATATTCATAGG -1187
-----+-----+-----+-----+-----+-----+-----+-----

-1186 TCCCTCCGAGTGGGCAGCCCCATCTACATAGGCTCAAAACCAGATTTTGTAAACATGTT -1127
-----+-----+-----+-----+-----+-----+-----+-----

-1126 AGACACTTTCAACTTCATCATAGACCATCAAGGAGCTGGCATGTGACAGTGATATATGTA -1067
-----+-----+-----+-----+-----+-----+-----+-----

-1066 TCAATTACCCATTCAACACGAATAGCTTGCTCATGCATGGTTAGTCTTGC GCGCGCGGGG -1007
-----+-----+-----+-----+-----+-----+-----+-----

-1006 CGGGACCATCGAACACACCGCCGGGCGGTCAGTAGGCTAGGGTTAGATAAAATCTAGCCG -947
-----+-----+-----+-----+-----+-----+-----+-----

-946 TTTTCATTCAAACCTTGTGATATATAATCAAATTTAAATAAAAACCTTTATTTTCGTGCAT -887
-----+-----+-----+-----+-----+-----+-----+-----

-886 TTTTATTTATTTGAGGGCGTGTTTGGGGGACACGGCTGGAAAGTGACATCCCCAAACACT -827
-----+-----+-----+-----+-----+-----+-----+-----

-826 GCACGAAGAAAACGCGTCGCCAAAAAATTCGATCCGGCGTCAGTCCTTTGGGAGACGATT -767
-----+-----+-----+-----+-----+-----+-----+-----

-766 TGGATGACGCGCTAGAGATGCTCTAAGTTCTCCACGCCATGTTTCTTTCTATATATACA -707
-----+-----+-----+-----+-----+-----+-----+-----

-706 CACAGCCCAAGGTCCATGAAAAGTAAAACGGCAGCAGACACGCACCGGCGACAACTTCA -647
-----+-----+-----+-----+-----+-----+-----+-----

-646 CATTACGGCACATCGCTATTACGGACCACATACTCCACCGCTATTCTCAGCCAAGTC -587
-----+-----+-----+-----+-----+-----+-----+-----

-586 ATACATGACATGATCCAATGGACGACTTTGTGAGCGAACTAGAACCTTGCGGGGTTTAG -527
-----+-----+-----+-----+-----+-----+-----+-----

-526 ATTTTCCAATGTGGATAAGTTGTACGCGCCGACTAGCTTTACACTTGGTTGAAAAAAGCT -467
-----+-----+-----+-----+-----+-----+-----+-----

FIGURE 38 CONTINUED

Sequence Name : CCR1genomicseq (SEQ ID NO: 18)

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-466 TATTGTAGCACGACTTCTCACTGACATAGGAATGTAAACAGTCTCTCCACGCCATGTTTC -407
 -----+-----+-----+-----+-----+-----+-----+-----
 -406 TTTCTAGTAGTAGCATACTAGTAGTAACCTTCTCTTTGTCTTACACACACCCAGGGTCCAA -347
 -----+-----+-----+-----+-----+-----+-----+-----
 -346 GAAAGGAAAACGGCACGACGGCACCCACCGACGACGACGACTCCACATCACGGTTCGGTA -287
 -----+-----+-----+-----+-----+-----+-----+-----
 -286 AAAAAAGTCAAACTCGCTGACGTGGCACCACCGGTCGCACTCAACTGACGCGCTCCTCT -227
 -----+-----+-----+-----+-----+-----+-----+-----
 -226 GCGCAGGTyTCACTtCAAGTTTCACCTACCACTGTGGGCCCCACCGCCAaTGTGGGCCCCG -167
 -----+-----+-----+-----+-----+-----+-----+-----
 -166 CGAGCTtCTtACTCACTGACCTGTCTCCACAGCCTCCTCGCCGGTATATTACCCCGGC -107
 -----+-----+-----+-----+-----+-----+-----+-----
 -106 CCCCAATTTCTCTGCCTTCCCACGAGCAGCAGCCGGAGCACGGAATCCCGGCCGCCATT -47
 -----+-----+-----+-----+-----+-----+-----+-----
 -46 CCTCCACCTTCAGCTCCGCCCAAAGATTTCATCCGGCGAGATCCATGGGCTCCATCGCG 13
 -----+-----+-----+-----+-----+-----+-----+-----
 M G S I A
 14 GCGGACGCGCCTCCCGCGGAGCTGGTGTTCGGTCCAAGCTCCCGGACATCGAGATCCCG 73
 -----+-----+-----+-----+-----+-----+-----+-----
 A D A P P A E L V F R S K L P D I E I P
 74 ACCCACCTGACGCTGCAGGACTACTGCTTCCAGCGCCTGCCGGAGCTCTCCGCGCGCGCC 133
 -----+-----+-----+-----+-----+-----+-----+-----
 T H L T L Q D Y C F Q R L P E L S A R A
 134 TGCCTCATCGACGGCGCCACGGGCGCCGCGCTCACCTACGCCGACGTGGACGCCCTCACG 193
 -----+-----+-----+-----+-----+-----+-----+-----
 C L I D G A T G A A L T Y A D V D A L T
 194 CGCCGCTGCGCCGCGGGCCTCCGCCGCTGGGGGTCCGCAAGGGCGACGTCGTTCATGGCG 253
 -----+-----+-----+-----+-----+-----+-----+-----
 R R C A A G L R R L G V R K G D V V M A
 254 CTGCTCCGCAACTGCCCGAGTTCGCCTTCGTGTTCCCTCGGCGCCGCCCGGCTCGGCGCC 313
 -----+-----+-----+-----+-----+-----+-----+-----
 L L R N C P E F A F V F L G A A R L G A

FIGURE 38 CONTINUED

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```

GCCACCACCACCGCCAACCCGTTCTACACGCCCCACGAGATCCACCGCCAGGCGACCGCC
314 -----+-----+-----+-----+-----+-----+-----+----- 373
A T T T A N P F Y T P H E I H R Q A T A

GCCGGGGCCAGGGTCATCGTCACCGAGGCCTGCGCCGTCGAGAAGGTGCGCGCCTTCGCC
374 -----+-----+-----+-----+-----+-----+-----+----- 433
A G A R V I V T E A C A V E K V R A F A

GCCGAGAGAG
434 -----+----- 443
A E R
```

FIGURE 38 CONTINUED

	TCGACGCCGGCCGCCTAATACGACTCACCTATAGGGCGAAAGAATTCCGGATCATATGGATTTCG	
-6735	-----+-----+-----+-----+-----+-----+-----+-----+	-6676
	ACACTGGAATTTACTCCCATCGGGAGCGTGCAACA AAAAGGTGTTATAGCAAGAAGACA	
-6675	-----+-----+-----+-----+-----+-----+-----+-----+	-6616
	CTGGCAACATTGCCAGCACAGAATTTGTTACAATCATAGAAAGTTTTATGACAGGACATT	
-6615	-----+-----+-----+-----+-----+-----+-----+-----+	-6556
	GTTTCAACCGAAAAGCAAGATTACAACAATATAATCAAGGGCTTGGGTCTGGTTGGACATG	
-6555	-----+-----+-----+-----+-----+-----+-----+-----+	-6496
	CTCGGTCCAATGGACGATTTATTTGCCGAGACCAGCTCAAGGAGTTGACGAGCACACTTA	
-6495	-----+-----+-----+-----+-----+-----+-----+-----+	-6436
	AGCGCCGAGATCTTAAAGGCACCCAAGTCAACAAGTCGCCCATCTTGCTCTTTTGGCAGC	
-6435	-----+-----+-----+-----+-----+-----+-----+-----+	-6376
	TCCTTGACATCTCTTCGATATTGGCTTTGAAGCCATGACCCATCATAAGCTGAAAGGCT	
-6375	-----+-----+-----+-----+-----+-----+-----+-----+	-6316
	AGGAGGGCACCATAGGTACGCGAAGTACGTTTGAATACCTCGAGGACCTCCCTCGTGTTG	
-6315	-----+-----+-----+-----+-----+-----+-----+-----+	-6256
	ATGGCGAAAAGCATCGATCAGCTGCCCAAAGGTCTTGTTTTGATCGATCTTGGGGAAGATC	
-6255	-----+-----+-----+-----+-----+-----+-----+-----+	-6196
	ATCGAGTGCATCCGCGTCATGGATCCTTTACCCCTTCTGAAGGAGGTCCTGAAAAAGCTGG	
-6195	-----+-----+-----+-----+-----+-----+-----+-----+	-6136
	TGAGACCCGAGGGTCATTGACAAAGCATTCGCCGGAGAATTATTCGGCAATTTATCTAGA	
-6135	-----+-----+-----+-----+-----+-----+-----+-----+	-6076
	GCCCTCAGCAGGGATGTAGGCAGCTTCTGGAGAAAGTGAAAGAGGAGAGCTCAC'TAACCA	
-6075	-----+-----+-----+-----+-----+-----+-----+-----+	-6016
	AAATCAAATCGATAAAGCAAAAAATCGGAAAGGAGGCCAAAAGGGGATTACTGAGCAAGGC	
-6015	-----+-----+-----+-----+-----+-----+-----+-----+	-5956
	CAAGGAAGATTGGCGAAGGAGCTCATCTTTTTTCAATCGCCGAGCTTCGGCAGCAAGCCT	
-5955	-----+-----+-----+-----+-----+-----+-----+-----+	-5896
	GGATGCCTCTTCATCCTTCAGCCTCTTTCTTAGCCCCCTCGAGCTCATCCTTAAAGGAATC	
-5895	-----+-----+-----+-----+-----+-----+-----+-----+	-5836
	AACCTCCTGGCGGGCCTCGGCAGCTATCTTTATCGCACCTCCAGCTTCGAGGAAGAAGA	
-5835	-----+-----+-----+-----+-----+-----+-----+-----+	-5776
	CTCGACCTCCTTTTGCAGCCGAGTCTTGTCAACTTCCAGAGAAGTGATTGGGAGGCGAA	
-5775	-----+-----+-----+-----+-----+-----+-----+-----+	-5716
	GGCCTCCAGAGAAGAGATAACAGCTCACAAATCCTTAAGAGATAAGGAAAAATAATTAGA	
-5715	-----+-----+-----+-----+-----+-----+-----+-----+	-5656
	CGAAGAACTGGTTGTCAACAAACTTATAATTTGATCAGGGAAATCGTCCACATGGATAT	
-5655	-----+-----+-----+-----+-----+-----+-----+-----+	-5596
	ATCGTTAAAAACAGGAAAAGCTTACAGGTTTCCCTGGAGGAGAAGCTGTAACCACGGCAGT	
-5595	-----+-----+-----+-----+-----+-----+-----+-----+	-5536

FIGURE 39

	CAAAGAAATCTCCTTCCCTTTGGAAAGGGAAGAAGTTGTTCGATATTTGAGCCATGGGGGC	
-5535	-----+-----+-----+-----+-----+-----+-----+-----	-5476
	TGCGGCAGGAGTCTGAAGCCTCGGAAGCGGCTGGATTTCGGCACGATGGCACCAGATTGGC	
-5475	-----+-----+-----+-----+-----+-----+-----+-----	-5416
	CTTCTTGGCCGGAGGCTCGATGAAGCCATCTTCACTGCAAGAACAAAAACTAGCGAAGT	
-5415	-----+-----+-----+-----+-----+-----+-----+-----	-5356
	CAGAATTCAATGCATATGGCGAAGTTAGAACAATCCTGGAAAAGGAAGCAAGGACTTA	
-5355	-----+-----+-----+-----+-----+-----+-----+-----	-5296
	CAATTCATAGAGACCATCTTCATCGGCAAAGCCGCCGGATGATCTCTTTGGAGGTAGTGC	
-5295	-----+-----+-----+-----+-----+-----+-----+-----	-5236
	CTCGGCCTTTTCCGTAGCTGCATCAACAAAGGCAGCACGATCAGCATCGTCATCATGCAT	
-5235	-----+-----+-----+-----+-----+-----+-----+-----	-5176
	TGACCCCGCTGTATCGCTCATATCATCGGCAGAGAATCGAGGATTGATGGAAAAGCCTC	
-5175	-----+-----+-----+-----+-----+-----+-----+-----	-5116
	AGGATTCATCGGATCATCATGTTGATCTATCGGGCTTGCAATCCCTAGAGTATGGGACCC	
-5115	-----+-----+-----+-----+-----+-----+-----+-----	-5056
	TACAAGGACTAAGGAATCCCTTTTCTTGGAAAAATGTTTCGACAGGTCTTGCAAACGTTC	
-5055	-----+-----+-----+-----+-----+-----+-----+-----	-4996
	AAGAGCCGTAAGGATCTGTCGTAGTTGACGAGTGAGAATAATGGCAGTTAAATAATCAA	
-4995	-----+-----+-----+-----+-----+-----+-----+-----	-4936
	AGGAACATGACAATAAGAGCATAAAGGGGAAATTTACCTCGGTTGGCAGATGACCAGCGT	
-4935	-----+-----+-----+-----+-----+-----+-----+-----	-4876
	CAAATGGCGGTTGAGGAGATATCAGTGGAATTGAATCTTCCTGGCTAAAGAGGGTGAGAC	
-4875	-----+-----+-----+-----+-----+-----+-----+-----	-4816
	ACCGGACTTCGTCAAGCAGTTCTTTTTCGGATAATTCAGCAATATTTACTCTAGTCTCGT	
-4815	-----+-----+-----+-----+-----+-----+-----+-----	-4756
	CCCTGGGACCCGAATACAACCACATCGGATGGGTCTTAGACATGATCGGCTGAACTCGAT	
-4755	-----+-----+-----+-----+-----+-----+-----+-----	-4696
	GTTTTAAGAACACAGCGGCTACCTCAGTACCTATCATGGTTTGACCATCGGATTCTTTGA	
-4695	-----+-----+-----+-----+-----+-----+-----+-----	-4636
	TCCGAAGGAATCTATCAAATAACTTGTCTACTGTTGGTTTTTCATCGGGTGAGAGGATAT	
-4635	-----+-----+-----+-----+-----+-----+-----+-----	-4576
	TTTTCCAAGACTTCTTGGGCTTGCTTCTAGAACATCGGAGAATTGGGGGGGAGCTGGGAG	
-4575	-----+-----+-----+-----+-----+-----+-----+-----	-4516
	TCGGCTGCTGATGAGTCCTTAATATAAAACCACTTCAGCCTCCAGCCTTGCACGGATTCT	
-4515	-----+-----+-----+-----+-----+-----+-----+-----	-4456
	TTCATCGGGAAGTTGAAGTAGTTGACTTCCTTACGAGCAACAAAACCAACCCACCAATG	
-4455	-----+-----+-----+-----+-----+-----+-----+-----	-4396

FIGURE 39 CONTINUED

2

	ACGAAGGACCACCACTGC	TGTATTATC	TTTTCACGAAGAAAATC	TTCTTCCACAAAACCA	
-4395	-----+	-----+	-----+	-----+	-4336
	AAGTGGGGCTCAATGCCA	AAAAACGCTTCGCAGAG	GGTGATAAAGATGGCAAG	GTAAGG	
-4335	-----+	-----+	-----+	-----+	-4276
	ATTGAGTTGGGGGTTAAC	TTCATAAATGAATCTCATACACTCGAAGGAGGTGGTGAAGA			
-4275	-----+	-----+	-----+	-----+	-4216
	AATTTGTGAGCGGGAAGCG	AAAGACCTCGGTACAAGAAGGATAAGAACATCACAGTAAAA			
-4215	-----+	-----+	-----+	-----+	-4156
	CCGGCAGGAGGATTGGGCCGTGAAATTG	CACCTGGAAGAATAACATTCCCCTCGTCAGAA			
-4155	-----+	-----+	-----+	-----+	-4096
	GAAATTATTACGAGGCTCCGGGCCCTCTTTTCATCTCGCTTCGTGGTGGTAGAAGCTGGC				
-4095	-----+	-----+	-----+	-----+	-4036
	CAATCGCCAGGGATAGGCCCGGCCGTGGAGCTTGACGGCGCTGGCGGTGCCGAGCTGAG				
-4035	-----+	-----+	-----+	-----+	-3976
	GGAGGAGCATCTGGCGCGCTTCTCCGCGCGGATT	CGAAGGAGCCCTGACGGTGGTGCCA			
-3975	-----+	-----+	-----+	-----+	-3916
	CTGCTCACGGCGCTGGTGGCGAGAGTGGGATTCTTCTTCTTACCATTGTGAGATTTGAG				
-3915	-----+	-----+	-----+	-----+	-3856
	GGAGATCTGGGAGTTGCGACGGTGGCGTGGTAGTTGCAAACGAAAAGGATGAATGAGGAA				
-3855	-----+	-----+	-----+	-----+	-3796
	GAAGGGACGCAAGGATGAAGTGTGGAAAGGGAGTTACCCCCAAGAGATTATAAAGTGAA				
-3795	-----+	-----+	-----+	-----+	-3736
	AGGAAAACCTGAGAATTGAGCGGGCACGTGTCGTTGCTCTCAATTTATTGAGGGGATTTT				
-3735	-----+	-----+	-----+	-----+	-3676
	TTCTCATCATAGATCGCGGAAATCGAGGAGTCACCTTGGTAAC	TGCACGCAAGTAGTGGT			
-3675	-----+	-----+	-----+	-----+	-3616
	CATTTCTTAAACAGAACCGCATAGAAGTAGGATGGGACCGTCAGGTCACGTCTATCAGT				
-3615	-----+	-----+	-----+	-----+	-3556
	CAGATTTACAACAGTAATTACATCATCACTGACGTCAAAGTATGCTTGAAGTATCCGAAG				
-3555	-----+	-----+	-----+	-----+	-3496
	AAAAGTCGAAATTTGGGCTCGAAGACTTTCTTG	CAGAGAAGCGCGTGAAAGGAATATCTA			
-3495	-----+	-----+	-----+	-----+	-3436
	AGGAAAGGGTCAAAACATT	CGGCTCGAGTCTACGCACGGATTGCAAGCATCCGTACCTAG			
-3435	-----+	-----+	-----+	-----+	-3376
	ACTCGGGGGCTACTCCCATCGGGAGCGCTGGACGTGCACCCGATAAAATTTAGACGAGGAT				
-3375	-----+	-----+	-----+	-----+	-3316
	GAA AACCGGAAACCAAGTGCTACTCCCATCGGGAGCGCCGATTACGCACCCGACAAACT				
-3315	-----+	-----+	-----+	-----+	-3256
	TTTTTGC	ACTCCAGGATCATGCCCGGGAC	TTAATTTCTGTGTAGAGTAGCGTTGTTTTGT		
-3255	-----+	-----+	-----+	-----+	-3196

FIGURE 39 CONTINUED

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-3195	CTTCGGCAGTTAACCAGCAAAGCTGGACACGTTACTCAATATCCTTTACGCATTAAACCC	-3136
-3135	TTACTTGAAGAATTGAAGCCCCGATGCAAATATATCGGATGACCTATGAAGGCCGCGGA	-3076
-3075	AAGCTTCGGGAGAAGAAGACATTCGAGTGGCACAACCTGAGTCTACGAACGGATTGCAAG	-3016
-3015	CATCCGTACCTAGACTCGGGGGCTACTCCCATCGGGAGCGCTGGACTCGCACCCGATAGA	-2956
-2955	AGGAGATGATGATATTACAAGAAGGACAAGAAGTATCAAGGGAGAAGAACATTTCGGTGGA	-2896
-2895	GGCATGCTTTAGTCTCTACCCGAAAAAAGTTCGGCTAGACACTCGGGGGGCTACTGACGT	-2836
-2835	GGGCATTACCTTTCGGGTAAGTATGATATGCCCTATCCTGTACGACCCAACTGGAGGCCCA	-2776
-2775	TGAAGACACTCGAAGGCAAGGTGGACCACTACGTCGGTGCCGAAGGGGGTTCCTTGAAGA	-2716
-2715	ACAAGACGAAGAAAAGAAGAATACAAGAAAAGTATAGAAGTACCTTTTGTAACTGG	-2656
-2655	TCGTACCCGGACAGATCTCTCGAGACCTGGCCCCCTACATATGGGCTAGGAGAGGGGCTG	-2596
-2595	CCGAGAGGGACACACACAATCTTAGCAATTTTAGCCACCATAAGTCCAGAGCAAGGTCCC	-2536
-2535	CGTAGAACTTAGCCTCTCGACGAGATCAGACCCGAAACCTTCGGCACCCCATTTGTAACCC	-2476
-2475	GATATTTTCATAGTCAAGATCAGACAGGTAGGACGTAAGGGTTTACCTCATCGAGGGCC	-2416
-2415	CCGAACCTGGGTAAATCGCTCTCCCCGCTTGTTTGATAACCGATGGCTTGTCAGCTTA	-2356
-2355	CATGATTCCATCTACCCCTAAACCTCAAACGGAGGGCATTGCCGAGGAGTACCCCTCGACAT	-2296
-2295	TCCCCCTCCACCAATGGTCTCACATAAATTCACAAAGCAAACCTCATAAAAAGTTTAATGA	-2236
-2235	GTTTCAGAAAGAAATAAACTAGGCCCTCCTTTGAGAATCTACGAATGATTACCATAT	-2176
-2175	CATCTCGCAGTTAGTGATGAGTAACCTAAGTCTCAAATTTCCCGACGCATGGCGAAAAAGG	-2116
-2115	TAGCGAACTTAAATGTGAGGAATGAATGCCACATATGCATGGTGCATCGAGTATTCTCA	-2056
-2055	TTTTAGTCTTGGATTACTCCCTTTAGATGTTGACACCATCCCAAAAATACAACCTTGACA	-1996

FIGURE 39 CONTINUED

68/76

AGTTGTTTCATTTCACTAGTATGAATTTTCAGTAAATCGGGCAATACTCCAACACTCATTCA
-1995 -----+-----+-----+-----+-----+-----+-----+----- -1936

CCCCCTAGGCGAGGTTAGCTCAGATCAACGTCGGGTGTCTTCATCGAGTTAATGTCGTCA
-1935 -----+-----+-----+-----+-----+-----+-----+----- -1876

CACGCACACACACGTACGCGCACACACACGTGCGCAAAACAAAAGAAAACCTAGGAACCTT
-1875 -----+-----+-----+-----+-----+-----+-----+----- -1816

CTCACGTAGCCTAGGTCTTGTCTGTAAAGAAAAACCCAGGTCCACCCTAGTTTCGAACC
-1815 -----+-----+-----+-----+-----+-----+-----+----- -1756

AAAATATTTTGAAGATACATTAGTAAGATATTTTGAATAAAACCGCAAAAGGGAA
-1755 -----+-----+-----+-----+-----+-----+-----+----- -1696

TTGAAAAATATGGACTGGCTGTTTGTGCCAAAACCACATCTTTCGGAGAACCACGAGGGT
-1695 -----+-----+-----+-----+-----+-----+-----+----- -1636

ATCTATTGATGGGCTCATACTATACCTGGGCATGTGTTGGGCCAGGCCTCATGTCGGGCC
-1635 -----+-----+-----+-----+-----+-----+-----+----- -1576

GAGGAAAGCCCGACGCTGAAAAATCAGGCCCAAGCTTAACCCGGCCCGACCAATACCCA
-1575 -----+-----+-----+-----+-----+-----+-----+----- -1516

CCAAACCCGTTGGGCCATCAGGTTGCGGGCCGGGCAGTAGTGTAACACCGATTTCGGG
-1515 -----+-----+-----+-----+-----+-----+-----+----- -1456

CTACATAGGCCCGGCTCGTTTGTGCGGCAACATTTCTAGACCTAAGCCCGAGTTTTTCG
-1455 -----+-----+-----+-----+-----+-----+-----+----- -1396

GGCCGGGCTGCCCATGGCCAGGTATAGCTCATAACGACGTATGACATTTTCGAGCAATTGA
-1395 -----+-----+-----+-----+-----+-----+-----+----- -1336

TGCAAAGCACGTGTAGGGTTTTATCCCATCCGTGTGGCGTGTGTAGGGTGTAATGAATA
-1335 -----+-----+-----+-----+-----+-----+-----+----- -1276

GGATAATTTCTCGCCGAAACTGGTCCCAAATTCGCTTTGAAGTGTCATATATGATTTT
-1275 -----+-----+-----+-----+-----+-----+-----+----- -1216

AAAGAATGTGACAAATAAGATATCCAATTTGAAATAGTGCTCCGATACGGTATAGGA
-1215 -----+-----+-----+-----+-----+-----+-----+----- -1156

TATGGTATAGCAAATAACATGCTGATATGGATTGTCCGATATTAAATTAAGATAATCCAA
-1155 -----+-----+-----+-----+-----+-----+-----+----- -1096

ATGTTTTAAACCGCATAATTCGATTTTTGAGTCAAAAGCGAATGCCAATTCAGAAGGTTA
-1095 -----+-----+-----+-----+-----+-----+-----+----- -1036

GCAGTTATTGAGTTTCAAATTTATTTGGCGAGCATATCTAGTTCTAAATTCATACAGT
-1035 -----+-----+-----+-----+-----+-----+-----+----- -976

AAATTGTGTCTTTTTTAATAACTACACAAGACTAAAAGTTTAAATCTCTCTCAAGATTT
-975 -----+-----+-----+-----+-----+-----+-----+----- -916

GCGAAAACATAGCTATCTACTGATATATATATCCGACTATATTGTTTTCGGACCGCAT
-915 -----+-----+-----+-----+-----+-----+-----+----- -856

GCGTCCATTTCCGATTTCGAATCTGCACTCCGATATATCCACATTGAATCTAAAACCGAT
-855 -----+-----+-----+-----+-----+-----+-----+----- -796

FIGURE 39 CONTINUED

69/76

-795	CAATATTTGCTCCGATCTAAATCCGAAAAATATGTGGTGAAGGATATGGTATAAGCAAA	-736
-735	ATCCGATTTGATCCATTTGTACCTCTAGGCGTGTGCAAGACCTGGAGGAAAGAATGGCGC	-676
-675	ATCTGTAGGGTGCAGTCCCACCGGTGAAAAATGTGAGCTCACCGTATTGTCCCCGATGG	-616
-615	AGCATCGAAACGGAGTCGGAACACGATTTGCGCCACGTACAGAGCATGCATGATTTCCCT	-556
-555	TGTATGCGGTCCAGGATCTTAACTGCCTTCCATTTCCAGGAACCTACCGATTGGCTGCA	-496
-495	AGCCGTAGCTAGCGGTTTGAAGTCACGGCATTGCCGCCCCCGATTAAACCCACCCGTCGCG	-436
-435	CGCGCGGTTCGTCGTTTACCCTCCTGCCTAGGCTACGCACGCGCGCGCGCAGTTGGGCC	-376
-375	AGTTGTAGGTAAGCCGACTCGAGATCACACACCCGGCCTCACCTACTACCTCTCGCCGTC	-316
-315	GCGGTCACCGTGTACACTCACGCCCAGGGGAGCCACCCGCCACACGGCGCCTAGCTCA	-256
-255	TCCCCTCTCACTACTCTTCTTCTCCTCCCTCTCACCTCGCCGTCGACCCAGCTCCCGGCT	-196
-195	CTATAAATTCCGCACTACTCGAACCAACATCGCCCAGGCCTTTGCCTTTTACGACGAATC	-136
-135	CTACCAAACCGAGCTACCAGATCCTTCTCTACTAATCGAGCTCCCTACGCTGCTCCGCCT	-76
-75	GTCTTCGTTTCCGCTCACCGCCGGCCGGTTCTCCGCTCCAAGCTACGTCCGTCCGTCCA	-16
-15	CATATATAGCATCGACATGACCATCGCCGAGGTCGTGGCTGCCGGAGACACCGCCGCCGC	44
45	GGTGGTGCAGCCCGCCGGAACGGGCAGACCGTGTGCGTGACCGGCGCCGCGGGTACAT	104
105	CGCGTCGTGGCTCGTCAAGCTGCTGCTGGAGAAGGGGTACACCGTCAAGGGCACCGTCAG	164
165	GAACCCAGGCATGTACCCATGCATTTCATATTTTCTTACTAGTCGTATGCGTTATGCGA	224
225	CTTGTGTATTAACTATTGTGGACTGCATGCAGACGACCCGAAGAACGCGCACCTGAGGGC	284

M T I A E V V A A G D T A A A

V V Q P A G N G Q T V C V T G A A G Y I

A S W L V K L L L E K G Y T V K G T V R

N P G

D P K N A H L R A

FIGURE 39 CONTINUED

70/76

GCTCGACGGCGCCGCGACCGGCTGGTCCTCTGCAAGGCCGACCTCCTCGACTACGACGC
 285 -----+-----+-----+-----+-----+-----+-----+-----+----- 344
 L D G A A D R L V L C K A D L L D Y D A

 CATCCGCCGCGCCATCGACGGCTGCCACGGCGTCTTCCACACCGCGTCCCCCGTCACCGA
 345 -----+-----+-----+-----+-----+-----+-----+-----+----- 404
 I R R A I D G C H G V F H T A S P V T D

 CGACCCCGTACGTACTCCATAGAACTCGGCACCCCTAGCTTCTCTCCGTTCTCTCTGTAT
 405 -----+-----+-----+-----+-----+-----+-----+-----+----- 464
 D P

 GTCTGTACCGTCGATCGCCATGGCAGCACGCATGCATGCGCGCGCAACGCTAGCTAGAC
 465 -----+-----+-----+-----+-----+-----+-----+-----+----- 524

 GCTGACCGACTCATTGTGTCAGGAGCAAATGGTGGAGCCGCGGTGAGGGGCACGCAGTAC
 525 -----+-----+-----+-----+-----+-----+-----+-----+----- 584
 E Q M V E P A V R G T Q Y

 GTCATAGACGCGGCGGCGGAGGCCGGCACGGTGC GCGGATGGTGCTCACCTCCTCCATC
 585 -----+-----+-----+-----+-----+-----+-----+-----+----- 644
 V I D A A A E A G T V R R M V L T S S I

 GGCGCCGTCACCATGGACCCCAACCGCGGGCCGGACGTGGTCGTCGACGAGTCGTGCTGG
 645 -----+-----+-----+-----+-----+-----+-----+-----+----- 704
 G A V T M D P N R G P D V V V D E S C W

 AGCGACCTCGACTTCTGCAAGAAAACCAGGGTGGGTGCTGCATGCTCAATTTTATTATC
 705 -----+-----+-----+-----+-----+-----+-----+-----+----- 764
 S D L D F C K K T R

 ATAGCTACCCTTTTCTGCACCATGCTGCATTTCTTTTCCAAAAACAACCTCTCAAAAGAT
 765 -----+-----+-----+-----+-----+-----+-----+-----+----- 824

 ATGCTACGTGGTGAGTTCTATAGCTGAATTATTACAACCTACCACCCTATCGATCACTAC
 825 -----+-----+-----+-----+-----+-----+-----+-----+----- 884

 CGCCCTAAAAGTGTTCAACTTTTGAAGGCAACCAAAACCAATACATGAACGACGATCGTG
 885 -----+-----+-----+-----+-----+-----+-----+-----+----- 944

 TCGCTTGTGTCGTTATCATTAGCCTCTGTAGCTCTAATTTTACCTATGTACGCATGG
 945 -----+-----+-----+-----+-----+-----+-----+-----+----- 1004

 ATAGACGATTTCGAAATACAGTTCAGTTTACCTACCATATACTATGCCGAAATCGAACGC
 1005 -----+-----+-----+-----+-----+-----+-----+-----+----- 1064

 ACACAGGTGTGAGGCAGCAGCCGCTCACGAGTTATGCGCCGAAACCGACATCTCGGAATC
 1065 -----+-----+-----+-----+-----+-----+-----+-----+----- 1124

 TTCAGTCCACAATCAAAAAATAGACACCTGGTACCACTACAAAATTATACTCCTACTGTA
 1125 -----+-----+-----+-----+-----+-----+-----+-----+----- 1184

 TATTGGTAAAACAAAACATTTTCTTTTATTTGATAGGAGTGCTGCAAATTAAAGTTCT
 1185 -----+-----+-----+-----+-----+-----+-----+-----+----- 1244

 TTGTGTCATTTTCAAAGGAAAAAAAAAACACCTTTACCACTCTTCTTCCTTGCCATCAT
 1245 -----+-----+-----+-----+-----+-----+-----+-----+----- 1304

FIGURE 39 CONTINUED

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1305	TTTTTTTTTACCAAAGTTTGTTCGTCAAATGAACATATATATAGTTCGGTGCTATGTCA -----+-----+-----+-----+-----+-----+-----+-----	1364
1365	GTGCCATTACCGGCCACTAGCTAGTAGGACTGCCATGTTCCAGCAAATTGTCTAGTGGA -----+-----+-----+-----+-----+-----+-----+-----	1424
1425	CCGGAGTGGCCAAAAGGAGCCAATTATGTAGGGTTGCAAGCGGGATCACACAAAAGCCTC -----+-----+-----+-----+-----+-----+-----+-----	1484
1485	GCCTCTAGTTCATTTTATCAATTAAGTGGTACTTTCTCAGGGACCCCCCTTGCAACTCTA -----+-----+-----+-----+-----+-----+-----+-----	1544
1545	CCATTACATCCGTGCAAAATAAAAGCTAGCATCACGCACCAGATTTAGTACTCCCTCCGT -----+-----+-----+-----+-----+-----+-----+-----	1604
1605	TTTTATTTAGTTCGCATTCTAGGTTGAGCCAAAGTCATACTTTGCAAAGTTTAACCAAAA -----+-----+-----+-----+-----+-----+-----+-----	1664
1665	TTATAAGAAAAAATATCAATAATCATCATACAAAATACATATAATATAAGAGTAAACCT -----+-----+-----+-----+-----+-----+-----+-----	1724
1725	TATAACGATTCTACAATAGATTTTTTTTATTGCATATGTCAATATTTTTTCATAAATATTT -----+-----+-----+-----+-----+-----+-----+-----	1784
1785	ACTCAAAATTATAAGGTTTGACTTTGACTAAACCCAGAACCTTCTTAGAGAGGAAGAAAT -----+-----+-----+-----+-----+-----+-----+-----	1844
1845	GCATGGGCAAAAGCAAATCATGCATATGGGCAGGAGTAACATTTTTTTGACTTTCATAGA -----+-----+-----+-----+-----+-----+-----+-----	1904
1905	AAGTACTGTATGGCACTAAACGGTCTAAACCGGACACTGGAAGCAAATCGTGCACGTGGG -----+-----+-----+-----+-----+-----+-----+-----	1964
1965	CAATATTATCTACCGTCGCGTCGCCAGTCTCCCATGCCCATGACCATGCTTGAATTTT -----+-----+-----+-----+-----+-----+-----+-----	2024
2025	AGTCTCGCCGGAGCTGCCGAGTGCATGCATAGTGACGAGTTTCAATAGGCCACTATATAT -----+-----+-----+-----+-----+-----+-----+-----	2084
2085	GTGATCATGGCTCTTGATTTGTCACTTTCTTTTTTTGCCGAAGGATATAGTAGTATTACT -----+-----+-----+-----+-----+-----+-----+-----	2144
2145	TTCTCTGCTATCACAAAGAAAGAACTGATTGTGTCTAGTCTAGGTGGTCTCAGAATTCTG -----+-----+-----+-----+-----+-----+-----+-----	2204
2205	CATGACTCCAGAGTATTCTTGATGCCACTTGTTTGTATTGCAAGAACTTAATTCGGAG -----+-----+-----+-----+-----+-----+-----+-----	2264
2265	ACAACCAAAAGCTCATCCCATGTCTCTGGAAC TAGTAGACATAAGAAAATCTCATGGTAT -----+-----+-----+-----+-----+-----+-----+-----	2324
2325	CAGTTTGCCTATTTATCTACAACGAAACGGCATGTTTGGTTTATTAAATTCAGAACTGG -----+-----+-----+-----+-----+-----+-----+-----	2384
		N W
2385	TACTGCTACGGGAAGGCGGTGCGGAGCAGGCGGCATCGGAGTTGGCGCGGCAGCGCGGC -----+-----+-----+-----+-----+-----+-----+-----	2444
	Y C Y G K A V A E Q A A S E L A R Q R G	

FIGURE 39 CONTINUED

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GTGGACCTTGTGGTGGTGAACCCGGTGCTGGTGATCGGCCCCCTGCTGCAGCCGACGGTG
2445 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2504
V D L V V V N P V L V I G P L L Q P T V

AACGCCAGCATCGGCCACATCCTCAAGTACCTGGACGGGTTCGGCCAGCAAGTTCGCCAAC
2505 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2564
N A S I G H I L K Y L D G S A S K F A N

GCCGTGCAGGCGTACGTGGACGTCCGCGACGTGGCCGACGCCACCTCCGCGTCTTCGAG
2565 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2624
A V Q A Y V D V R D V A D A H L R V F E

TGCGCCGCGCGTCCGGCCGCCACCTCTGCGCCGAGCGCGTCTCCACGCGAGGACGTC
2625 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2684
C A A A S G R H L C A E R V L H R E D V

GTGCGCATCTCGCCAAGCTCTTCCCCGAGTACCCCGTCCCCACCAGGTACGCGTACGAC
2685 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2744
V R I L A K L F P E Y P V P T R

CTGCTTGCTAGCCGCTTCCGTTAATTCCATTGCCTTAATTGATTGCATGATGCCGCTCCT
2745 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2804

AATTTACTCACTTGCGTAACCTAATTGCATTATATATGATCTACCAACCGTGGAGAAAAT
2805 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2864

TAGCAAGAGTCTGTGCGGGCGTCCCGGTCCAGTGCAGTTAACCTGCATGTCGATGGTCTG
2865 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2924

CAGGTTGCAGCTTACTTGTGGTTCTTTAGTTTCAGAGACACAGAGCAATTGGGCACTAAGC
2925 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 2984

AAAAGTGCATCACTGGTAATTAGGTAGCTCCACACACTGAAGTGGGTGGATCCCATCG
2985 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3044

GTAGTAGGTAAGGGTGGATAGTACTGGACGAGAGCTCGATCGTTGTTGTAAAAAAGCGAG
3045 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3104

TGACCACCACTTCACCATCCACTGCAAGTAGCTGCTAGTGAACCATCCAACCAGCTCCCT
3105 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3164

GGATCACTCTGCTCCGTCCGTACCTTCAGTTACCTACAGAAGCGACATGAACACACAGAC
3165 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3224

ACACAAGGCCGGCTCACCATTTCGCATAGGTCAAACCAAATGTTGGTGAACGGCAACATCG
3225 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3284

CCACAAGTCGCGTGCTAGTTTCGAGGTTGTGTCCGGTGTACCGAGGCCACACTATTTCGTGC
3285 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3344

TGCCCCGTCGCTGATATTTGCACGCGTAGCTGTGACGAAAGTAGGTGGACTGACAGATAC
3345 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3404

ACATATCCTCATTCGCTTCTCTGCTCGGTTTCTGCTAGGATTGCCATCTTCAGGAGTGCC
3405 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3464

TATCCGCACGGCAGAAACGCGTAGCATCAGGCCAGAAAGCAGCGTGCGTGATATCGTAAC
3465 -----+-----+-----+-----+-----+-----+-----+-----+-----+----- 3524

FIGURE 39 CONTINUED

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3525 CCAGACGGTCTTCACCTGTCCATTCTGGGCTACCTGGCATACTACCTCGGTGCCGCTGTG 3584
-----+-----+-----+-----+-----+-----+-----+-----
3585 CCGCTGACCAATTCGTGCACGACCCTATAGCAAAACCTATGCATGTAACCTGCTTCAAG 3644
-----+-----+-----+-----+-----+-----+-----+-----
3645 ATCAGCAGTGACATGTGCAATATAAACCTCAAGTGTGCACTCTAGTGCGTACTGATAAAA 3704
-----+-----+-----+-----+-----+-----+-----+-----
3705 CCGTATAACTGGTGACCCAGTCATTCTTCTCTTTTTTATTTGTTTGGACCAAACGAACAC 3764
-----+-----+-----+-----+-----+-----+-----+-----
3765 AGCATGTTATCCATCACCAACAAGTGGCGCTGATTTTCAAACCTACACGGGATCATACT 3824
-----+-----+-----+-----+-----+-----+-----+-----
3825 GGAAACCAAAGCAGGAGAACATCTTCGAACCAAGAGATGTTTACTAAATTTGAAAGAAAA 3884
-----+-----+-----+-----+-----+-----+-----+-----
3885 TGTACTGACAAGTAATCTGTCTGAAGCAAGACACATACTACCTCGGTTTGAACGTGGGAC 3944
-----+-----+-----+-----+-----+-----+-----+-----
3945 ACCATGCCCCGTGCCATATTTGCTAGGCACCACTCTGCCGTCGATTGTATCCCAACGGAGG 4004
-----+-----+-----+-----+-----+-----+-----+-----
4005 GAGTATCGATTTGCGCAAAGTTCCTACATACATAGCCGCTCAAGATATAATCTTACGACC 4064
-----+-----+-----+-----+-----+-----+-----+-----
4065 TTCCGTCGAAATCGGTGATACGTCGCAACCTATAGCTAACTTGGCAGAGCATAAAATAAC 4124
-----+-----+-----+-----+-----+-----+-----+-----
4125 TATCTAAGGTTGGGGTCTCCCTCTTTTCAATCAACCTTTCATACCGAATGATGGGAGTGT 4184
-----+-----+-----+-----+-----+-----+-----+-----
4185 TTGTGAAAACATCTCTTGGTCGACTCAGCATTAGCGCCCTACCAATTTCTCTGTGGACAA 4244
-----+-----+-----+-----+-----+-----+-----+-----
4245 TGCCACCTTAAATCGTTTTTTAGTCTTCATGATTTACTCCCCCTTATATCTGGCCGTAGT 4304
-----+-----+-----+-----+-----+-----+-----+-----
4305 CCCTCTTTTCCATTTTCTTGTCTGGTTTAAAGTCAAATTTAGACTACTAAAACAACAGC 4364
-----+-----+-----+-----+-----+-----+-----+-----
4365 AAGATTTTATGGAAGGGAGGTAGTGCAAAACAGAAAGTCCGATCGAAATGCGTGCCAATT 4424
-----+-----+-----+-----+-----+-----+-----+-----
4425 TGTGTCGCGCGGCCGGGACTAAAATGGATCTGCATGTGCATACCGTTGTCGGAGTATC 4484
-----+-----+-----+-----+-----+-----+-----+-----
4485 CTGCGAACGGTCGTGTGTTTAGTCAACATTAATGTGAGGTTTCATGTGATACTCTTGCTTG 4544
-----+-----+-----+-----+-----+-----+-----+-----
4545 AAAGATACTACTACTGCTACCTCGTAGAACTGAATGAAAGTATGTGGGACTGTTACGCTC 4604
-----+-----+-----+-----+-----+-----+-----+-----
4605 TCTGCACATGTCAAATGTCGTTACTCATACCTTTCGTCAGAGCATCCTGCGACGCGCGCC 4664
-----+-----+-----+-----+-----+-----+-----+-----
4665 GGTGCCGAAATTCGCCGTGTGTTTAGTCAAGATCAACGTGAGGTTTCATGCGGTACCCCTA 4724
-----+-----+-----+-----+-----+-----+-----+-----

FIGURE 39 CONTINUED

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TCTGGCTTCGAAGATACCAAGCAGACTGCGGCTAGATTGTCATTTTGATGTCGCAATCTT
4725 -----+-----+-----+-----+-----+-----+-----+----- 4784
CACCAAACCTGCCCTTCCGGACCACAGCAGCAGTACGTAAACAATGGTGTTCATCGCCATGC
4785 -----+-----+-----+-----+-----+-----+-----+----- 4844
GTTGCTCGTGTCCAAGGAAACGGAGGAATCTCGGCTTCCCACAAGTCACGCATCGATGTT
4845 -----+-----+-----+-----+-----+-----+-----+----- 4904
CACACCTGAATTGGTCGACGTTTCTTCTTCTAGACTAGAAAAAGATTACAGAACAACGCA
4905 -----+-----+-----+-----+-----+-----+-----+----- 4964
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4965 -----+-----+-----+-----+-----+-----+-----+----- 5024
ATGTCTATTCTGAATTTTTCGACTTCTATTCAAAGGATGGGCTGGAATTGCTACTGACTT
5025 -----+-----+-----+-----+-----+-----+-----+----- 5084
TGGTGTGATGTGTGTGGCACAGGTGCTCTGATGAGACGAACCCGAGGAAGCAGCCATACA
5085 -----+-----+-----+-----+-----+-----+-----+----- 5144
                                C S D E T N P R K Q P Y K
AGATGTGCAACCAGAAGCTCCAGGACCTCGGACTCGAGTTCAGGCCGGTGAGCCAGTCCC
5145 -----+-----+-----+-----+-----+-----+-----+----- 5204
      M S N Q K L Q D L G L E F R P V S Q S L
TGTACGAGACGGTGAAGAGCCTCCAGGAGAAGGGCCACCTTCCGGTGCTCAGCGAGCAGG
5205 -----+-----+-----+-----+-----+-----+-----+----- 5264
      Y E T V K S L Q E K G H L P V L S E Q A
CAGAGGCGGACAAGGAAACCCCTAGCTGCCGAGCTGCAGGCAGGGGTTACCATCCGAGCAT
5265 -----+-----+-----+-----+-----+-----+-----+----- 5324
      E A D K E T L A A E L Q A G V T I R A *
GAGGAACAAGAAATCAACCATGTCCATACTGCTACTGTCATGTAAACCAGCTGTTGAATG
5325 -----+-----+-----+-----+-----+-----+-----+----- 5384
CCTAAAATCTAAGTTCTTGTAATACTGTGTTGTTTCATGTGGACTAGATTGATCG
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FIGURE 39 CONTINUED

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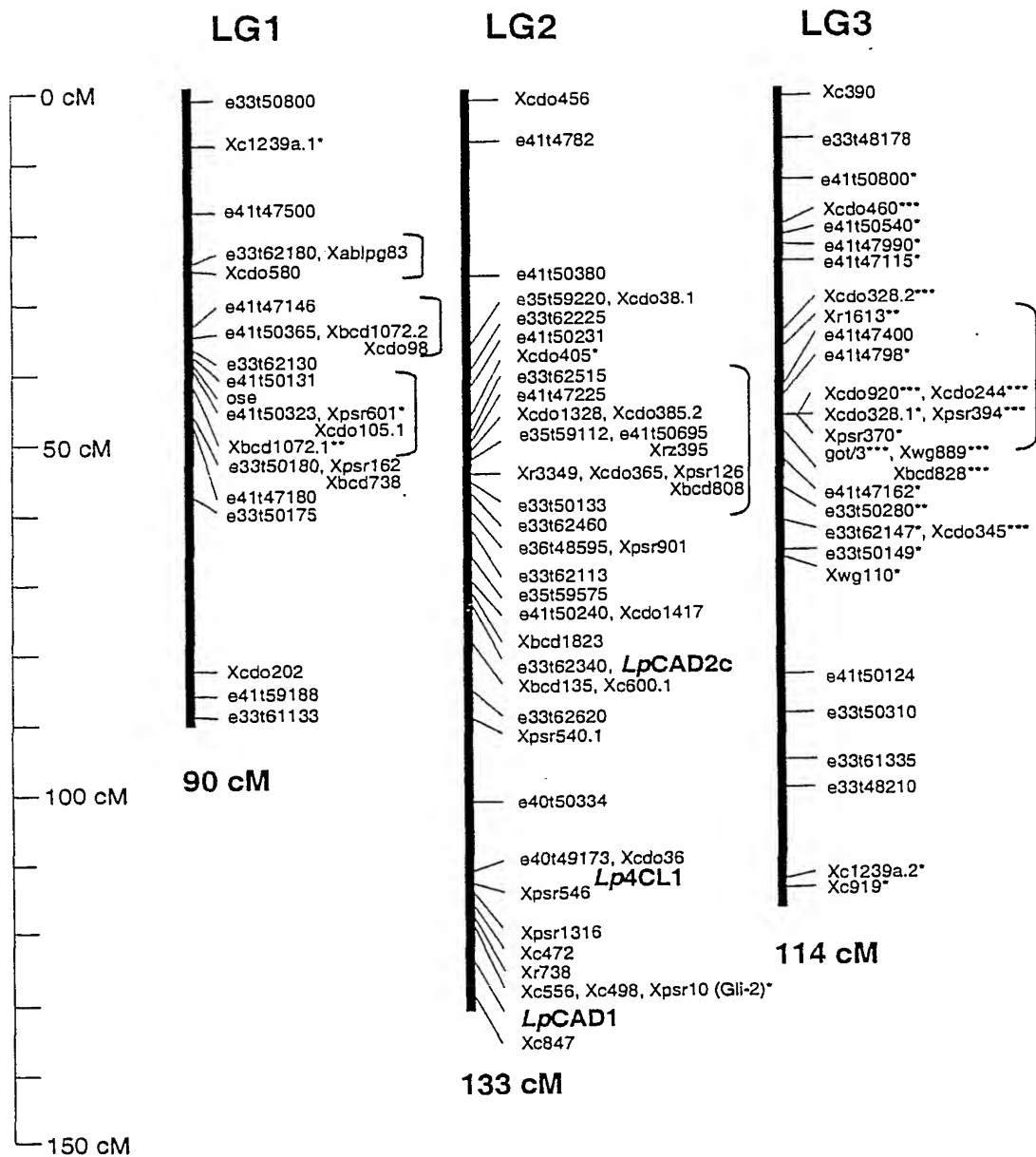


FIGURE 40

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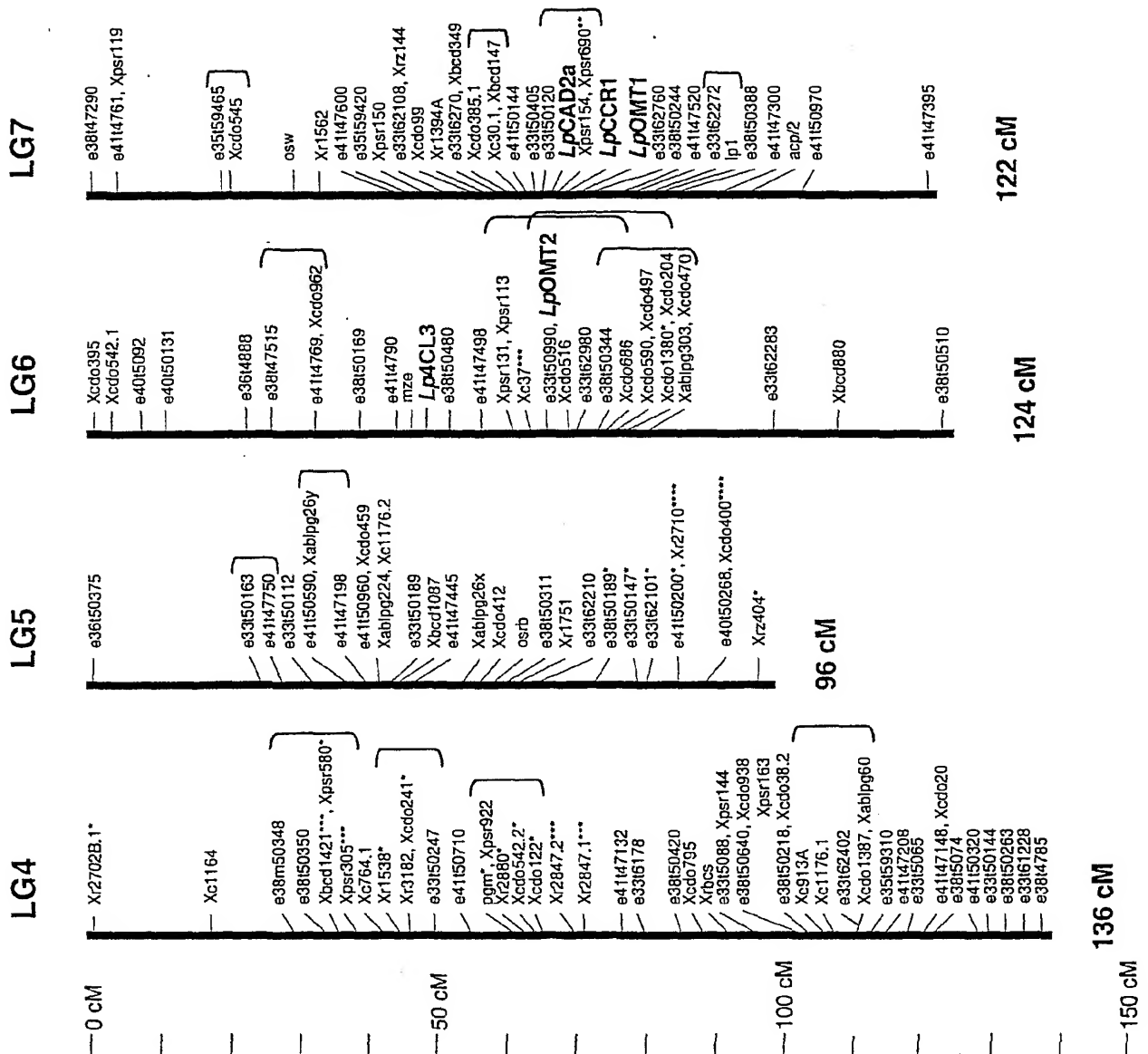


FIGURE 40 CONTINUED

Organization Applicant

Street : 15th Floor, 8 Nicholson Street
City : East Melbourne
State : Victoria
Country : Australia
PostalCode : 3002
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : State of Victoria as represented by Department of Natural Resources and Environment

Organization Applicant

Street : North Terrace
City : Adelaide
State : South Australia
Country : Australia
PostalCode : 5005
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : The University of Adelaide

Organization Applicant

Street : Lisboa 27, Apartado Postal 6-641
City : Mexcio
State : DF
Country : Mexico
PostalCode : 06600
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : International Maize and Wheat Improvement Center

Organization Applicant

Street : Waite Road
City : Glen Osmond
State : South Australia
Country : Australia
PostalCode : 5064
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : State of South Australia as represented by South Australian Research and Development Institute

Organization Applicant

Street : Military Road
City : Lismore
State : New South Wales
Country : Australia
PostalCode : 2580
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : Southern Cross University

Organization Applicant

Street : Level 3, 84 William Street
City : Melbourne
State : Victoria
Country : Australia

PostalCode : 3000
 PhoneNumber :
 FaxNumber :
 EmailAddress :

<110> OrganizationName : Dairy Research and Development Corporation

Application Project

<120> Title : Modification of Lignin Biosynthesis
 <130> AppFileReference : 40494788
 <140> CurrentAppNumber : AU PQ8154
 <141> CurrentFilingDate : 2001-06-14

Earlier Applications

<150> PriorAppNumber : AU PQ8154
 <151> PriorFilingDate : 2000-06-14

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<213> OrganismName : Lolium perenne
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<212> Type : DNA

<211> Length : 2284

SequenceName : 4CL1cDNA (SEQ ID NO: 1)

SequenceDescription :

Custom Codon

Sequence Name : 4CL1cDNA (SEQ ID NO: 1)

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 MITVAAPEVQ QPQIAAAAAA VEAAPEATT IFRSRLPDID IPTHMPLHDY CFATAASAPD 60APCLITAAT
 G KTYTFAETHL LCRKAAAALH GLGVRHGDRI MLLLQNSVEF ALAFFGASML 120GAVSTAANPF CTPQEIH
 KQL VASGAKLVVT QSAYVDKLRH EAFPRIGEAL TVITIDEDDG 180TPDGCQPFWA LVSAADENSV PESPI
 SPDDA VALPYSSGTT GLPKGVVLTH GGLVSSVAQQ 240VDGENPNLHM RAGEDVVLVCV LPLFHIFSLN SVL
 LCLARAG AAVMLMPRFE MGAMLEGIER 300WRVTVAAVVP PLVLALAKNP GVEKHDLSI RIVLSGAAPL G
 KELEDALRG RLPQAIFGQG 360YGMTEAGPVL SMCPAFAREP TPAKSGSCGT VVRNAQLKVV DPDTGVS LGR
 NLPGEICIRG 420PQIMKGYLND PVATAATIDV EGWLHTGDIG YVDDDDEVFI VDRVKELIKF KGFQVPPA
 EL 480EALLIAHPSI ADAAVVPQKD DAAGEVPVAF VVRAADSDIA EEAIKEFVSK QVVFYKRLHK 54
 OVYFTHAIPKS ASGKILRKEL RAKLAAPATA RVVHGFMLII SIRKALLAYM FHLLFHLEDC 600IPASGQ
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<212> Type : PRT

<211> Length : 606

SequenceName : 4CL1pep (SEQ ID NO: 2)

SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
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 c atcgcggcgg acgcgcctcc cgcggagctg gtgttccggg ccaagctccc 120ggacatcgag atcccg
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 1992

<212> Type : DNA

<211> Length : 1992

SequenceName : 4CL2cDNA (SEQ ID NO: 3)

SequenceDescription :

Custom Codon

Sequence Name : 4CL2cDNA (SEQ ID NO: 3)

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
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A GLRRLGVGKG DVVMALLRNC PEFAFVFLGA ARLGAATTTA NPFYTPHEIH 120RQATAAGARV IVTEACA
 VEK VRAFAAERGI PVVSDEGVD GGCLPFAETL LGEESGERFV 180DEAVDPDDVV ALPYSSGTTG LPKG
 MLTHR SLVTSVAQOV DGENPNLHFS SSDVLLCVLP 240LFHIYSLNSV LLAGLRAGCA IVIMRKFDHG ALV
 DLVRTHG VTVAPFVPPI VVEIAKSARV 300TAADLASIRL VMSGAPMGK ELQDAFMAKI PNAVLGQGYG M
 TEAGPVLAM CLAFAKEPFA 360VKSGSCGTVV RNAELKIVDP DTGASLGRNL PGEICIRGKQ IMKGYLNDPV
 ATKNTIDKDG 420WLHTGDIGYV DDDDEIFIVD RLKEIIKYKG FQVPPAELEA LLITHPEIKD AAVVSMQD
 EL 480AGEVPVAFVV RTEGSEISEN EIKQFVAKEV VFYKRICKVF FADSIPKSPS GKILRKDLRA 54
 OKLAAGIPSSN TTQSKS 556

<212> Type : PRT

<211> Length : 556

SequenceName : 4CLpep (SEQ ID NO: 4)

SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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 g cgggtgaggt agctagctag ctactcgtac tagaccatta ccatgggttc 120cgtgccggag gagtcag
 tgg tggcgggtggc accggcggag acggtgttcc ggctgaagct 180ccccgacatc gagatcaaca acgag
 cagac gctgcagagc tactgcttcg agaagatggc 240cgaggtcgcg tcccgccct gcacatcga cgg
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 gccgcatgg gcgtggggaa 360gggcgacgtg gtgatgaacc tgctgcgcaa ctgcccgag ttgccttct
 cttctctggg 420cgccggcgcg ctgggcgcgc ccaccaccac cgccaaccgc ttctacacct cgcacgag
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 0gaaggtgctg gagttcgcg cggggcgggg cgtgcccgtg gtcaccgtcg acgggagggc 600cgacgggt
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 gacg acgtcgtgc cctgccctac tctccggca ccaccgggct 720ccccaagggc gtcagtctca ccca
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 tccacatc tactcgtgc acacggtgct gctggcgggg ctccgcgtcg gcgcgcccat 900cgtcatcatg
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<212> Type : DNA

<211> Length : 2038

SequenceName : 4CLcDNA (SEQ ID NO: 5)

SequenceDescription :

Custom Codon

Sequence Name : 4CLcDNA (SEQ ID NO: 5)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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 R RAAAGLRRMG VGKGDVVMNL LRNCPEFAFS FLGAARLGAA TTTANPFYTP 120HEIHRQAEAA GAKLIVT
 EAC AVEKVFLEFAA GRGVPVVTVD GRRDGCVDFA ELIAGEELPE 180ADEAGVLPDD VVALPYSSGT TGLPK
 GVMLT HRSLVTSVAQ LVDGSPNVC FNKDDALLCL 240LPLFHIYSLH TVLLAGLRVG AAIVIMRKFD VGA
 LVDLVRA HRITIAFPVP PIVVEIAKSD 300RVGADDLASI RMVLSGAAPM GKDLQDAFMA KIPNAVLGQG Y
 GMTEAGPVL AMCLAFAKEP 360FKVKSGSCGT VVRNAELKVV DPDTGASLGR NQPGEICVRG KQIMIGYLND

PESTKNTIDK 420DGWLHTGDIG LVDDDDDEIFI VDRLKEIIKY KGFQVAPAEI EALLLTNPEV KDAAVVGV
 KD 480DLCGEVPVAF IKRIEGSEIN ENEIKQFVSK EVVIFYKRINK VYFTDSIPKN PSGKILRKDL 54
 ORARLAAGIPT EVAAPRS 557

<212> Type : PRT

<211> Length : 557

SequenceName : 4CLpep (SEQ ID NO: 6)

SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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 g cctgtcttcg tttccgcctc accgccggcc ggttctccgc tccaagctac 120gtccgtccgt ccacata
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 aaaaaaaaa aaaa 1395

<212> Type : DNA

<211> Length : 1395

SequenceName : CCR1cDNA (SEQ ID NO: 7)

SequenceDescription :

Custom Codon

Sequence Name : CCR1cDNA (SEQ ID NO: 7)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

MTIAEVVAAG DTAAAVVQPA GNGQTVCVTG AAGYIASWLV KILLEKGYTV KGTVRNPDDP 60KNAHLRALD
 G AADRLVLCKA DLLDYDAIRR AIDGCHGVFH TASFVTDDPE QMVEPAVRGT 120QYVIDAAAEA GTVRRMV
 LTS SIGAVTMDPN RGPDVVVDES CWSLDLDFCK TRNWYCYGKA 180VAEQAASELA RQGVLDLVVV NPVLV
 IGPLL OPTVNASIGH ILKYLDGSAS KFANAVQAYV 240DVRDVADAH LRVFECAAAASG RHLCAERVLH RED
 VVRILAK LFPEYVPVTR CSDETNPRKQ 300PYKMSNQKLQ DLGLEFRPVS QSLYETVKSL QEKGHLPLVS E
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<212> Type : PRT

<211> Length : 362

SequenceName : CCR1pep (SEQ ID NO: 8)

SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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<212> Type : DNA

<211> Length : 1325

SequenceName : CAD1cDNA (SEQ ID NO: 9)

SequenceDescription :

Custom Codon

Sequence Name : CAD1cDNA (SEQ ID NO: 9)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

MAEGLPALGW AARDASGHL S YSFSRSVPK DDDVTIKVLF CGICHTDLHI IKNDWGNALY 60PIVPGHEIV
 G VVASVSGSV SFKAGDTGV GYFLDSCRTC YSCSKGYENF CPTLTLSNG 120VDGGGATTQG GFSDELV
 VNK DYVIRVPDNL PLAGAAPLLC AGVTYSPMV EYGLNAPGKH 180XGVVGLGGLG HVAVKFGKAF GMTVT
 VISSS DRKRDEALGR LGADAFVSS DPEQMKAAG 240TMDGIIDTVS AGHPIVPLLD LLKPMGQMVV VGA
 PSKPLEL PAFAIIGGK RLAGSGTGSV 300AHCQAMLDFA GKHGITADVE VVKMDYGOHR HRAAREERRQ V
 PLRHRRRRQ PPAGHRRRLTC 360ATQCGRALVW SRKRFAGSQP HEQVNESLVC CLSSFHIWDA VPDFHVK
 407

<212> Type : PRT

<211> Length : 407

SequenceName : CAD1pep (SEQ ID NO: 10)

SequenceDescription :

Sequence

<213> OrganismName : Lolium preenne

<400> PreSequenceString :

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<212> Type : DNA

<211> Length : 1377

SequenceName : CAD2cvEllettcDNA (SEQ ID NO: 11)
SequenceDescription :

Custom Codon

Sequence Name : CAD2cvEllettcDNA (SEQ ID NO: 11)

Sequence

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
MAPTAAEQTE HHQHTRKAVG LAARDDAGHL SPLAITRRST GDDDVVILKIL YCGICHSDLH 60ALKNDWKNS
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VTY GGYSSMVVVH ERFVVRFPDA MPLDKGAPLL CAGITVYSPM 180KYHGLNVPGL HLGVLGLGGL GHVAV
KFGKA FGMKVTVISS SPGKKEEALG RLGADAFIVS 240KDADEMKAVI APWMASXNTV SANIPLPLF GLL
KPNGKMI MVGLPEKPIE IPPFALVATN 300KTLAGSIIGG MSDTQEMLDL AAKHGVTDI EVVGAEYVNT A
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370
<212> Type : PRT
<211> Length : 370
SequenceName : CADcvEllettep (SEQ ID NO: 12)
SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
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5119

<212> Type : DNA

<211> Length : 5119

SequenceName : LpOmt1promoter (SEQ ID NO: 13)

SequenceDescription :

Custom Codon

Sequence Name : LpOmt1promoter (SEQ ID NO: 13)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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<212> Type : DNA

<211> Length : 4555

SequenceName : CAD2cvBarlanogenomic (SEQ ID NO: 14)

SequenceDescription :

Custom Codon

Sequence Name : CAD2cvBarlanogenomic (SEQ ID NO: 14)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

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<212> Type : PRT

<211> Length : 312

SequenceName : CAD2cvBarlanopep (SEQ ID NO: 15)

SequenceDescription :

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<213> OrganismName : Lolium perenne

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<212> Type : DNA

<211> Length : 1378

SequenceName : CAD2cvBarlanocDNA (SEQ ID NO: 16)

SequenceDescription :

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Sequence Name : CAD2cvBarlanocDNA (SEQ ID NO: 16)

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<213> OrganismName : Lolium perenne

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<212> Type : DNA

<211> Length : 2650

SequenceName : 4CL2promoterseq (SEQ ID NO: 17)

SequenceDescription :

Custom Codon

Sequence Name : 4CL2promoterseq (SEQ ID NO: 17)

Sequence

<213> OrganismName : Lolium perenne

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/00699

A. CLASSIFICATION OF SUBJECT MATTERInt. Cl. ⁷: A01H 5/00, C12N 9/00, C12N 9/02, C12N 9/04, C12N 9/10, C12N 15/29

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

GenBank, EMBL, PDB Nucleic Acids, GenPept, TREMBL, SWISS-PROT, PIR, Medline, ChemAbs, AGRICOLA, WPIDS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 952 486 A (BLOKSBERG <i>et al</i>) 14 September 1999 whole of document	1-26
X	Civardi L <i>et al</i> , "Molecular Cloning and Characterization of two cDNAs Encoding Enzymes Required for Secondary Cell Wall Biosynthesis in Maize", <i>NATO ASI Series, Volume H 104 (Cellular Integration of Signalling Pathways in Plant Development)</i> , 1998, pages 135-146 whole of document	1, 2, 4-12, 14-17, 20a, 19b, 23-26
X	WO 99/31243 A (INTERNATIONAL PAPER COMPANY) 24 June 1999 whole of document	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26

☒ Further documents are listed in the continuation of Box C
 ☒ See patent family annex

* Special categories of cited documents:		"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance		"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent but published on or after the international filing date		"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)		"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

23 July 2001

Date of mailing of the international search report

29 August 2001

Name and mailing address of the ISA/AU

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GARETH COOK

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/00699

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GenBank accession AF052223, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL3 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37734, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL3 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052222, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL2 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37733, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL2 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052221, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL1 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37732, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL1 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	WO 98/39454 A (ZENECA LIMITED) 11 September 1998 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	Pichon M <i>et al</i> , "Cloning and characterization of two maize cDNAs encoding Cinnamoyl-CoA Reductase (CCR) and differential expression of the corresponding genes", <i>Plant Molecular Biology</i> , 1998, 38:671-676 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	GenBank accession AJ231134, Selman-Housein G <i>et al</i> , "Saccharum officinarum mRNA for cinnamoyl-CoA reductase" 25 January 2000	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	WO 93/05159 A (IMPERIAL CHEMICAL INDUSTRIES PLC) 18 March 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	WO 93/24638 A (ZENECA LIMITED) 9 December 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	Baucher M <i>et al</i> , "Down-regulation of cinnamyl alcohol dehydrogenase in transgenic alfalfa (<i>Medicago sativa</i> L.) and the effect on lignin composition and digestibility", <i>Plant Molecular Biology</i> , 1999, 39:437-447 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	GenBank accession AF010290, McAlister FM <i>et al</i> , "Lolium perenne cinnamyl alcohol dehydrogenase mRNA, complete cds", 23 September 1997	1, 2, 5-12, 15-17, 19b, 23-26
X	GenPept accession AAB70908, McAlister FM, "cinnamyl alcohol dehydrogenase [Lolium perenne]", 22 September 1997	1, 2, 5-12, 15-17, 19b, 23-26

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU01/00699

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Heath R <i>et al</i> , "cDNA Cloning and Differential Expression of Three Caffeic Acid O-Methyltransferase Homologues from Perennial Ryegrass (<i>Lolium perenne</i>)," <i>Journal of Plant Physiology</i> , 1998, 153:649-657 whole of document	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033540, Heath RL et al, "Lolium perenne caffeic acid O-methyltransferase (OMT3) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033539, Heath RL et al, "Lolium perenne caffeic acid O-methyltransferase (OMT2) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033548, Heath RL et al, "Lolium perenne caffeic acid O-methyltransferase (OMT1) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF010291, McAlister FM <i>et al</i> , "Lolium perenne bispecific caffeic acid/hydroxyferulic acid O-methyltransferase mRNA, complete cds", 3 June 1998	17, 18, 19b, 20b, 23-26
X	Capellades M <i>et al</i> , "The maize caffeic acid O-methyltransferase gene promoter is active in transgenic tobacco and maize plants," <i>Plant Molecular Biology</i> , 1996, 31:307-322 whole of document	17, 18, 19b, 20b, 23-26
Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.		

Supplemental Box

(To be used when the space in any of Boxes I to VIII is not sufficient)

Continuation of Box No: II, Observations where unity of invention is lacking

The international search report has been drawn up in respect of the entire international application but the International Searching Authority is of the opinion that the application does not appear to comply with the requirements of unity of invention as set forth in the PCT regulations (Article 34(3), Rule 68(1) PCT).

The separate groups of invention are:

1. Claims 1, 2, 6 to 12, 16, 17, 19a, 19b and 23 to 26 (partial) and claims 3, 13 and 21 (complete) are to 4-coumarate-CoA ligase (4CL) from ryegrass (*Lolium*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from ryegrass is considered to be the first "special technical feature".
2. Claims 1, 6 to 11, 16, 17, 19a, 19b and 23 to 26 (partial) are to 4CL from fescue (*Festuca*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from fescue is considered to be the second "special technical feature".
3. Claims 1, 2, 6 to 12, 16, 17, 20a, 19b and 23 to 26 (partial) and claims 4, 14 and 22 (complete) are to cinnamoyl-CoA reductase (CCR) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from ryegrass is considered to be the third "special technical feature".
4. Claims 1, 6 to 11, 16, 17, 19b, 20a and 23 to 26 (partial) are to CCR from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from fescue is considered to be the fourth "special technical feature".
5. Claims 1, 2, 6 to 12, 16, 17, 19b and 23 to 26 (partial) and claims 5 and 15 (complete) are to cinnamyl alcohol dehydrogenase (CAD) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from ryegrass is considered to be the fifth "special technical feature".
6. Claims 1, 6 to 11, 16, 17, 19b, and 23 to 26 (partial) are to CAD from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from fescue is considered to be the sixth "special technical feature".
7. Claims 17, 18, 19b and 23 to 26 (partial) and claim 20b (complete) are to caffeic acid O-methyltransferase (OMT) gene promoter from ryegrass and various uses of it. Caffeic acid OMT from ryegrass is considered to be the seventh "special technical feature".
8. Claims 17, 18, 19b and 23 to 26 (partial) are to caffeic acid O-methyltransferase (OMT) gene promoter from fescue and various uses of it. Caffeic acid OMT from fescue is considered to be the eighth "special technical feature".

In order for there to be unity between the four types of enzymes claimed, they have to share a significant structural element, that is a structural element that defines the specific biological activity of the enzymes, and the significant structural element must be disclosed in the specification. No significant structural element has been identified as being shared by the four types of enzymes, hence there is lack of unity between the enzymes. In addition, all four types of enzymes are known in the prior art, for example, in US 5 952 486. Hence unity of invention is also lacking between the enzymes from ryegrass and the enzymes from fescue.

Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.
PCT/AU01/00699

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member	
US 5 952 486	AU 44036/97	BR 9711745	EP 929 682
	US 5 850 020	WO 98/11205	ZA 9710451
	ZA 9810574	US 6 204 434	
WO 99/31243	US 6 252 135	ZA 9811568	
WO 98/39454	AU 63041/98	EP 970 222	
WO 93/05159	AU 16581/92	BR 9205934	CA 2 109 222
	EP 584 117	US 5 451 514	US 6 066 780
WO 93/24638	AU 43345/93	BR 9306445	EP 643 774
	US 5 633 439		
END OF ANNEX			